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(54) Title: NOVEL PLANT ACYLTRANSFERASES (57) Abstract <p>By this invention, novel nucleic acid sequences encoding for acyltransferase related proteins are provided, wherein said acyltransferase-like protein is active in the transfer of a fatty acyl group from a fatty acyl donor to a fatty acyl acceptor. Also considered are amino acid and nucleic acid sequences obtainable from AT-like nucleic acid sequences and the use of such sequences to provide transgenic host cells capable of producing modified lipid content and composition.</p>		

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NOVEL PLANT ACYLTRANSFERASES

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INTRODUCTION

This application claims the benefit of U.S. Provisional Application Serial No. 60/101,939 filed September 25, 1998.

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Technical Field

The present invention is directed to nucleic acid and amino acid sequences and constructs, and methods related thereto.

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Background

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Through the development of plant genetic engineering techniques, it is now possible to produce transgenic varieties of plant species to provide plants which have novel and desirable characteristics. For example, it is now possible to genetically engineer plants for tolerance to environmental stresses, such as resistance to pathogens and tolerance to herbicides and to improve the quality characteristics of the plant, for example improved fatty acid compositions. However, the number of useful nucleotide sequences for the engineering of such characteristics is thus far limited and the speed with which new useful nucleotide sequences for engineering new characteristics is slow.

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The characterization of various acyltransferase proteins is useful for the further study of plant fatty acid synthesis systems and for the development of novel and/or alternative oils sources. Studies of plant mechanisms may provide means to further enhance, control, modify, or otherwise alter the total fatty acyl composition of triglycerides and oils.

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Furthermore, the elucidation of the factor(s) critical to the natural production of fatty acids in plants is desired, including the purification of such factors and the characterization of element(s) and/or cofactors which enhance the efficiency of the system. Of particular interest are the nucleic acid sequences of genes encoding proteins which may be useful for applications in genetic engineering.

SUMMARY OF THE INVENTION

5 The present invention provides nucleic acid encoding for amino acid sequences for a class of proteins which are related to acyltransferase proteins. Such proteins are referred to herein as acyltransferase related or acyltransferase like proteins.

 By this invention, nucleic acid sequences encoding these acyltransferase related proteins may now be characterized with respect to enzyme activity. In particular,
10 identification and isolation of nucleic acid sequences encoding for acyltransferase related proteins from *Arabidopsis*, yeast, corn, and soybean are provided.

 Thus, this invention encompasses acyltransferase related nucleic acid sequences and the corresponding amino acid sequences, and the use of these nucleic acid sequences in the preparation of oligonucleotides containing such acyltransferase related encoding sequences
15 for analysis and recovery of plant acyltransferase related gene sequences. The acyltransferase related encoding sequence may encode a complete or partial sequence depending upon the intended use. All or a portion of the genomic sequence, or cDNA sequence, is intended.

 Of special interest are recombinant DNA constructs which provide for transcription or transcription and translation (expression) of the acyltransferase related sequences in host
20 cells. In particular, constructs which are capable of transcription or transcription and translation in plant host cells are preferred. For some applications a reduction in sequences encoding acyltransferase related sequences may be desired. Thus, recombinant constructs may be designed having the acyltransferase related sequences in a reverse orientation for expression of an anti-sense sequence or use of co-suppression, also known as "transwitch",
25 constructs may be useful. Such constructs may contain a variety of regulatory regions including transcriptional initiation regions obtained from genes preferentially expressed in plant seed tissue. For some uses, it may be desired to use the transcriptional and translational initiation regions of the acyltransferase related gene either with the acyltransferase related encoding sequence or to direct the transcription and translation of a heterologous sequence.

30 Also considered in this invention are the plants and seeds containing the constructs and polynucleotides of this invention.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 provides the 204 amino acid conserved sequence profile identified from comparisons of glycerol-3-phosphate acyltransferase and various lysophosphatidic acid acyltransferase using PSI-BLAST.

Figure 2 provides an amino acid sequence alignment for the acyltransferase sequences. The alignment shown is of the regions of the protein extending from about 30 amino acids prior to the conserved H in the conserved sequence HXXXXD to 100 amino acids after, or downstream, of the P in the conserved PEG sequence motif of the acyltransferase-like sequences.

Figure 3 provides schematics showing the relationship of the identified acyltransferases. The relationships described are derived from an alignment of the regions of the protein extending from about 30 amino acids prior to the conserved H in the conserved sequence HXXXXD to 100 amino acids after, or downstream, of the P in the conserved PEG sequence motif of the acyltransferase-like sequences. Figure 3A provide a phylogenetic tree showing the relationship of several acyltransferases. Figure 3B provides a table showing the percent similarities and percent divergence of the novel acyltransferases and known acyltransferases using the Clustal method with PAM250 residue weight table.

DETAILED DESCRIPTION OF THE INVENTION

In accordance with the subject invention, nucleotide sequences are provided which are capable of coding sequences of amino acids, such as, a protein, polypeptide or peptide, which are related to nucleic acid sequences encoding acyltransferase proteins, referred to herein as acyltransferase-like or acyltransferase related. The novel nucleic acid sequences find use in the preparation of constructs to direct their expression in a host cell. Furthermore, the novel nucleic acid sequences may find use in the preparation of plant expression constructs to modify the fatty acid composition of a plant cell.

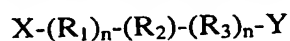
In one embodiment of the present invention, nucleic acid sequences, also referred to herein as polynucleotides, are identified from databases which are related to acyltransferases.

Isolated proteins, Polypeptides and Polynucleotides

A first aspect of the present invention relates to isolated acyltransferase polynucleotides. The polynucleotide sequences of the present invention include isolated polynucleotides that encode the polypeptides of the invention having a deduced amino acid sequence selected from the group of sequences set forth in the Sequence Listing and to other polynucleotide sequences closely related to such sequences and variants thereof.

The invention provides a polynucleotide sequence identical over its entire length to each coding sequence as set forth in the Sequence Listing. The invention also provides the coding sequence for the mature polypeptide or a fragment thereof, as well as the coding sequence for the mature polypeptide or a fragment thereof in a reading frame with other coding sequences, such as those encoding a leader or secretory sequence, a pre-, pro-, or prepro- protein sequence. The polynucleotide can also include non-coding sequences, including for example, but not limited to, non-coding 5' and 3' sequences, such as the transcribed, untranslated sequences, termination signals, ribosome binding sites, sequences that stabilize mRNA, introns, polyadenylation signals, and additional coding sequence that encodes additional amino acids. For example, a marker sequence can be included to facilitate the purification of the fused polypeptide. Polynucleotides of the present invention also include polynucleotides comprising a structural gene and the naturally associated sequences that control gene expression.

The invention also includes polynucleotides of the formula:



wherein, at the 5' end, X is hydrogen, and at the 3' end, Y is hydrogen or a metal, R_1 and R_3 are any nucleic acid residue, n is an integer between 1 and 3000, preferably between 1 and 1000 and R_2 is a nucleic acid sequence of the invention, particularly a nucleic acid sequence selected from the group set forth in the Sequence Listing and preferably SEQ IDNOs: 1, 3, 5, 7, 9, 10, 12, 14, 16, 18, 20, 22, and 226-233. In the formula, R_2 is oriented so that its 5' end residue is at the left, bound to R_1 , and its 3' end residue is at the right, bound to R_3 . Any stretch of nucleic acid residues denoted by either R group, where R is greater than 1, may be either a heteropolymer or a homopolymer, preferably a heteropolymer.

The invention also relates to variants of the polynucleotides described herein that encode for variants of the polypeptides of the invention. Variants that are fragments of the polynucleotides of the invention can be used to synthesize full-length polynucleotides of the

invention. Preferred embodiments are polynucleotides encoding polypeptide variants wherein 5 to 10, 1 to 5, 1 to 3, 2, 1 or no amino acid residues of a polypeptide sequence of the invention are substituted, added or deleted, in any combination. Particularly preferred are substitutions, additions, and deletions that are silent such that they do not alter the properties or activities of the polynucleotide or polypeptide.

Nucleotide sequences encoding acyltransferases may be obtained from natural sources or be partially or wholly artificially synthesized. They may directly correspond to an acyltransferase endogenous to a natural source or contain modified amino acid sequences, such as sequences which have been mutated, truncated, increased or the like. Acyltransferases may be obtained by a variety of methods, including but not limited to, partial or homogenous purification of protein extracts, protein modeling, nucleic acid probes, antibody preparations and sequence comparisons. Typically an acyltransferase will be derived in whole or in part from a natural source. A natural source includes, but is not limited to, prokaryotic and eukaryotic sources, including, bacteria, yeasts, plants, including algae, and the like.

Of special interest are acyltransferases which are obtainable from eukaryotic sources, including those which are obtained, from plants, or from acyltransferases which are obtainable through the use of these sequences. "Obtainable" refers to those acyltransferases which have sufficiently similar sequences to that of the sequences provided herein to provide a biologically active protein of the present invention.

Further preferred embodiments of the invention that are at least 50%, 60%, or 70% identical over their entire length to a polynucleotide encoding a polypeptide of the invention, and polynucleotides that are complementary to such polynucleotides. More preferable are polynucleotides that comprise a region that is at least 80% identical over its entire length to a polynucleotide encoding a polypeptide of the invention and polynucleotides that are complementary thereto. In this regard, polynucleotides at least 90% identical over their entire length are particularly preferred, those at least 95% identical are especially preferred. Further, those with at least 97% identity are highly preferred and those with at least 98% and 99% identity are particularly highly preferred, with those at least 99% being the most highly preferred.

Preferred embodiments are polynucleotides that encode polypeptides that retain substantially the same biological function or activity as the mature polypeptides encoded by the polynucleotides set forth in the Sequence Listing.

The invention further relates to polynucleotides that hybridize to the above-described sequences. In particular, the invention relates to polynucleotides that hybridize under stringent conditions to the above-described polynucleotides. As used herein, the terms "stringent conditions" and "stringent hybridization conditions" mean that hybridization will generally occur if there is at least 95% and preferably at least 97% identity between the sequences. An example of stringent hybridization conditions is overnight incubation at 42°C in a solution comprising 50% formamide, 5x SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5x Denhardt's solution, 10% dextran sulfate, and 20 micrograms/milliliter denatured, sheared salmon sperm DNA, followed by washing the hybridization support in 0.1x SSC at approximately 65°C. Other hybridization and wash conditions are well known and are exemplified in Sambrook, *et al.*, Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY (1989), particularly Chapter 11.

The invention also provides a polynucleotide consisting essentially of a polynucleotide sequence obtainable by screening an appropriate library containing the complete gene for a polynucleotide sequence set forth in the Sequence Listing under stringent hybridization conditions with a probe having the sequence of said polynucleotide sequence or a fragment thereof; and isolating said polynucleotide sequence. Fragments useful for obtaining such a polynucleotide include, for example, probes and primers as described herein.

As discussed herein regarding polynucleotide assays of the invention, for example, polynucleotides of the invention can be used as a hybridization probe for RNA, cDNA, or genomic DNA to isolate full length cDNAs or genomic clones encoding a polypeptide and to isolate cDNA or genomic clones of other genes that have a high sequence similarity to a polynucleotide set forth in the Sequence Listing. Such probes will generally comprise at least 15 bases. Preferably such probes will have at least 30 bases and can have at least 50 bases.

Particularly preferred probes will have between 30 bases and 50 bases, inclusive.

The coding region of each gene that comprises or is comprised by a polynucleotide sequence set forth in the Sequence Listing may be isolated by screening using a DNA sequence provided in the Sequence Listing to synthesize an oligonucleotide probe. A labeled oligonucleotide having a sequence complementary to that of a gene of the invention is then used to screen a library of cDNA, genomic DNA or mRNA to identify members of the library which hybridize to the probe. For example, synthetic oligonucleotides are prepared which correspond to the N-terminal sequence of the polypeptide. The partial sequences so prepared can then be used as probes to obtain acyltransferase clones from a gene library prepared from

a cell source of interest. Alternatively, where oligonucleotides of low degeneracy can be prepared from particular peptides, such probes may be used directly to screen gene libraries for gene sequences. In particular, screening of cDNA libraries in phage vectors is useful in such methods due to lower levels of background hybridization.

5 Typically, a sequence obtainable from the use of nucleic acid probes will show 60-70% sequence identity between the target acyltransferase sequence and the encoding sequence used as a probe. However, lengthy sequences with as little as 50-60% sequence identity may also be obtained. The nucleic acid probes may be a lengthy fragment of the nucleic acid sequence, or may also be a shorter, oligonucleotide probe. When longer nucleic acid
10 fragments are employed as probes (greater than about 100 bp), one may screen at lower stringencies in order to obtain sequences from the target sample which have 20-50% deviation (i.e., 50-80% sequence homology) from the sequences used as probe.

Oligonucleotide probes can be considerably shorter than the entire nucleic acid sequence encoding an acyltransferase enzyme, but should be at least about 10, preferably at least about
15 15, and more preferably at least about 20 nucleotides. A higher degree of sequence identity is desired when shorter regions are used as opposed to longer regions. It may thus be desirable to identify regions of highly conserved amino acid sequence to design oligonucleotide probes for detecting and recovering other related genes. Shorter probes are often particularly useful for polymerase chain reactions (PCR), especially when highly conserved sequences can be
20 identified. (*See, Gould, et al., PNAS USA (1989) 86:1934-1938*).

The skilled artisan will appreciate that, in many cases, an isolated cDNA sequence will be incomplete, in that the region coding for the polypeptide is truncated with respect to the 5' terminus of the cDNA. This is a consequence of the reverse transcriptase, an enzyme with low 'processivity' (a measure of the ability of the enzyme to remain attached to the
25 template during the polymerization reaction) employed during the first strand cDNA synthesis.

There are several methods available and are well know to the skilled artisan to obtain full-length cDNAs, or extend short cDNAs, for example those based on the method of Rapid Amplification of cDNA Ends (RACE) (see, for example, Frohman *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:8998-9002). Recent modifications of the technique, exemplified by the
30 Marathon™ technology (Clontech Laboratories, Inc.) for example, have significantly simplified obtaining full-length cDNA sequences.

Another aspect of the present invention relates to isolated acyltransferase polypeptides. Such polypeptides include isolated polypeptides set forth in the Sequence Listing, as well as polypeptides and fragments thereof, particularly those polypeptides which exhibit acyltransferase activity and also those polypeptides which have at least 50%, 60% or 70% identity, preferably at least 80% identity, more preferably at least 90% identity, and most preferably at least 95% identity to a polypeptide sequence selected from the group of sequences set forth in the Sequence Listing, and also include portions of such polypeptides, wherein such portion of the polypeptide preferably includes at least 30 amino acids and more preferably includes at least 50 amino acids.

“Identity”, as is well understood in the art, is a relationship between two or more polypeptide sequences or two or more polynucleotide sequences, as determined by comparing the sequences. In the art, “identity” also means the degree of sequence relatedness between polypeptide or polynucleotide sequences, as determined by the match between strings of such sequences. “Identity” can be readily calculated by known methods including, but not limited to, those described in *Computational Molecular Biology*, Lesk, A.M., ed., Oxford University Press, New York (1988); *Biocomputing: Informatics and Genome Projects*, Smith, D.W., ed., Academic Press, New York, 1993; *Computer Analysis of Sequence Data, Part I*, Griffin, A.M. and Griffin, H.G., eds., Humana Press, New Jersey (1994); *Sequence Analysis in Molecular Biology*, von Heinje, G., Academic Press (1987); *Sequence Analysis Primer*, Gribskov, M. and Devereux, J., eds., Stockton Press, New York (1991); and Carillo, H., and Lipman, D., *SIAM J Applied Math*, 48:1073 (1988). Methods to determine identity are designed to give the largest match between the sequences tested. Moreover, methods to determine identity are codified in publicly available programs. Computer programs which can be used to determine identity between two sequences include, but are not limited to, GCG (Devereux, J., et al., *Nucleic Acids Research* 12(1):387 (1984); suite of five BLAST programs, three designed for nucleotide sequences queries (BLASTN, BLASTX, and TBLASTX) and two designed for protein sequence queries (BLASTP and TBLASTN) (Coulson, *Trends in Biotechnology*, 12: 76-80 (1994); Birren, et al., *Genome Analysis*, 1: 543-559 (1997)). The BLAST X program is publicly available from NCBI and other sources (BLAST Manual, Altschul, S., et al., NCBI NLM NIH, Bethesda, MD 20894; Altschul, S., et al., *J. Mol. Biol.*, 215:403-410 (1990)). The well known Smith Waterman algorithm can also be used to determine identity.

Parameters for polypeptide sequence comparison typically include the following:

Algorithm: Needleman and Wunsch, *J. Mol. Biol.* 48:443-453 (1970)

Comparison matrix: BLOSSUM62 from Hentikoff and Hentikoff, *Proc. Natl. Acad. Sci USA* 89:10915-10919 (1992)

5 Gap Penalty: 12

Gap Length Penalty: 4

A program which can be used with these parameters is publicly available as the "gap" program from Genetics Computer Group, Madison Wisconsin. The above parameters along with no penalty for end gap are the default parameters for peptide comparisons.

10 Parameters for polynucleotide sequence comparison include the following:

Algorithm: Needleman and Wunsch, *J. Mol. Biol.* 48:443-453 (1970)

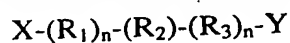
Comparison matrix: matches = +10; mismatches = 0

Gap Penalty: 50

Gap Length Penalty: 3

15 A program which can be used with these parameters is publicly available as the "gap" program from Genetics Computer Group, Madison Wisconsin. The above parameters are the default parameters for nucleic acid comparisons.

The invention also includes polypeptides of the formula:



20 wherein, at the amino terminus, X is hydrogen, and at the carboxyl terminus, Y is hydrogen or a metal, R_1 and R_3 are any amino acid residue, n is an integer between 1 and 1000, and R_2 is an amino acid sequence of the invention, particularly an amino acid sequence selected from the group set forth in the Sequence Listing and preferably SEQ ID NOs: 2, 4, 6, 8, 11, 13, 15, 17, 19, 21, 23, and 218-225. In the formula, R_2 is oriented so that its amino terminal residue
25 is at the left, bound to R_1 , and its carboxy terminal residue is at the right, bound to R_3 . Any stretch of amino acid residues denoted by either R group, where R is greater than 1, may be either a heteropolymer or a homopolymer, preferably a heteropolymer.

Polypeptides of the present invention include isolated polypeptides encoded by a polynucleotide comprising a sequence selected from the group of a sequence contained in
30 SEQ ID NOs: 1, 3, 5, 7, 9, 10, 12, 14, 16, 18, 20, 22, and 226-233.

The polypeptides of the present invention can be mature protein or can be part of a fusion protein.

Fragments and variants of the polypeptides are also considered to be a part of the invention. A fragment is a variant polypeptide which has an amino acid sequence that is entirely the same as part but not all of the amino acid sequence of the previously described polypeptides. The fragments can be "free-standing" or comprised within a larger polypeptide of which the fragment forms a part or a region, most preferably as a single continuous region. Preferred fragments are biologically active fragments which are those fragments that mediate activities of the polypeptides of the invention, including those with similar activity or improved activity or with a decreased activity. Also included are those fragments that antigenic or immunogenic in an animal, particularly a human.

Variants of the polypeptide also include polypeptides that vary from the sequences set forth in the Sequence Listing by conservative amino acid substitutions, substitution of a residue by another with like characteristics. In general, such substitutions are among Ala, Val, Leu and Ile; between Ser and Thr; between Asp and Glu; between Asn and Gln; between Lys and Arg; or between Phe and Tyr. Particularly preferred are variants in which 5 to 10; 1 to 5; 1 to 3 or one amino acid(s) are substituted, deleted, or added, in any combination.

Variants that are fragments of the polypeptides of the invention can be used to produce the corresponding full length polypeptide by peptide synthesis. Therefore, these variants can be used as intermediates for producing the full-length polypeptides of the invention.

The polynucleotides and polypeptides of the invention can be used, for example, in the transformation of various host cells, as further discussed herein.

The invention also provides polynucleotides that encode a polypeptide that is a mature protein plus additional amino or carboxyl-terminal amino acids, or amino acids within the mature polypeptide (for example, when the mature form of the protein has more than one polypeptide chain). Such sequences can, for example, play a role in the processing of a protein from a precursor to a mature form, allow protein transport, shorten or lengthen protein half-life, or facilitate manipulation of the protein in assays or production. It is contemplated that cellular enzymes can be used to remove any additional amino acids from the mature protein.

A precursor protein, having the mature form of the polypeptide fused to one or more prosequences may be an inactive form of the polypeptide. The inactive precursors generally are activated when the prosequences are removed. Some or all of the prosequences may be removed prior to activation. Such precursor protein are generally called proproteins.

The polynucleotide and polypeptide sequences can also be used to identify additional sequences which are homologous to the sequences of the present invention. The most preferable and convenient method is to store the sequence in a computer readable medium, for example, floppy disk, CD ROM, hard disk drives, external disk drives and DVD, and then to use the stored sequence to search a sequence database with well known searching tools.

Examples of public databases include the DNA Database of Japan

(DDBJ)(<http://www.ddbj.nig.ac.jp/>); Genebank

(<http://www.ncbi.nlm.nih.gov/web/Genbank/Index.html>); and the European Molecular

Biology Laboratory Nucleic Acid Sequence Database (EMBL)

(http://www.ebi.ac.uk/ebi_docs/embl_db.html). A number of different search algorithms are available to the skilled artisan, one example of which are the suite of programs referred to as

BLAST programs. There are five implementations of BLAST, three designed for nucleotide sequences queries (BLASTN, BLASTX, and TBLASTX) and two designed for protein sequence queries (BLASTP and TBLASTN) (Coulson, *Trends in Biotechnology*, 12: 76-80

(1994); Birren, *et al.*, *Genome Analysis*, 1: 543-559 (1997)). Additional programs are available in the art for the analysis of identified sequences, such as sequence alignment programs, programs for the identification of more distantly related sequences, and the like, and are well known to the skilled artisan.

20 Plant Constructs and Methods of Use

Of interest in the present invention, is the use of the nucleotide sequences, or polynucleotides, in recombinant DNA constructs to direct the transcription or transcription and translation (expression) of the acyltransferase sequences of the present invention in a host cell.

Of particular interest is the use of the nucleotide sequences, or polynucleotides, in recombinant DNA constructs to direct the transcription or transcription and translation (expression) of the acyltransferase sequences of the present invention in a host cell. The expression constructs generally comprise a promoter functional in a host cell operably linked to a nucleic acid sequence encoding an acyltransferase of the present invention and a transcriptional termination region functional in a host cell.

By "host cell" is meant a cell which contains a vector and supports the replication, and/or transcription or transcription and translation (expression) of the expression construct.

Host cells for use in the present invention can be prokaryotic cells, such as *E. coli*, or eukaryotic cells such as yeast, plant, insect, amphibian, or mammalian cells. Preferably, host cells are monocotyledenous or dicotyledenous plant cells.

Of particular interest in the present invention is the use of the polynucleotides of the present invention for the preparation of constructs to direct the transcription or transcription and translation of the nucleotide sequences encoding an acyltransferase in a host plant cell. Plant expression constructs generally comprise a promoter functional in a plant host cell operably linked to a nucleic acid sequence of the present and a transcriptional termination region functional in a host plant cell.

Those skilled in the art will recognize that there are a number of promoters which are functional in plant cells, and have been described in the literature. Chloroplast and plastid specific promoters, chloroplast or plastid functional promoters, and chloroplast or plastid operable promoters are also envisioned.

One set of promoters are constitutive promoters such as the CaMV35S or FMV35S promoters that yield high levels of expression in most plant organs. Enhanced or duplicated versions of the CaMV35S and FMV35S promoters are useful in the practice of this invention (Odell, *et al.* (1985) *Nature* 313:810-812; Rogers, U.S. Patent Number 5,378, 619). In addition, it may also be preferred to bring about expression of the protein of interest in specific tissues of the plant, such as leaf, stem, root, tuber, seed, fruit, etc., and the promoter chosen should have the desired tissue and developmental specificity.

Of particular interest is the expression of the nucleic acid sequences of the present invention from transcription initiation regions which are preferentially expressed in a plant seed tissue. Examples of such seed preferential transcription initiation sequences include those sequences derived from sequences encoding plant storage protein genes or from genes involved in fatty acid biosynthesis in oilseeds. Examples of such promoters include the 5' regulatory regions from such genes as napin (Kridl *et al.*, *Seed Sci. Res.* 1:209:219 (1991)), phaseolin, zein, soybean trypsin inhibitor, ACP, stearyl-ACP desaturase, soybean α' subunit of β -conglycinin (soy 7s, (Chen *et al.*, *Proc. Natl. Acad. Sci.*, 83:8560-8564 (1986))) and oleosin.

It may be advantageous to direct the localization of proteins conferring acyltransferase to a particular subcellular compartment, for example, to the mitochondrion, endoplasmic reticulum, vacuoles, chloroplast or other plastidic compartment. For example, where the genes of interest of the present invention will be targeted to plastids, such as chloroplasts, for

expression, the constructs will also employ the use of sequences to direct the gene to the plastid. Such sequences are referred to herein as chloroplast transit peptides (CTP) or plastid transit peptides (PTP). In this manner, where the gene of interest is not directly inserted into the plastid, the expression construct will additionally contain a gene encoding a transit peptide to direct the gene of interest to the plastid. The chloroplast transit peptides may be derived from the gene of interest, or may be derived from a heterologous sequence having a CTP. Such transit peptides are known in the art. See, for example, Von Heijne *et al.* (1991) *Plant Mol. Biol. Rep.* 9:104-126; Clark *et al.* (1989) *J. Biol. Chem.* 264:17544-17550; della-Cioppa *et al.* (1987) *Plant Physiol.* 84:965-968; Romer *et al.* (1993) *Biochem. Biophys. Res Commun.* 196:1414-1421; and, Shah *et al.* (1986) *Science* 233:478-481. Additional transit peptides for the translocation of the protein to the endoplasmic reticulum (ER), or vacuole may also find use in the constructs of the present invention.

Depending upon the intended use, the constructs may contain the nucleic acid sequence which encodes the entire acyltransferase protein, or a portion thereof. For example, where antisense inhibition of a given acyltransferase protein is desired, the entire sequence is not required. Furthermore, where acyltransferase sequences used in constructs are intended for use as probes, it may be advantageous to prepare constructs containing only a particular portion of a acyltransferase encoding sequence, for example a sequence which is discovered to encode a highly conserved acyltransferase region.

The skilled artisan will recognize that there are various methods for the inhibition of expression of endogenous sequences in a host cell. Such methods include, but are not limited to antisense suppression (Smith, *et al.* (1988) *Nature* 334:724-726), co-suppression (Napoli, *et al.* (1989) *Plant Cell* 2:279-289), ribozymes (PCT Publication WO 97/10328), and combinations of sense and antisense, such as those described by Waterhouse, *et al.* (1998) *Proc. Natl. Acad. Sci. USA* 95:13959-13964. Methods for the suppression of endogenous sequences in a host cell typically employ the transcription or transcription and translation of at least a portion of the sequence to be suppressed. Such sequences may be homologous to coding as well as non-coding regions of the endogenous sequence.

Regulatory transcript termination regions may be provided in plant expression constructs of this invention as well. Transcript termination regions may be provided by the DNA sequence encoding the acyltransferase or a convenient transcription termination region derived from a different gene source, for example, the transcript termination region which is naturally associated with the transcript initiation region. The skilled artisan will recognize

that any convenient transcript termination region which is capable of terminating transcription in a plant cell may be employed in the constructs of the present invention.

Alternatively, constructs may be prepared to direct the expression of the acyltransferase sequences directly from the host plant cell plastid. Such constructs and methods are known in the art and are generally described, for example, in Svab, *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87:8526-8530 and Svab and Maliga (1993) *Proc. Natl. Acad. Sci. USA* 90:913-917 and in U.S. Patent Number 5,693,507.

A plant cell, tissue, organ, or plant into which the recombinant DNA constructs containing the expression constructs have been introduced is considered transformed, transfected, or transgenic. A transgenic or transformed cell or plant also includes progeny of the cell or plant and progeny produced from a breeding program employing such a transgenic plant as a parent in a cross and exhibiting an altered genotype resulting from the presence of an introduced acyltransferase nucleic acid sequence.

The term "introduced" in the context of inserting a nucleic acid sequence into a cell, means "transfection", or "transformation" or "transduction" and includes reference to the incorporation of a nucleic acid sequence into a eukaryotic or prokaryotic cell where the nucleic acid sequence may be incorporated into the genome of the cell (for example, chromosome, plasmid, plastid, or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (for example, transfected mRNA).

Plant expression or transcription constructs having an acyltransferase as the DNA sequence of interest for increased or decreased expression thereof may be employed with a wide variety of plant life, particularly, plant life involved in the production of vegetable oils for edible and industrial uses. Plants of interest in the present invention include monocotyledenous and dicotyledenous plants. Most especially preferred are temperate oilseed crops. Plants of interest include, but are not limited to, rapeseed (Canola and High Erucic Acid varieties), sunflower, safflower, cotton, soybean, peanut, coconut and oil palms, and corn. Depending on the method for introducing the recombinant constructs into the host cell, other DNA sequences may be required. Importantly, this invention is applicable to dicotyledons and monocotyledons species alike and will be readily applicable to new and/or improved transformation and regulation techniques.

As used herein, the term "plant" includes reference to whole plants, plant organs (for example, leaves, stems, roots, etc.), seeds, and plant cells and progeny of same. Plant cell, as used herein includes, without limitation, seeds suspension cultures, embryos, meristematic

regions, callus tissue, leaves roots shoots, gametophytes, sporophytes, pollen, and microspores. The class of plants which can be used in the methods of the present invention is generally as broad as the class of higher plants amenable to transformation techniques, including both monocotyledenous and dicotyledenous plants. Particularly preferred plants of interest include, but are not limited to, rapeseed (Canola and High Erucic Acid varieties), sunflower, safflower, cotton, soybean, peanut, coconut and oil palms, and corn. Most especially preferred plants include *Brassica*, soybean, and corn.

As used herein, "transgenic plant" includes reference to a plant which comprises within its genome a heterologous polynucleotide. Generally, the heterologous polynucleotide is stably integrated within the genome such that the polynucleotide is passed on to successive generations. The heterologous polynucleotide may be integrated into the genome alone or as part of a recombinant expression cassette. "Transgenic" is used herein to include any cell, cell line, callus, tissue, plant part or plant, the genotype of which has been altered by the presence of heterologous nucleic acid including those transgenics initially so altered as well as those created by sexual crosses or asexual propagation from the initial transgenic.

Thus a plant having within its cells a heterologous polynucleotide is referred to herein as a transgenic plant. The heterologous polynucleotide can be either stably integrated into the genome, or can be extra-chromosomal. Preferably, the polynucleotide of the present invention is stably integrated into the genome such that the polynucleotide is passed on to successive generations. The polynucleotide is integrated into the genome alone or as part of a recombinant expression cassette. "Transgenic" is used herein to include any cell, cell line, callus, tissue, plant part or plant, the genotype of which has been altered by the presence of heterologous nucleic acids including those transgenics initially so altered as well as those created by sexual crosses or asexual reproduction of the initial transgenics.

As used herein, "heterologous" in reference to a nucleic acid is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by deliberate human intervention. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form. A heterologous protein may originate from a foreign species, or, if from the same species, is substantially modified from its original form by deliberate human intervention.

As used herein, a "recombinant expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements which permit transcription of a particular nucleic acid in a target cell. The recombinant expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid sequence to be transcribed and a promoter.

It is contemplated that the gene sequences may be synthesized, either completely or in part, especially where it is desirable to provide plant-preferred sequences. Thus, all or a portion of the desired structural gene (that portion of the gene which encodes the acyltransferase protein) may be synthesized using codons preferred by a selected host. Host-preferred codons may be determined, for example, from the codons used most frequently in the proteins expressed in a desired host species.

One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover "homologous" or "related" acyltransferase from a variety of plant sources. Homologous sequences are found when there is an identity of sequence, which may be determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions between a known acyltransferase and a candidate source. Conservative changes, such as Glu/Asp, Val/Ile, Ser/Thr, Arg/Lys and Gln/Asn may also be considered in determining sequence homology. Amino acid sequences are considered homologous by as little as 25% sequence identity between the two complete mature proteins. (*See generally, Doolittle, R.F., OF URFS and ORFS* (University Science Books, CA, 1986.)

Thus, other acyltransferase sequences can be obtained from the specific exemplified sequences provided herein. Furthermore, it will be apparent that one can obtain natural and synthetic sequences, including modified amino acid sequences and starting materials for synthetic-protein modeling from the exemplified sequences and from acyltransferases which are obtained through the use of such exemplified sequences. Modified amino acid sequences include sequences which have been mutated, truncated, increased and the like, whether such sequences were partially or wholly synthesized. Sequences which are actually purified from plant preparations or are identical or encode identical proteins thereto, regardless of the method used to obtain the protein or sequence, are equally considered naturally derived.

For immunological screening, antibodies to the acyltransferase protein can be prepared by injecting rabbits or mice with the purified protein or portion thereof, such methods of preparing antibodies being well known to those in the art. Either monoclonal or polyclonal antibodies can be produced, although typically polyclonal antibodies are more useful for gene isolation. Western analysis may be conducted to determine that a related protein is present in a crude extract of the desired plant species, as determined by cross-reaction with the antibodies to the acyltransferase protein. When cross-reactivity is observed, genes encoding the related proteins are isolated by screening expression libraries representing the desired plant species. Expression libraries can be constructed in a variety of commercially available vectors, including lambda gt11, as described in Sambrook, *et al.* (*Molecular Cloning: A Laboratory Manual*, Second Edition (1989) Cold Spring Harbor Laboratory, Cold Spring Harbor, New York).

The nucleic acid sequences associated with acyltransferase proteins will find many uses. For example, recombinant constructs can be prepared which can be used as probes, or which will provide for expression of the acyltransferase protein in host cells to produce a ready source of the enzyme and/or to modify the composition of triglycerides found therein. Other useful applications may be found when the host cell is a plant host cell, either *in vitro* or *in vivo*.

The modification of fatty acid compositions may also affect the fluidity of plant membranes. Different lipid concentrations have been observed in cold-hardened plants, for example. By this invention, one may be capable of introducing traits which will lend to chill tolerance. Constitutive or temperature inducible transcription initiation regulatory control regions may have special applications for such uses.

As discussed above, nucleic acid sequence encoding an acyltransferase of this invention may include genomic, cDNA or mRNA sequence. By "encoding" is meant that the sequence corresponds to a particular amino acid sequence either in a sense or anti-sense orientation. By "extrachromosomal" is meant that the sequence is outside of the plant genome of which it is naturally associated. By "recombinant" is meant that the sequence contains a genetically engineered modification through manipulation via mutagenesis, restriction enzymes, and the like.

Once the desired acyltransferase nucleic acid sequence is obtained, it may be manipulated in a variety of ways. Where the sequence involves non-coding flanking regions, the flanking regions may be subjected to resection, mutagenesis, etc. Thus, transitions,

transversions, deletions, and insertions may be performed on the naturally occurring sequence. In addition, all or part of the sequence may be synthesized. In the structural gene, one or more codons may be modified to provide for a modified amino acid sequence, or one or more codon mutations may be introduced to provide for a convenient restriction site or other purpose involved with construction or expression. The structural gene may be further modified by employing synthetic adapters, linkers to introduce one or more convenient restriction sites, or the like.

The nucleic acid or amino acid sequences encoding an acyltransferase of this invention may be combined with other non-native, or "heterologous", sequences in a variety of ways. By "heterologous" sequences is meant any sequence which is not naturally found joined to the acyltransferase, including, for example, combinations of nucleic acid sequences from the same plant which are not naturally found joined together.

The DNA sequence encoding an acyltransferase of this invention may be employed in conjunction with all or part of the gene sequences normally associated with the acyltransferase. In its component parts, a DNA sequence encoding acyltransferase is combined in a DNA construct having, in the 5' to 3' direction of transcription, a transcription initiation control region capable of promoting transcription and translation in a host cell, the DNA sequence encoding plant acyltransferase and a transcription and translation termination region.

Potential host cells include both prokaryotic cells, such as *E.coli* and eukaryotic cells such as yeast, insect, amphibian, or mammalian cells. A host cell may be unicellular or found in a multicellular differentiated or undifferentiated organism depending upon the intended use. Preferably, host cells of the present invention include plant cells, both monocotyledenous and dicotyledenous. Cells of this invention may be distinguished by having a sequence foreign to the wild-type cell present therein, for example, by having a recombinant nucleic acid construct encoding an acyltransferase therein.

The methods used for the transformation of the host plant cell are not critical to the present invention. The transformation of the plant is preferably permanent, i.e. by integration of the introduced expression constructs into the host plant genome, so that the introduced constructs are passed onto successive plant generations. The skilled artisan will recognize that a wide variety of transformation techniques exist in the art, and new techniques are continually becoming available. Any technique that is suitable for the target host plant can be employed within the scope of the present invention. For example, the constructs can be

introduced in a variety of forms including, but not limited to as a strand of DNA, in a plasmid, or in an artificial chromosome. The introduction of the constructs into the target plant cells can be accomplished by a variety of techniques, including, but not limited to calcium-phosphate-DNA co-precipitation, electroporation, microinjection, *Agrobacterium* infection, liposomes or microprojectile transformation. The skilled artisan can refer to the literature for details and select suitable techniques for use in the methods of the present invention.

Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformant cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

Where *Agrobacterium* is used for plant cell transformation, a vector may be used which may be introduced into the *Agrobacterium* host for homologous recombination with T-DNA or the Ti- or Ri-plasmid present in the *Agrobacterium* host. The Ti- or Ri-plasmid containing the T-DNA for recombination may be armed (capable of causing gall formation) or disarmed (incapable of causing gall formation), the latter being permissible, so long as the *vir* genes are present in the transformed *Agrobacterium* host. The armed plasmid can give a mixture of normal plant cells and gall.

In some instances where *Agrobacterium* is used as the vehicle for transforming host plant cells, the expression or transcription construct bordered by the T-DNA border region(s) will be inserted into a broad host range vector capable of replication in *E. coli* and *Agrobacterium*, there being broad host range vectors described in the literature. Commonly used is pRK2 or derivatives thereof. See, for example, Ditta, *et al.*, (*Proc. Nat. Acad. Sci., U.S.A.* (1980) 77:7347-7351) and EPA 0 120 515, which are incorporated herein by reference. Alternatively, one may insert the sequences to be expressed in plant cells into a vector containing separate replication sequences, one of which stabilizes the vector in *E. coli*, and the other in *Agrobacterium*. See, for example, McBride and Summerfelt (*Plant Mol. Biol.* (1990) 14:269-276), wherein the pRiHRI (Jouanin, *et al.*, *Mol. Gen. Genet.* (1985) 201:370-374) origin of replication is utilized and provides for added stability of the plant expression vectors in host *Agrobacterium* cells.

Included with the expression construct and the T-DNA will be one or more markers, which allow for selection of transformed *Agrobacterium* and transformed plant cells. A number of markers have been developed for use with plant cells, such as resistance to chloramphenicol, kanamycin, the aminoglycoside G418, hygromycin, or the like. The particular marker employed is not essential to this invention, one or another marker being preferred depending on the particular host and the manner of construction.

For transformation of plant cells using *Agrobacterium*, explants may be combined and incubated with the transformed *Agrobacterium* for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

There are several possible ways to obtain the plant cells of this invention which contain multiple expression constructs. Any means for producing a plant comprising a construct having a nucleic acid sequence of the present invention, and at least one other construct having another DNA sequence encoding an enzyme are encompassed by the present invention. For example, the expression construct can be used to transform a plant at the same time as the second construct either by inclusion of both expression constructs in a single transformation vector or by using separate vectors, each of which express desired genes. The second construct can be introduced into a plant which has already been transformed with the first expression construct, or alternatively, transformed plants, one having the first construct and one having the second construct, can be crossed to bring the constructs together in the same plant.

In general, acyltransferase proteins are active in the transfer of acyl groups from a donor to a variety of different substrates. For example, diacylglycerol acyltransferases add acyl groups to diacylglycerol to form triacylglycerol (TAG), or acyl:CoA:cholesterol acyltransferase uses an acyl-CoA as a donor to transfer an acyl group to a sterol to form a sterol ester. Typically, the substrates include, but are not limited to glycerides, including mono and diglycerides, sterols, stanols, phosphatides, and the like. Donors include, but are not limited to acyl-CoA and acyl-ACP molecules.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

5

EXAMPLES

Example 1: RNA Isolations

10

Total RNA from the inflorescence and developing seeds of *Arabidopsis thaliana* is isolated for use in construction of complementary (cDNA) libraries. The procedure is an adaptation of the DNA isolation protocol of Webb and Knapp (D.M. Webb and S.J. Knapp, (1990) Plant Molec. Reporter, 8, 180-185). The following description assumes the use of 1g fresh weight of tissue. Frozen seed tissue is powdered by grinding under liquid nitrogen. The powder is added to 10ml REC buffer (50mM Tris-HCl, pH 9, 0.8M NaCl, 10mM EDTA, 0.5% w/v CTAB (cetyltrimethyl-ammonium bromide)) along with 0.2g insoluble polyvinylpolypyrrolidone, and ground at room temperature. The homogenate is centrifuged for 5 minutes at 12,000 xg to pellet insoluble material. The resulting supernatant fraction is extracted with chloroform, and the top phase is recovered.

15

20

The RNA is then precipitated by addition of 1 volume RecP (50mM Tris-HCL pH9, 10mM EDTA and 0.5% (w/v) CTAB) and collected by brief centrifugation as before. The RNA pellet is redissolved in 0.4 ml of 1M NaCl. The RNA pellet is redissolved in water and extracted with phenol/chloroform. Sufficient 3M potassium acetate (pH 5) is added to make the mixture 0.3M in acetate, followed by addition of two volumes of ethanol to precipitate the RNA. After washing with ethanol, this final RNA precipitate is dissolved in water and stored frozen.

25

30

Alternatively, total RNA may be obtained using TRIzol reagent (BRL-Lifetechnologies, Gaithersburg, MD) following the manufacturers protocol. The RNA precipitate is dissolved in water and stored frozen.

Example 2: Identification of Acyltransferase Homology Sequences

Searches are performed on a Silicon Graphics Unix computer using additional Bioaccelerator hardware and GenWeb software supplied by Compugen Ltd. This software and hardware enables the use of the Smith-Waterman algorithm in searching DNA and protein databases using profiles as queries. The program used to query protein databases is profilesearch. This is a search where the query is not a single sequence but a profile based on a multiple alignment of amino acid or nucleic acid sequences. The profile is used to query a sequence data set, i.e., a sequence database. The profile contains all the pertinent information for scoring each position in a sequence, in effect replacing the "scoring matrix" used for the standard query searches. The program used to query nucleotide databases with a protein profile is tprofilesearch. Tprofilesearch searches nucleic acid databases using an amino acid profile query. As the search is running, sequences in the database are translated to amino acid sequences in six reading frames. The output file for tprofilesearch is identical to the output file for profilesearch except for an additional column that indicates the frame in which the best alignment occurred.

The Smith-Waterman algorithm, (Smith and Waterman (1981) *supra*), is used to search for similarities between one sequence from the query and a group of sequences contained in the database. E score values as well as other sequence information, such as conserved peptide sequences of HXXXXD and PEG are used to identify related sequences. By using the conserved peptide sequence information, E score values of greater than E-12 and E-8 are considered. For example, the EST sequence originally used to identify ATAT2 had an E score of 0.0094, while the EST sequence originally used to identify ATLPAAT1 had an E score of 0.0868.

A protein sequence of glycerol-3-phosphate from *E. coli* (Swiss Prot Accession P00482) is used to search the NCBI non-redundant protein database using BLAST. In the first round of searches, other membrane forms of G3PAAT are identified. In subsequent PSI-BLAST searches (Altschul, *et al.* (1997) *Nucleic Acids Res* 25:3389-3402), LPAATs and other acyltransferases are identified. Using sequence alignment software programs, G3PAAT and different LPAAT amino acid sequences are aligned, and a profile is generated using a homologous sequence region, between amino acids 256 and 459 of the *E. coli* sequence.

The identified 204 amino acid is used to query the protein database using PSI-BLAST. After 5 iterations of PSI-BLAST, the profile generated from this new query (Figure 1)

identified soluble forms of G3PAAT. Prior to this identification, no sequence homology had been identified between the membrane and soluble forms of G3PAAT.

5 **Example 3: Excision of PSI-BLAST Profile**

10 The profile generated from the queries using PSI-BLAST is excised from the hyper text markup language (html) file. The worldwide web (www)/html interface to psiblast at ncbi stores the current generated profile matrix in a hidden field in the html file that is returned after each iteration of psiblast. However, this matrix has been encoded into string62 (s62) format for ease of transport through html. String62 format is a simple conversion of the values of the matrix into html legal ascii characters.

15 The encoded matrix width (x axis) is 26 characters, and comprise the consensus characters, the probabilities of each amino acid in the order A,B,C,D,E,F,G,H,I,K,L,M,N, P,Q,R,S,T,V,W,X,Y,Z (where B represents D and N, and Z represents Q and E, and X represents any amino acid), gap creation value, and gap extension value.

20 The length (y axis) of the matrix corresponds to the length of the sequences identified by PSI-BLAST. The order of the amino acids corresponds to the conserved amino acid sequence of the sequences identified using PSI-BLAST, with the N-terminal end at the top of the matrix. The probabilities of other amino acids at that position are represented for each amino acid along the x axis, below the respective single letter amino acid abbreviation.

Thus, each row of the profile consists of the highest scoring (consensus) amino acid, followed by the scores for each possible amino acid at that position in sequence matrix, the score for opening a gap at that position, and the score for continuing a gap at that position.

25 The string62 file is converted back into a profile for use in subsequent searches. The gap open field is set to 11 and the gap extension field is set to 1 along the x axis. The gap creation and gap extension values are known, based on the settings given to the PSI-BLAST algorithm. The matrix is exported to the standard GCG profile form. This format can be read by GenWeb.

30 The algorithm used to convert the string62 formatted file to the matrix is outlined in Table 1.

Table 1

1. if encoded character z then the value is blast score min
2. if encoded character Z then the value is blast score max
- 5 3. else if the encoded character is uppercase then its value is (64-(ascii # of char))
4. else if the encoded character is a digit the value is ((ascii # of char)-48)
5. else if the encoded character is not uppercase then the value is ((ascii # of char) - 87)
6. ALL B positions are set to min of D and N amino acids at that row in sequence matrix
7. ALL Z positions are set to min of Q and E amino acids at that row in sequence matrix
- 10 8. ALL X positions are set to min of all amino acids at that row in sequence matrix
9. kBLAST_SCORE_MAX=999;
10. kBLAST_SCORE_MIN=-999;
11. all gap opens are set to 11
12. all gap lens are set to 1

Example 4: Identification of Novel Acyltransferase Related Amino Acid Sequences

20 The profile (Figure 1) is used in further queries to identify a number of previously unidentified proteins from yeast as novel acyltransferases. A protein is identified from an *Arabidopsis* protein sequence database (ATAT1) (SEQ ID NO:2). Sequences are also identified from nucleic acid databases (Table 2)

Table 2

Database ID Number	BLAST Search Hits	Log probability
<u><i>Saccharomyces cerevisiae</i></u>		
gi 1078509	Limnanthes putative LPAAT	e-10 (SEQ ID
NO:217)		
30 gi 586485	Limnanthes putative LPAAT	e-13 (SEQ ID
NO:218)		

	gi 320748 NO:219)	Limnanthes putative LPAAT	e-19 (SEQ ID
	gi 2506920	SUPPRESSES CTR1 (choline transport mutant) (SEQ ID NO:220)	
5	gi 549627 NO:221)	similar to CTR1	e-118 (SEQ ID
	gi 2133031 NO:222)	unidentified	(SEQ ID
	gi 2132939 NO:223)	unidentified	(SEQ ID
10	gi 2132299 NO:224)	TAFAZZIN	e-14 (SEQ ID

In Table 2, the gi number is the database identifier, the middle column shows the results of BLAST searches against the NCBI NR protein database, and the log probability number shows represents the log of the probability of such a match occurring by random chance. These proteins, including the ATAT1 protein sequence, are identified using the original PSI-BLAST search of the NCBI NR protein database. Thus, these proteins are novel acyltransferase related proteins with unidentified activities.

The *Arabidopsis* acyltransferase sequence, herein referred to as ATAT1, is also identified using the original PSI-BLAST search of the NCBI NR protein database, and did not have an annotated function.

Additional *Arabidopsis* amino acid sequences related to acyltransferases are identified from the databases, referred to as ATAT2est, ATAT3est, ATAT4est, ATAT5est, ATAT6est, ATAT7est, ATAT8est, ATAT9, ATAT10, and ATAT11est. Furthermore, *Arabidopsis* amino acid sequences are identified which demonstrate sequence similarity to known lysophosphatidic acid, referred to as ATLPAAT1. The sequences of ATAT9 and ATAT10 are identified from the database as genomic sequences, all other *Arabidopsis* sequences are identified as ESTs.

Example 5: Sequence Analysis of the Novel Acyltransferases

To obtain the entire coding region corresponding to the *Arabidopsis* acyltransferase sequences, synthetic oligo-nucleotide primers are designed to amplify the 5' and 3' ends of partial cDNA clones containing acyltransferase related sequences. Primers are designed according to the respective *Arabidopsis* acyltransferase related sequences (Table 3) and used
5 in Rapid Amplification of cDNA Ends (RACE) reactions (Frohman *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:8998-9002) using the Marathon cDNA amplification kit (Clontech Laboratories Inc, Palo Alto, CA). Primers with an R designation are used for 5' RACE reactions, and primers with an F designation are used for 3' RACE reactions.

Table 3

ATAT2

ATAT2R1 CCATCCGCTTCAAGGGAACGACACCCATCA (SEQ ID NO:135)

ATAT2R2 TCCCTGTCTTGCTTGATGAACTTAAAGCTTG (SEQ ID NO:136)

5 ATAT2R3 ACAGCAGGAGTGTCTGATGATGGCAGATTC (SEQ ID NO:137)

ATAT3

ATAT3R1 ACTGGAGTTCCAGCCAAAAATGCACCTGTC (SEQ ID NO:138)

ATAT3R2 GATACACCCTTGAAATCAGGCGATTTTGCT (SEQ ID NO:139)

10

ATAT4

ATAT4R1 TTGCAAATTCAATTCCTGTTTCACCGGGCC (SEQ ID NO:140)

ATAT4R2 GTTTTCTGCTATTCCAGAAGGCGTCAACAA (SEQ ID NO:141)

15

ATAT5

ATAT5R1 CATTGAAGATCCGTCCGTGAAGTTNCCTTACC (SEQ ID NO:142)

ATAT5R2 TCGAGCTGTGATCGATGATTGGCTGTGAAG (SEQ ID NO:143)

ATAT5F1 GTCTCTTCAAAAACACACACACACGTCTCT (SEQ ID NO:144)

ATAT5F2 GTCTCTTCAAAAACACACACACACGTCTCT (SEQ ID NO:145)

20

ATAT6

H76348-F1 GTAGAGAGCCTTACTTGCTTCGGTTTAGTC (SEQ ID NO:146)

H76348-F2 ACGTCATCGTACCTGTTGCTATTGACTCAC (SEQ ID NO:147)

H76348-R1 ACTTTTCCATTGTCAGGGACTCCTCGACAC (SEQ ID NO:148)

25 H76348-R2 ACGGTGTAGGAAGGGAAAGGATTCAAAAGG (SEQ ID NO:149)

ATAT7

ATTS0193-F1 GCGATGAACTACAGAGTCGGATTCTTCCTC (SEQ ID NO:150)

ATTS0193-F2 CCGGTTTACGAGATTACGTTCTTGAACCAG (SEQ ID NO:151)

30 ATTS0193-R1 CAATGGAGACAAGGCTCGAAAGTGCTAACC (SEQ ID NO:152)

ATTS0193-R2 ATTCTCTGAACATAGTTCGCCACGGTCATG (SEQ ID NO:153)

ATAT8

AA042618-F1 GAAATCCAACGCCTTCCCAATATCACTCTG (SEQ ID NO:154)

AA042618-F2 CTTCAACTTTCCATCAGGATCTTGGCACGT (SEQ ID NO:155)

AA042618-R1 ACCACTTGTTAGAGACCTTACCTGCTTAGG (SEQ ID NO:156)

5 AA042618-R2 TCCTACCTACACCATCCAATTTCTCGACCC (SEQ ID NO:157)

ATAT11

ATAT11R1 CTGCGTCAAGTGAGCAACTCAGTTCTTGCA (SEQ ID NO:158)

ATAT11R2 TGGGAAGCAGCACGTTGTTTCAGTATCGGAA (SEQ ID NO:159)

10 ATAT11R3 TAGCCTCTGTGTAATCTGTGCCCTCGGGGA (SEQ ID NO:160)

From the nucleic acid sequences obtained from the RACE reactions, protein sequence is predicted for each nucleic acid sequence using Macvector software. Nucleic acid sequences are provided for ATAT1 (SEQ ID NO:1), ATAT2 (SEQ ID NO:3), ATAT3 (SEQ ID NO:5), ATAT4 (SEQ ID NO:7), ATAT5 (SEQ ID NO:9), ATAT6 (SEQ ID NO:10), ATAT7 (SEQ ID NO:12), ATAT8 (SEQ ID NO:14), ATAT9 (SEQ ID NO:16), ATAT10 (SEQ ID NO:18), ATAT11 (SEQ ID NO:20) and ATLPAAT1 (SEQ ID NO:22), respectively.

The protein sequence derived from the ATAT1 (SEQ ID NO:2) nucleic acid sequence from Arabidopsis has a predicted molecular mass of 32.5 kDa, and a PI of 9.74. Alignment of the Arabidopsis acyltransferase with several LPAAT and G3PAAT shows that some of the domains that are conserved between LPAAT and G3PAAT are conserved in the new acyltransferase protein.

The ATAT2 nucleic acid sequence is predicted to encode a 312 amino acid protein (SEQ ID NO:4), with a molecular weight of 34.6 kD, and a pI of 9.99. The ATAT2 protein may also contain 2 to 3 transmembrane domains. However, the protein encoded by the ATAT2 nucleic acid sequence may be longer than predicted because of the absence of an inframe stop codon upstream of the ATG start codon used.

The ATAT3 nucleic acid sequence is predicted to encode a 398 amino acid protein (SEQ ID NO:6), with a molecular weight of 44.7 kD, and a pI of 5.62. The ATAT3 protein may contain 1 to 4 transmembrane domains. The ATAT4 nucleic acid sequence is predicted to encode a 317 amino acid protein (SEQ ID NO:8), with a molecular weight of 36.5 kD, and a pI of 9.67. The ATAT4 protein is predicted to have 2 to 5 transmembrane domains.

The ATLPAAT1 nucleic acid sequence is predicted to encode a 389 amino acid protein (SEQ ID NO:23), with a molecular weight of 43.7 kD, and a pI of 9.52. The ATLPAAT1 protein is predicted to have up to 3 transmembrane domains. The protein predicted from the ATLPAAT1 nucleic acid sequence is similar to LPAATs reported for *Brassica*, maize, and meadowfoam (described in PCT Publication WO 94/13814). The ATAT11 nucleic acid sequence is predicted to encode a 375 amino acid protein (SEQ ID NO:21), with a molecular weight of 43.5 kD, and a pI of 9.45. The deduced amino acid sequences of ATAT6 (SEQ ID NO:11), ATAT7 (SEQ ID NO:13), ATAT8 (SEQ ID NO:15), ATAT9 (SEQ ID NO:17), and ATAT10 (SEQ ID NO:19) are also provided

A sequence region approximately 30 amino acids upstream through approximately 100 amino acids downstream of the conserved amino acid sequences HXXXXD (Heath and Rock, (1998) *J. Bacteriol.* 180(6):1425-1430) and PEG (Neuwald (1997) *Curr Biol* 7:R465-R466) of the predicted amino acid sequences derived from the nucleic acid sequences of ATAT1, ATAT2, ATAT3, ATAT4, ATAT6, ATAT7, ATAT8, ATAT9, ATAT10, ATLPAAT1, and ATAT11 are compared to the amino acid sequences of lysophosphatidic acid acyltransferase (Jojoba AT (SEQ ID NO:162, the nucleic acid sequence is provided in SEQ ID NO:161), maize AT (PCT Publication WO 94/13814), PLSC coco(GenBank accession 1098605), PLSC Lim(GenBank accession 1209507), PLSC, Ecoli (GenBank accession 1209507), and PLSC Yeast(GenBank accession 464422)) and glycerol-3-phosphate acyltransferase (PLSB Ecoli(GenBank accession 130326) and PLSB Mouse(GenBank accession 2498786)) (Figure 2), and similarities are identified (Figure 2 and Figure 3).

Sequence comparisons reveal several classes of acyltransferases exist based on conserved amino acid sequences identified in the comparisons in Figure 2. For example, ATAT1, ATAT6, ATAT7, ATAT8, and ATAT9, contain the conserved amino acid sequences of VTYSXS(SEQ ID NO: 128), VXLTRXR(SEQ ID NO: 129), LXXGDLV(SEQ ID NO: 132) between the HXXXXD and PEG sequences. In addition, ATAT1, ATAT6, ATAT7, ATAT8, and ATAT9 also contain the conserved sequences CPEGT(SEQ ID NO: 130) which comprises the PEG sequence, as well as IVPVA(SEQ ID NO: 131) and VANXXQ (SEQ ID NO: 134)(Figure 2) downstream of the PEG sequence. The sequences corresponding to ATAT1, ATAT7, and ATAT9 are the most closely related in this class, with similarities between ATAT1 and ATAT9 of 67.0%, between ATAT1 and ATAT7 of 58.2% and between ATAT9 and ATAT7 of 63.9% (Figure 3B).

Sequence comparisons also demonstrate that the sequence of ATLPAAT1 is most closely related to the jojoba LPAAT (82.3% similar), and maize (78.0% similar).

Furthermore, sequence analysis demonstrates that ATAT4 is the most divergent sequence with the highest similarity to ATAT10 (18.5%). The highest similarity (15.3%) to a known sequence is with a meadowfoam (*Limnanthes douglassi*) LPAAT. However, the sequences of ATAT4 and ATAT10 share several conserved peptide sequences with the amino acid sequences of ATAT2 and ATAT3 (Figure 2), VXNHXS (SEQ ID NO: 127) where the H comprises the conserved H of the HXXXXD sequence and FXXGAF (SEQ ID NO: 133) downstream of the PEG sequence.

Example 6: Identification of Additional Acyltransferase Sequences

The novel *Arabidopsis* sequences identified above are used to search proprietary databases containing soybean and corn EST sequences. The results of this search identifies EST sequences from soybean (SEQ ID NO:24 through SEQ ID NO: 85) as well as from corn (SEQ ID NO: 86 through SEQ ID NO:126) as encoding acyltransferase related proteins.

Sequence comparisons between the various EST sequences and the complete *Arabidopsis* sequences reveals that the identified EST sequences demonstrate higher similarity to the various *Arabidopsis* sequences as determined by BLAST scores.

Expressed Sequence Tag (EST) sequences from soybean and corn databases are identified which are most closely related by BLAST score to ATAT1 (SEQ ID NOS:24-29 and SEQ ID NOS:86-88, respectively), ATAT2 (SEQ ID NO: 30 and SEQ ID NO:89, respectively), ATAT3 (SEQ ID NOS:31-35 and SEQ ID NOS:90-94, respectively), ATAT4 (SEQ ID NOS:36-44 and SEQ ID NOS:95-100, respectively), ATAT6 (SEQ ID NOS:45-49 and SEQ ID NO:101, respectively), ATAT7 (SEQ ID NOS:50-54 and SEQ ID NOS:102-103, respectively), ATAT8 (SEQ ID NOS:55-56 and SEQ ID NO:104, respectively), ATAT9 (SEQ ID NOS:57-79 and SEQ ID NOS:105-111, respectively), ATAT10 (SEQ ID NOS:80-81 and SEQ ID NO:112, respectively), ATAT11, (SEQ ID NOS:82-85 and SEQ ID NOS:123-126, respectively), and ATLPAAT1 (SEQ ID NOS: 113-122 respectively).

Example 7: Expression Construct Preparation

A series of synthetic oligo nucleotide primers were prepared for use in Polymerase Chain Reactions (PCR) to amplify the entire DNA sequences encoding the various acyltransferase sequences identified above. The sequences are listed in Table 3.

Table 3

Primer	Sequence (listed 5'-3')	SEQ ID NO:
ATAT1F	AAGCTTGCATGCGTCGACACAATGGTTCATGCGACCAAGT CAG	163
ATAT1R	GGTACCGTCGACTCACTTCTTGGTGTGTTGATAG	164
ATAT2F	GGATCCGCGGCCGCGACAATGACGAGCTTTACTACTTCCCT TCAT	165
ATAT2R	GGATCCCCTGCAGGTTAGAGATCCATTGATTCTGCAAT	166
ATAT3F	GGATCCGCGGCCGCGATAATGGAATCAGAGCTCAAAGAT	167
ATAT3R	GGATCCCCTGCAGGTCATTCTTCTTTCTGATGGAAATC	168
ATAT4F	GGATCCGCGGCCGCGACAATGACTCGTTCACAAGATGTTTC A	169
ATAT4R	GGATCCCCTGCAGGTCACTTCTCTTCCAATCTAGCCAG	170
ATAT6F	GGATCCGCGGCCGCGACAATGTCCGGTAATAAGATCTCGAC TCTTCA	171
ATAT6R	GGATCCCCTGCAGGTTATTTTTTCTTGACAACCTCCGTTAT TACCGG	172
ATAT7F	ATATCCGCGGCCGCGACAATGGTTATGGAGCAAGCTGGAA	173
ATAT7R	GGATCCCCTGCAGGTC AATGGAGACAAGGCTCGAAAGT	174
ATAT8F	GGATCCGCGGCCGCGACAATGTCCGCCAAGATTTCAATATT CC	175
ATAT8R	GGATCCCCTGCAGGTTAATTTTTCTTA ACTACTCCATT	176
ATAT9F	GGATCCGCGGCCGCGACAATGGGAGCTCAGGAGAAACGGCG CC	177
ATAT9R	GGATCCCCTGCAGGTCACGTCTTCTCCTTCTTCACCGG	178
ATAT10F	GGATCCGCGGCCGCGACAATGGCGGATCCTGATCTGTCTTC TCCT	179
ATAT10R	GGATCCCCTGCAGGTTATGTTGGGGCCAAGTCAGGTGCAA AGAT	180
ATAT11F	GGATCCGCGGCCGCAAAATGGAAAAAAGAGTG TACCAAA	181

	TTCT	
ATAT11R	GGATCCCCTGCAGGTTATTTGTTTACTAATTTGAGGGAAT	182
	TTTTTG	
ATLPAAT	TCGACCTGCAGGAAGCTTAAGGATGGTGATTGCTGC	183
1F		
ATLPAAT	GGATCCGCGGCCGCTTACTTCTCCTTCTCCG	184
1R		
YSCAT1F	GGATCCGCGGCCGCGCACAAATGTCTTTTAGGGATGTCCTAG	185
YSCAT1R	GGATCCCCTGCAGGTCAATCATCCTTACCCTTTGGTTTAC	186
	C	
YSCAT 1	ATGTCTTTTAGGGATGTCCTAGAAAGAGGAGATGAATTTT	187
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 1	TCAATCATCCTTACCCTTTGGTTTACCCTCTGGAGGCAGA	188
KO R	AGATTGTACTGAGAGTGCAC	
YSCAT2F	GGATCCGCGGCCGCGCACAAATGAAGCATTCCCAAAAATACCG	189
	TAGG	
YSCAT2R	GGATCCCCTGCAGGTCAATGATTTTTTTTCATCACAAATA	190
	C	
YSCAT 2	ATGAAGCATTCCCAAAAATACCGTAGGTATGGAATTTATG	191
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 2	TCAATGATTTTTTTTCATCACAAATACAAGAATAAGAAAA	192
KO R	AGATTGTACTGAGAGTGCAC	
YSCAT	GGATCCGCGGCCGCGCACAAATGGGTTTTGTGATTCTTCGA	193
3F	AAC	
YSCAT	GGATCCCCTGCAGGTTATTTGGTCTCAATTTTAATATTTT	194
3R	TTTGC	
YSCAT 3	ATGGGTTTTGTGATTCTTTCGAAACATATATGGTCGGTT	195
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 3	TTATTTGGTCTCAATTTTAATATTTTTTTTGCAAGGACTCG	196
KO R	AGATTGTACTGAGAGTGCAC	
YSCAT	GGATCCGCGGCCGCGCACAAATGGAAAAGTACACCAATTGGAG	197
4F	AGAC	
YSCAT	GGATCCCCTGCAGGCTACTTCCTCTTTTACGTTGATCGC	198
4R	TG	
YSCAT 4	ATGGAAAAGTACACCAATTGGAGAGACAATGGTACGGGAA	199
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 4	CTACTTCCTCTTTTTACGTTGATCGCTGATATATTCCTTC	200
KO R	AGATTGTACTGAGAGTGCAC	

YSCAT	GGATCCGCGGCCGCACAATGCCTGCACCAAACTCACGGA	201
5F	G	
YSCAT	GGATCCCCTGCAGGCTACGCATCTCCTTCTTTCCCTTC	202
5R		
YSCAT 5	ATGCCTGCACCAAACTCACGGAGAAATCTGCCTCTTCCA	203
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 5	CTACGCATCTCCTTCTTTCCCTTCTTCTTCTTCTCCTCT	204
KO R	AGATTGTACTGAGAGTGCAC	
YSCAT	GGATCCGCGGCCGCACAATGTCTGCTCCCGCTGCCGATCA	205
6F	TAACGC	
YSCAT	GGATCCCCTGCAGGTCATTCTTTCTTTTCGTGTTCTCTTT	206
6R	TCTG	
YSCAT 6	ATGTCTGCTCCCGCTGCCGATCATAACGCTGCCAAACCTA	207
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 6	TCATTCTTTCTTTTCGTGTTCTCTTTTCTGTCTTACCAGC	208
KO R	AGATTGTACTGAGAGTGCAC	
YSCAT	GGATCCGCGGCCGCACAATGCTGCATCAAAAAATAGCTCA	209
7F	TAAAGTTCG	
YSCAT	GGATCCCCTGCAGGTCAAAAAATAAAACAATAAAGTTTAT	210
7R	AAACTAACC	
YSCAT 7	ATGCTGCATCAAAAAATAGCTCATAAAGTTCGAAAAGTCG	211
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 7	TCAAAAAATAAAACAATAAAGTTTATAAACTAACCAAATT	212
KO R	AGATTGTACTGAGAGTGCAC	
YSCAT	GGATCCGCGGCCGCACAATGAGTGTGATAGGTAGGTCTT	213
8F	G	
YSCAT	GGATCCCCTGCAGGTTAATGCATCTTTTTTACAGATGAAC	214
8R	C	
YSCAT 8	ATGAGTGTGATAGGTAGGTTCTTGTATTACTTGAGGTCCG	215
KO F	CTGTGCGGTATTTTCACACCG	
YSCAT 8	TTAATGCATCTTTTTTACAGATGAACCTTCGTTATGGGTA	216
KO R	AGATTGTACTGAGAGTGCAC	

The entire coding regions for each of the acyltransferase sequences were amplified using the respective primers listed in the Table 3 above, cloned into the vector pCR2.1Topo (Invitrogen) or pZero (Invitrogen), and labeled as pCGN8558 (ATAT1), pCGN8564

(ATAT2), pCGB8565 (ATAT3), pCGN8566 (ATAT4), pCGN8918 (ATAT6), pCGN8913 (ATAT7), pCGN8904 (ATAT8), pCGN9970 (ATAT9), pCGN9940 (ATAT10), pCGN8567 (ATAT11), pCGN8632 (ATLPAAT1), pCGN9901 (YSCAT1 also referred to as gi2132299), pCGN9902 (YSCAT2, also referred to as gi1078509), pCGN9903 (YSCAT3, also referred to as gi2132939), pCGN9904 (YSCAT4, also referred to gi2133031), pCGN9905 (YSCAT5, also referred to as gi320748), pCGN9906 (YSCAT6, also referred to as gi549627), pCGN9907 (YSCAT7, also referred to as gi586485), and pCGN9908 (YSCAT8, also referred to as gi464422). The nucleic acid sequences for the respective yeast acyltransferase are provided YSCAT1 (SEQ ID NO:225), YSCAT2 (SEQ ID NO:226), YSCAT3 (SEQ ID NO:227), YSCAT4 (SEQ ID NO:228), YSCAT5 (SEQ ID NO:229), YSCAT6 (SEQ ID NO:230), YSCAT7 (SEQ ID NO:231), and YSCAT8 (SEQ ID NO:232).

7A. Baculovirus Expression Constructs

Constructs are prepared to direct the expression of the *Arabidopsis* ATAT sequences in cultured insect cells. The entire coding regions of ATAT1, 2, 3, 4, 6, 7, 8, 9, 10, and 11 are cloned into the vector pFastBac1 (Gibco-BRL, Gaithersburg, MD) digested with *NotI* and *PstI*. The respective coding sequences were cloned as *NotI/Sse8387I* fragments. Double stranded DNA sequence was obtained to verify that no errors were introduced by PCR amplification. The resulting plasmid were designated pCGN9723 (ATAT1), pCGN9724 (ATAT2), pCGN9725 (ATAT3), pCGN9726 (ATAT4), pCGN9727 (ATAT5), pCGN9728 (ATAT7), pCGN9729 (ATAT8), pCGN9991 (ATAT9) pCGN9730 (ATAT10), pCGN9731 (ATAT11).

7B. Plant Expression Construct Preparation

A plasmid containing the napin cassette derived from pCGN3223 (described in USPN 5,639,790, the entirety of which is incorporated herein by reference) was modified to make it more useful for cloning large DNA fragments containing multiple restriction sites, and to allow the cloning of multiple napin fusion genes into plant binary transformation vectors. An adapter comprised of the self annealed oligonucleotide of sequence CGCGATTTAAATGGCGCGCCCTGCAGGCGGCCCTGCAGGGCGCGCCATTTAA (SEQ ID NO:233) AT was ligated into the cloning vector pBC SK+ (Stratagene) after digestion with the restriction endonuclease BssHII to construct vector pCGN7765. Plasmids pCGN3223 and pCGN7765 were digested with *NotI* and ligated together. The resultant vector, pCGN7770, contains the pCGN7765 backbone with the napin seed specific expression cassette from pCGN3223.

The cloning cassette, pCGN7787, essentially the same regulatory elements as pCGN7770, with the exception of the napin regulatory regions of pCGN7770 have been replaced with the double CAMV 35S promoter and the tml polyadenylation and transcriptional termination region.

A binary vector for plant transformation, pCGN5139, was constructed from pCGN1558 (McBride and Summerfelt, (1990) Plant Molecular Biology, 14:269-276). The polylinker of pCGN1558 was replaced as a HindIII/Asp718 fragment with a polylinker containing unique restriction endonuclease sites, *AscI*, *PacI*, *XbaI*, *SwaI*, *BamHI*, and *NotI*. The Asp718 and HindIII restriction endonuclease sites are retained in pCGN5139.

A series of turbo binary vectors are constructed to allow for the rapid cloning of DNA sequences into binary vectors containing transcriptional initiation regions (promoters) and transcriptional termination regions.

The plasmid pCGN8618 was constructed by ligating oligonucleotides 5'-
5 TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGG-3') (SEQ ID NO:234) and 5'-
TCGACCTGCAGGAAGCTTGCGGCCGCGGATCC-3') (SEQ ID NO:235) into SalI/XhoI-
digested pCGN7770. A fragment containing the napin promoter, polylinker and napin 3'
region was excised from pCGN8618 by digestion with Asp718I; the fragment was blunt-
ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that
10 had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs
with Klenow fragment. A plasmid containing the insert oriented so that the napin promoter
was closest to the blunted Asp718I site of pCGN5139 and the napin 3' was closest to the
blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation
and the integrity of cloning junctions. The resulting plasmid was designated pCGN8622.

15 The plasmid pCGN8619 was constructed by ligating oligonucleotides 5'-
TCGACCTGCAGGAAGCTTGCGGCCGCGGATCC -3') (SEQ ID NO:236) and 5'-
TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGG-3') (SEQ ID NO:237) into SalI/XhoI-
digested pCGN7770. A fragment containing the napin promoter, polylinker and napin 3'
region was removed from pCGN8619 by digestion with Asp718I; the fragment was blunt-
20 ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that
had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs
with Klenow fragment. A plasmid containing the insert oriented so that the napin promoter
was closest to the blunted Asp718I site of pCGN5139 and the napin 3' was closest to the
blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation
25 and the integrity of cloning junctions. The resulting plasmid was designated pCGN8623.

The plasmid pCGN8620 was constructed by ligating oligonucleotides 5'-
TCGAGGATCCGCGGCCGCAAGCTTCCTGCAGGAGCT -3') (SEQ ID NO:238) and 5'-
CCTGCAGGAAGCTTGCGGCCGCGGATCC-3') (SEQ ID NO:239) into SalI/SacI-
digested pCGN7787. A fragment containing the d35S promoter, polylinker and tml 3' region
30 was removed from pCGN8620 by complete digestion with Asp718I and partial digestion with
NotI. The fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment
then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-
ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert

oriented so that the d35S promoter was closest to the blunted Asp718I site of pCGN5139 and the tml 3' was closest to the blunted HindIII site was subjected to sequence analysis to confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8624.

5 The plasmid pCGN8621 was constructed by ligating oligonucleotides 5'-TCGACCTGCAGGAAGCTTGCGGCCGCGGATCCAGCT -3') (SEQ ID NO:240) and 5'-GGATCCGCGGCCGCAAGCTTCCTGCAGG-3') (SEQ ID NO:241) into SalI/SacI-digested pCGN7787. A fragment containing the d35S promoter, polylinker and tml 3' region was removed from pCGN8621 by complete digestion with Asp718I and partial digestion with
10 NotI. The fragment was blunt-ended by filling in the 5' overhangs with Klenow fragment then ligated into pCGN5139 that had been digested with Asp718I and HindIII and blunt-ended by filling in the 5' overhangs with Klenow fragment. A plasmid containing the insert oriented so that the d35S promoter was closest to the blunted Asp718I site of pCGN5139 and the tml 3' was closest to the blunted HindIII site was subjected to sequence analysis to
15 confirm both the insert orientation and the integrity of cloning junctions. The resulting plasmid was designated pCGN8625.

The coding regions of the various acyltransferase sequences were cloned as *NotI/Sse8387I* fragments into pCGN8622, pCGN8623, pCGN8624, and pCGN8625, for expression in sense or antisense orientations from a tissue preferential promoter, napin, or the
20 35S promoter. Fragments which were cloned into the pCGN8622 vector created the constructs pCGN8901 (ATAT1), pCGN8571 (ATAT2), pCGN8909 (ATAT3), pCGN8596 (ATAT4), pCGN8919 (ATAT6), pCGN8914 (ATAT7), pCGN8905 (ATAT8), pCGN9973 (ATAT9), pCGN9942 (ATAT10), pCGN8575 (ATAT11), and pCGN8633 (ATLPAAT1) for the sense expression of the respective coding sequences from the napin promoter. Fragments
25 which were cloned into the pCGN8623 vector created the constructs pCGN8900 (ATAT1), pCGN8572 (ATAT2), pCGN8910 (ATAT3), pCGN8597 (ATAT4), pCGN8920 (ATAT6), pCGN8915 (ATAT7), pCGN8906 (ATAT8), pCGN9972 (ATAT9), pCGN9943 (ATAT10), pCGN8576 (ATAT11), and pCGN8634 (ATLPAAT1) for the antisense expression of the respective coding sequences from the napin promoter. Fragments which were cloned into the
30 pCGN8624 vector created the constructs pCGN8903 (ATAT1), pCGN8573 (ATAT2), pCGN8911 (ATAT3), pCGN8598 (ATAT4), pCGN8921 (ATAT6), pCGN8916 (ATAT7), pCGN8907 (ATAT8), pCGN9971 (ATAT9), pCGN9944 (ATAT10), pCGN8577 (ATAT11), and pCGN8635 (ATLPAAT1) for the sense expression of the respective coding sequences

from the 35S promoter. Fragments which were cloned into the pCGN8625 vector created the constructs pCGN8902 (ATAT1) and pCGN9974 (ATAT9) for the antisense expression of the respective coding sequences from the 35S promoter.

In addition, the yeast acyltransferase coding sequences were cloned into the vector pCGN8624 creating the constructs pCGN9926 (YSCAT1), pCGN9927 (YSCAT2), pCGN9928 (YSCAT3), pCGN9929 (YSCAT4), pCGN9930 (YSCAT5), pCGN9931 (YSCAT6), pCGN9932 (YSCAT7), and pCGN9933 (YSCAT8). These constructs allow for the sense expression of the respective acyltransferase coding sequences from the 35S promoter in plant cells.

Example 8: Plant Transformation

A variety of methods have been developed to insert a DNA sequence of interest into the genome of a plant host to obtain the transcription or transcription and translation of the sequence to effect phenotypic changes.

Transgenic *Brassica* plants are obtained by *Agrobacterium*-mediated transformation as described by Radke *et al.* (*Theor. Appl. Genet.* (1988) 75:685-694; *Plant Cell Reports* (1992) 11:499-505). Transgenic *Arabidopsis thaliana* plants may be obtained by *Agrobacterium*-mediated transformation as described by Valverkens *et al.*, (*Proc. Nat. Acad. Sci.* (1988) 85:5536-5540), or as described by Bent *et al.* ((1994), *Science* 265:1856-1860), or Bechtold *et al.* ((1993), *C.R.Acad.Sci, Life Sciences* 316:1194-1199) or Clough, *et al.* (1998) *Plant J.*, 16:735-43. Other plant species may be similarly transformed using related techniques.

Alternatively, microprojectile bombardment methods, such as described by Klein *et al.* (*Bio/Technology* 10:286-291) may also be used to obtain nuclear transformed plants.

The above results demonstrate that the nucleic acid sequences identified encode proteins which are related to protein sequences encoding acyltransferase proteins. Such acyltransferase sequences find use in preparing expression constructs for plant transformations.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All

publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

- 5 Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

Claims

What is Claimed is:

1. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like
5 proteins,
wherein said enzyme includes the amino acid sequence of SEQ ID NO: 127
(VxNHxS) wherein the H is the conserved Histidine residue in the conserved peptide
sequence HXXXXD of said acyltransferase-like protein, x representing any amino acid.

10 2. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like
proteins,
wherein said enzyme includes the amino acid sequence of SEQ ID NO: 128
(VTYSxS) within about 30 amino acids downstream from the conserved amino acid sequence
HXXXXD of said acyltransferase-like protein, x representing any amino acid.

15 3. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like
proteins,
wherein said enzyme includes the amino acid sequence of SEQ ID NO: 129
(VxLTRxR) within about 60 amino acids downstream from the conserved amino acid
20 sequence HXXXXD of said acyltransferase-like protein, x representing any amino acid.

4. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like
proteins,
wherein said enzyme includes the amino acid sequence of SEQ ID NO: 132
25 (LxxGDLV) within about 20 amino acids upstream of the conserved amino acid sequence
PEG of said acyltransferase-like protein, x representing any amino acid.

5. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like
proteins,
30 wherein said enzyme includes the amino acid sequence of SEQ ID NO: 130 (CPEGT)
containing the conserved amino acid sequence PEG of said acyltransferase-like protein.

6. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like proteins,

wherein said enzyme includes the amino acid sequence of SEQ ID NO: 133 (FxxGAF) within about 20 amino acids downstream from the conserved amino acid sequence PEG of said acyltransferase-like protein, x representing any amino acid.

7. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like proteins,

wherein said enzyme includes the amino acid sequence of SEQ ID NO: 131 (IVPVA) within about 40 amino acids downstream from the conserved amino acid sequence PEG of said acyltransferase-like protein.

8. An isolated DNA sequence encoding an enzyme of the class of acyltransferase-like proteins,

wherein said enzyme includes the amino acid sequence of SEQ ID NO: 134 (VANxxQ) within about 110 amino acids downstream from the conserved amino acid sequence PEG of said acyltransferase-like protein, x representing any amino acid.

9. A DNA sequence encoding an enzyme of the class of acyltransferase-like proteins, said DNA sequence obtainable by the steps comprising:

(a) using the profile of Figure 1 to search a nucleic acid sequence database;

(b) obtaining a probability score for nucleic acid sequences in said sequence database using the Smith-Waterman algorithm; and

(c) selecting a nucleic acid sequence having a probability score of less than about 1.

10. The DNA encoding sequence according to Claim 9, wherein said DNA sequence is an encoding sequence.

11. The DNA encoding sequence according to Claim 9, wherein said DNA sequence is an EST.

12. The DNA encoding sequence according to any one of Claims 1 to 11, wherein said acyltransferase-like protein is from a plant.

13. A construct comprising a DNA sequence of any one of Claims 1 to 11 linked to a
5 heterologous transcriptional and translational initiation region functional in a host cell.

14. The construct according to Claim 13 wherein said host cell is a plant cell.

15. A plant cell comprising a DNA construct according to Claim 13.

10

16. A plant comprising a cell according to Claim 15.

17. The DNA encoding sequence of any one of 1 to 11 wherein said acyltransferase-
15 like protein is from *Arabidopsis thaliana*.

18. The DNA encoding sequence of any one of 1 to 11 wherein said acyltransferase-
like protein is from corn.

19. The DNA encoding sequence of Claim 18 wherein said sequence comprises and
20 EST selected from the group consisting of SEQ ID NO: 86 through SEQ ID NO: 126.

20. The DNA encoding sequence of any one of 1 to 11 wherein said acyltransferase-
like protein is from soybean.

25

21. The DNA encoding sequence of Claim 20 wherein said sequence comprises and
EST selected from the group consisting of SEQ ID NO: 24 through SEQ ID NO: 85.

22. The DNA encoding sequence of any one of Claims 2, 3, 4, 5, 7 and 8 wherein
30 said acyltransferase-like protein is selected from the group consisting of SEQ ID NO: 1, SEQ
ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14 and SEQ ID NO: 16.

23 . The DNA encoding sequence of either of Claim 1 and Claim 6 wherein said acyltransferase-like protein is selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7 and SEQ ID NO: 18.

Con	A	B	C	D	E	F	G	H	I	K	L	M	N	P	Q	R	S	T	V	W	X	Y	Z	Gap	Len
!10	0	0	-1	-1	0	-2	-1	-1	-2	0	-2	-1	0	-1	0	-1	5	0	-2	-3	-4	-2	0	11	1
S	0	-1	-3	-3	0	-3	-2	-1	-2	5	3	5	0	-1	0	0	1	0	-2	-4	-4	-2	0	11	1
K	-2	-1	-3	-1	0	-1	-1	-1	-2	3	1	0	2	-2	4	4	-1	-1	-1	-3	-3	-2	0	11	1
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!40	-2	-5	-4																						

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Figure 2/5

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Figure 3/5

Y	2	-4	-4	-4	-4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
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Figure 4/5

1170	5	-4	-2	-4	-3	-4	0	-4	1	-3	0	-2	-4	-3	-3	-3	0	0	1	-5	-5	-4	-3	-11	1
A	-5	-3	-5	-5	-4	0	-5	-4	2	-4	2	3	-4	-4	0	-1	-3	-1	3	-4	-5	1	-4	11	1
M	-2	-5	-3	0	3	0	-1	-2	-1	0	2	2	2	-2	-3	-1	0	-1	-3	3	-5	-3	3	11	1
E	-4	4	4	-4	-3	-4	0	-4	-2	0	0	-2	-2	-3	-3	-3	-1	0	0	-5	-5	-4	-3	11	1
A	0	-4	0	-4	-1	1	-5	1	0	0	3	4	-4	-4	-3	-3	0	-2	2	-4	-5	-2	-3	11	1
M	1	-2	-4	1	0	-4	-1	-3	-3	2	0	-2	-2	0	2	0	1	-2	-1	-5	-5	-4	0	11	1
K	0	-4	-4	-4	1	2	-1	2	-4	1	-2	3	0	0	2	3	1	-1	-4	-5	-3	0	11	1	
R	0	2	-4	0	1	1	0	1	1	0	0	3	2	2	-3	0	0	-2	0	-5	-3	0	11	1	
M	-3	0	-4	0	-2	-3	0	-3	0	-2	0	3	1	1	0	3	0	-3	-3	-5	-3	-3	-2	11	1
R	0	-4	-4	0	0	-3	0	-3	0	4	-3	0	0	-4	0	-1	0	-2	-4	-4	0	0	11	1	
K	0	0	-4	0	0	0	-1	-3	0	0	-3	0	2	-3	0	0	0	1	-2	-4	-4	0	11	1	
1180	0	-3	-3	-3	-3	1	-3	-3	0	0	-2	0	1	2	-3	0	0	2	1	-4	-4	-3	11	1	
P	-3	0	-5	-4	1	-5	2	-1	-5	3	-4	0	0	0	3	-1	-2	-3	-2	-5	-4	1	11	1	
D	-4	-4	4	-4	-4	-4	-4	-4	3	0	0	-1	-4	0	0	0	-1	-1	3	-5	-3	-4	11	1	
C	0	-4	4	-4	-3	-2	-4	-3	0	0	0	-3	-3	5	-3	-3	0	1	-1	-4	-4	-3	11	1	
P	-3	-5	-3	-5	-5	-2	-6	-5	6	-2	2	-1	-5	-1	-5	-4	-4	-3	3	-5	-6	-3	11	1	
I	-1	-5	-3	-5	-1	-1	-5	-3	4	-4	1	-1	-5	-4	-4	-5	-4	-4	4	-6	-4	-4	11	1	
I	0	-3	-5	-3	0	-6	-4	-4	-5	-3	-5	-4	-3	8	-4	-5	-2	-3	-4	-6	-5	-3	11	1	
P	0	-5	4	-5	-4	-1	-5	-5	2	-4	0	2	-5	-4	-4	-5	-4	0	6	-5	-3	-4	11	1	
V	1	-3	4	-3	-4	-3	2	-5	-1	-3	-3	-3	0	-4	-3	0	2	3	0	3	-4	0	11	1	
T	1	-5	0	-5	-4	-3	0	-3	-1	-3	-4	-4	-4	-4	-3	0	2	3	0	3	-4	0	11	1	
I	-3	-5	0	-5	-5	0	-6	0	6	-4	1	4	-5	-5	-4	-4	-4	-3	2	-4	2	-5	11	1	
1190	0	-3	-4	-4	-3	-4	3	0	-1	0	-3	0	0	-4	0	0	1	0	1	1	-4	-1	11	1	
G	-3	-4	-4	-2	0	2	4	-1	-1	-4	-3	-3	-2	-3	-5	-3	-1	-4	-2	-1	-5	-3	11	1	
Y	0	-2	-4	-4	-2	0	3	-3	-3	-2	-3	1	-2	0	0	-1	2	1	-3	-4	-2	8	11	1	
F	-1	0	-5	-1	3	0	0	6	-5	0	-5	-3	0	-3	2	0	-2	-1	-4	-4	-5	1	11	1	
H	-1	0	-3	-3	0	-3	-1	-4	1	1	2	-1	-3	0	-2	0	2	0	1	-5	-3	-2	11	1	
L	-1	-3	-3	-3	0	0	0	-4	4	-4	-4	2	1	-4	-3	-4	0	0	2	-4	-2	-4	11	1	
I	0	-4	-3	-3	-4	0	-4	3	4	-2	2	1	-4	-4	-3	2	-3	-3	0	8	-4	3	11	1	
W	-3	-3	-4	-4	-2	-4	-4	-4	-2	-3	0	-2	-4	-4	-3	2	-3	0	3	-5	-4	-3	11	1	
P	-2	-4	-4	-4	-3	-3	-4	-4	-1	-3	-1	-2	-4	-4	-3	7	-3	-4	-3	-5	-4	-3	11	1	
D	-2	1	-4	-4	4	0	4	-3	-5	1	-5	-4	1	1	-2	1	-2	-1	-4	-4	-5	-4	11	1	
T	0	-2	-2	-2	1	-2	-3	-3	3	-2	0	-1	-2	-3	0	-3	1	4	0	-4	-4	-3	11	1	
1200	-3	-3	-4	-2	0	-2	3	4	-2	-2	-2	4	-3	-3	-2	5	-2	-3	-2	-2	-4	3	11	1	
P	3	-3	-2	-3	-2	0	-3	-3	-2	-2	1	-1	-3	-3	-2	5	-2	2	-2	-4	-3	-2	11	1	
P	0	-2	-3	-3	-2	0	-3	5	-3	4	-3	-2	-1	-1	0	-1	-2	1	-3	-4	-4	-1	11	1	
H	-2	0	-4	-4	0	5	-3	-3	-1	2	0	-1	2	-2	-1	-2	2	-2	-2	-4	-2	2	11	1	
E	-2	-5	-4	-4	-3	-4	0	-4	-2	-2	-3	-1	-4	-4	-3	-4	-1	-2	-4	-5	-2	-4	11	1	
L	-2	-5	-2	-2	-5	-4	0	-5	-4	-4	0	1	-4	-4	-3	-4	-4	-4	0	-3	-2	-4	11	1	

Figure 5/5

ATAT1	Y - - T Y E M L G I H L T T I R G H R - P P P P S P G T L G N L Y V L N H R T A L D D P	10	30	40
ATAT9	N Y K L T G I K L V V N G H P - P P P P A A G K S G V L F V C N H R T V L D D P			
ATAT7	- - S - Q I F G G H I I V V K G K P - P Q P P A A G K S G V L F V C N H R T L L M D P			
ATAT8	F S G C R L T V T N D Y V V N N H N - D L I S A D R K K R G C L F V C N H R T L L M D P			
ATAT6	A F S G I H L T T L T - - V N N A - - E R V R Q L A H D G H E L L F V C N H R S H I D D Y			
PLSB_ECOLI	N R L Y Q G I N V H N A - - E R V R Q L A H D G H E L L F V C N H R S H I D D Y			
PLSB_MOUSE	S - F F W N I Q I H K G Q L E M V K A A T E T N L P L E H A L L V I S N H R S D I D D W			
ATLPAAT1	I V D W W A G V K I Q V F A D N E T F - - R L M G K E H A L L V I S N H R S D I D D W			
Jojoba AT	V - D W W A S V K I K L F T D P D T F - - R L M G K E H A L L V I S N H R S D I D D W			
Maize AT	V - D W W A G V K V Q L H A D E E T Y - - R S M G K E H A L L V I S N H R S D I D D W			
ATAT11	L - - W P F L F E K I N K T K V I F S G D K V P C E D R V L L I A N H R T E V D D W			
PLSC_COCO	V T G R M L M W I L G N P T T - - I E G S E H T K K R A I Y I S N H A S P L D A			
PLSC_LIM	I I G G L V I W I Y G T P I K - - I Q G S E H T K K R A I Y I S N H A S P L D A			
PLSC_ECOLI	F - G R L A P L - F G L K V E C R K P T D A E S Y G N A I Y I S N H A S P L D A			
PLSC_YEAST	F - Y H V M K L M L G L D V - - K V V G E E N L A K K P A I Y I S N H A S P L D A			
ATAT2	W A S I S I Y P F Y K I N I E - - G L E N L P S S D T P A V Y V S N H V S Y L D I			
ATAT3	M D S N P K T T S T E - - I I R R - K G K P A R - - R E I A P T I V V S N H V S Y L D I			
ATAT10	R C I L F S F G - Y Q W - - T G V V K Y H G P R P S I R P K Q V Y V A N H T S M I			
ATAT4	M I C S F F V A S - - W - - T G V V K Y H G P R P S I R P K Q V Y V A N H T S M I			
ATAT1	I V A I A L G R K - I C C V T T Y S V S R L S L S P I - - P A V A A	50	60	70
ATAT9	V V T A V A L G R R K - I S C V T T Y S S I S R L S L S P I - - P A V A A			
ATAT7	V V L S Y A F A L R K K K N I K A P Y I A S G N N L N T P V - - K A T T V R R			
ATAT8	L Y V A F A L R K K K N I K A P Y I A S G N N L N T P V - - K A T T V R R			
ATAT6	L L L S Y Y I L F C H N I K A P Y I A S G N N L N T P V - - K A T T V R R			
PLSB_ECOLI	L L L S Y Y I L F C H N I K A P Y I A S G N N L N T P V - - K A T T V R R			
PLSB_MOUSE	L V G W I L A Q R S S G C L G S T L A V M K K S S K K F L P P V I - - F A V A A			
ATLPAAT1	L V G W I L A Q R S S G C L G S T L A V M K K S S K K F L P P V I - - F A V A A			
Jojoba AT	L I G W I L A Q R S S G C L G S T L A V M K K S S K K F L P P V I - - F A V A A			
Maize AT	M Y F W D L A L R K K G - - T V T V G V A K K E I I W Y P P L F F F			
ATAT11	F L I M W L A P P T G - - T V T V G V A K K E I I W Y P P L F F F			
PLSC_COCO	F F V M W L A P P T G - - T V T V G V A K K E I I W Y P P L F F F			
PLSC_LIM	V T A S N I V Q P P - - T V T V G V A K K E I I W Y P P L F F F			
PLSC_ECOLI	F M L G R I F P P G - - T V T V G V A K K E I I W Y P P L F F F			
PLSC_YEAST	Y T L L S L - G K S - - F K F T S K T G I F V I P I V - - G L I S K C L			
ATAT2	L Y H M S A S F P S - - F K F T S K T G I F V I P I V - - G L I S K C L			
ATAT3	I F Y F Y E L S P T - - F K F T S K T G I F V I P I V - - G L I S K C L			
ATAT10	I V L E Q M T A F A - - F K F T S K T G I F V I P I V - - G L I S K C L			
ATAT4	I V L E Q M T A F A - - F K F T S K T G I F V I P I V - - G L I S K C L			

Figure 2
1/3

	90	100	110	120	
ATAT1	L	T	R	R	C
ATAT9	L	T	R	R	C
ATAT7	L	T	R	R	C
ATAT8	L	T	R	R	C
ATAT6	L	T	R	R	C
PLSB_ECOLI	L	T	R	R	C
PLSB_MOUSE	L	T	R	R	C
ATLPAAT1	L	T	R	R	C
Jojoba AT	L	T	R	R	C
Maize AT	L	T	R	R	C
ATAT11	L	T	R	R	C
PLSC_COCO	L	T	R	R	C
PLSC_LIM	L	T	R	R	C
PLSC_ECOLI	L	T	R	R	C
PLSC_YEAST	L	T	R	R	C
ATAT2	L	T	R	R	C
ATAT3	L	T	R	R	C
ATAT10	L	T	R	R	C
ATAT4	L	T	R	R	C

[illegible]

Figure 2
2/3

[illegible]

Figure 2
3/3

9/10

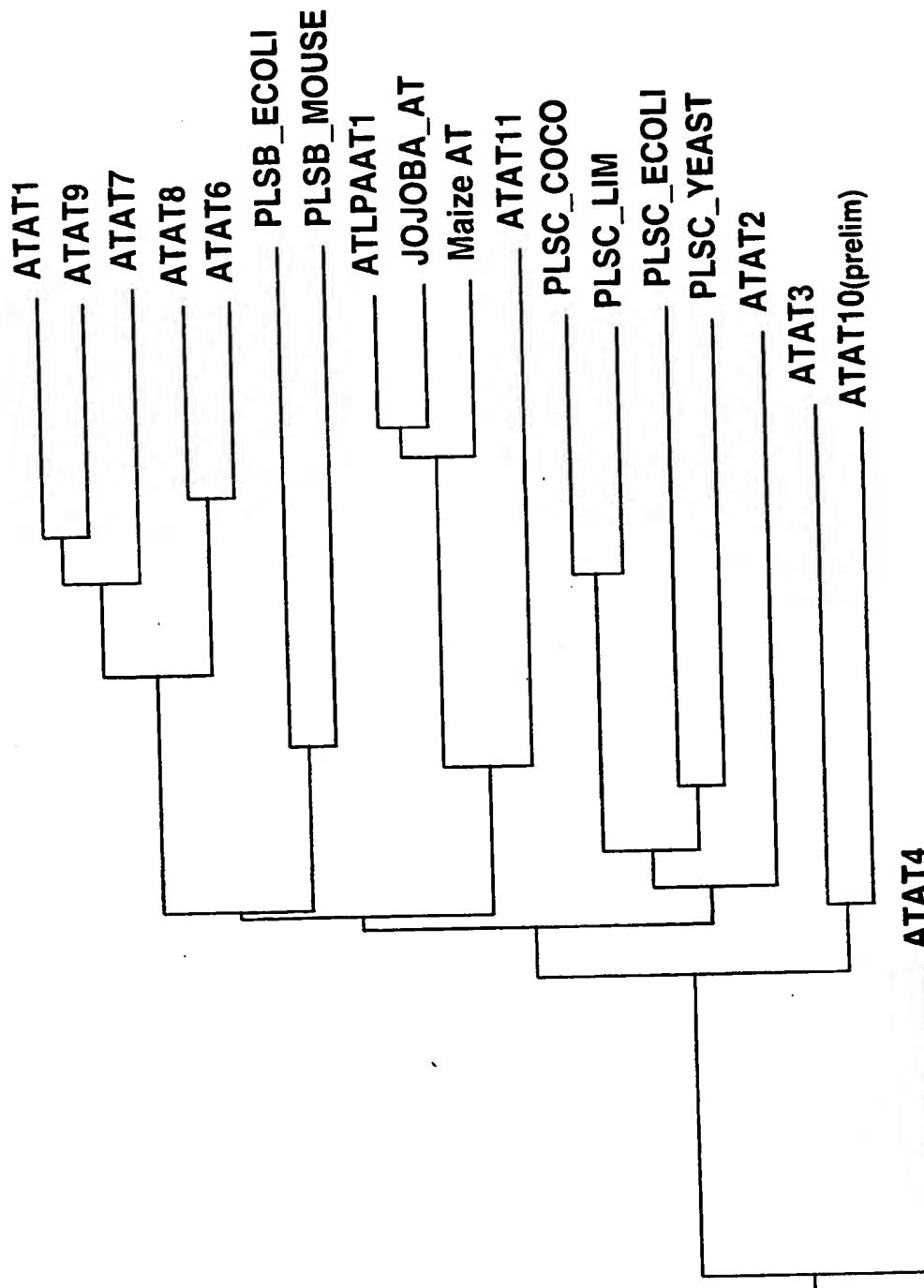


Figure 3 1/2

10/10

Percent Similarity

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2	31.4		63.9	44.8	42.8	12.9	12.4	14.9	14.4	13.9	16.5	11.3	12.4	12.4	11.3	11.2	13.4	13.7	14.4
3	40.2	35.8		44.8	44.8	12.9	14.4	14.9	13.4	11.3	12.9	12.4	12.9	11.9	13.9	13.4	13.4	17.1	14.4
4	49.7	50.0	50.3		67.2	10.8	13.3	11.8	11.8	10.8	16.4	11.8	11.8	12.8	13.3	12.3	17.4	15.1	12.8
5	50.3	50.3	48.6	25.7		12.3	12.3	13.8	12.8	12.3	12.3	12.3	12.8	12.8	12.3	12.8	15.9	13.7	15.9
6	85.6	86.3	85.6	86.2	86.1		28.5	12.6	12.1	11.6	9.7	13.9	14.3	14.8	11.8	17.6	13.5	12.3	10.6
7	83.8	86.8	82.8	82.7	84.3	66.2		12.6	13.9	12.9	13.1	12.4	13.3	14.3	13.8	15.0	11.7	11.6	10.0
8	82.9	78.4	81.2	83.1	81.2	83.6	85.1		82.3	78.0	31.6	12.4	12.8	13.3	15.8	13.9	12.2	16.4	14.4
9	83.5	77.8	81.8	85.9	84.6	85.6	87.1	18.2		77.5	32.1	11.9	14.3	13.3	16.3	15.5	12.4	15.1	12.0
10	83.5	82.4	84.1	87.6	85.1	84.4	87.1	22.5	22.5		30.6	13.9	16.7	12.8	16.3	14.4	12.9	16.4	12.0
11	84.1	81.4	83.1	85.1	85.1	90.3	87.4	65.4	65.0	67.5		14.4	14.8	11.8	12.8	13.4	14.6	15.8	12.9
12	83.6	84.6	84.6	83.3	83.2	84.6	86.8	80.9	82.2	81.1	82.4		66.7	27.9	28.4	23.5	14.9	17.1	12.9
13	82.1	84.0	81.7	81.0	82.0	87.6	86.9	79.5	80.5	79.5	81.6	33.3		26.1	28.1	19.3	14.8	14.4	15.3
14	83.2	81.4	85.0	83.6	82.9	89.1	86.8	82.1	81.6	80.5	84.3	71.3	71.6		30.0	19.3	18.7	17.8	12.3
15	83.1	80.6	83.0	79.3	81.0	88.2	87.4	81.4	82.0	79.8	84.1	70.3	70.1	62.8		20.3	15.3	15.1	13.8
16	82.7	82.6	83.9	78.4	78.8	80.8	86.1	82.9	82.2	81.6	84.5	72.3	75.0	76.5	73.5		18.2	15.8	17.6
17	83.7	82.0	86.6	78.4	80.2	86.7	89.8	86.4	85.5	85.5	84.4	80.1	78.2	78.6	80.6	77.8		30.8	17.2
18	78.5	82.5	82.5	81.7	81.8	88.7	87.1	79.1	80.5	78.9	82.8	81.8	78.1	76.1	78.1	79.3	64.8		18.5
19	84.7	84.8	84.8	84.7	85.5	86.5	83.6	87.4	86.5	87.6	91.0	85.0	83.1	85.7	81.8	83.7	79.4	74.1	
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19

Percent Divergence

ATAT1
 ATAT9
 ATAT7
 ATAT8
 ATAT6
 PLSB_ECOLI
 PLSB_MOUSE
 ATLPAAAT1
 JOIOBA_AT
 Maize AT
 ATAT11
 PLSB_COCO
 PLSB_LIM
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 PLSB_YEAST
 ATAT2
 ATAT3
 ATAT10(prellim)
 ATAT4

Figure 3 2/2

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Emig, Robin A
Ruezinsky, Diane
Van Eenennaam, Alison

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<130> 17029/00/WO

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Emig, Robin A
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Leu Ser Ile Ile Arg Val Tyr Phe Asn Leu Pro Leu Pro Glu Arg Phe
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Leu Tyr Gly Thr Met Gly Leu Gly Pro Phe Pro Met Thr Glu Asn Ile
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Lys Leu Ala Val Ala Leu Val Thr Leu Val Pro Leu Arg Phe Leu Leu
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Ser Met Ser Ile Leu Leu Leu Tyr Tyr Leu Ile Cys Arg Val Phe Thr
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Leu Phe Ser Ala Pro Tyr Arg Gly Pro Glu Glu Glu Glu Asp Glu Gly
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Val Phe Gly Phe Tyr Trp Ile His Glu Ser Cys Pro Asp Arg Asp Ser
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Asp Met Asp Ser Asn Pro Lys Thr Thr Ser Thr Glu Ile Asn Gln Lys
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Gly Glu Ala Ala Thr Glu Glu Pro Glu Arg Pro Gly Ala Ile Val Ser
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Asn His Val Ser Tyr Leu Asp Ile Leu Tyr His Met Ser Ala Ser Phe
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Pro Ser Phe Val Ala Lys Arg Ser Val Gly Lys Leu Pro Leu Val Gly
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Leu Ile Ser Lys Cys Leu Gly Cys Val Tyr Val Gln Arg Glu Ala Lys
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Ser Pro Asp Phe Lys Gly Val Ser Gly Thr Val Asn Glu Arg Val Arg

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 305 310 315 320
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 Pro Val Tyr Tyr Pro Ser Gln Glu Glu Lys Asp Asp Pro Lys Leu Tyr
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 Ala Ser Asn Val Arg Lys Leu Met Ala Thr Glu Gly Asn Leu Ile Leu
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 Arg Cys Phe Lys Ser Asn Pro Pro Glu Pro Trp Asn Trp Asn Ile Tyr
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 325 330 335
 Ala Gly Leu Lys Lys Val Pro Trp Asp Gly Tyr Leu Lys Tyr Ser Arg
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      50      55      60
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Val Ile Arg Ser Leu Phe Leu Leu Val Leu Tyr Pro Phe Ile Ser Leu
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Met Ser Tyr Glu Met Gly Leu Lys Thr Met Val Met Leu Ser Phe Phe
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Pro Leu Ile Phe His Asp Gly Arg Leu Ala Val Lys Pro Thr Pro Leu
      260      265      270
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      275      280      285
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Cys Asn His Arg Thr Leu Leu Asp Pro Leu Tyr Ile Ser Tyr Ala Leu
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Arg Lys Lys Asn Met Lys Ala Val Thr Tyr Ser Leu Ser Arg Leu Ser

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Lys Asp Gly Gln Ala Met Glu Lys Leu Leu Ser Gln Gly Asp Leu Val
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Val Lys Leu Leu Asp Pro Val Ser Gly Ser Ser Ser Ser Thr Cys Arg
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Gln His Glu Ile Gly Asn Ala Leu Gly Phe Glu Cys Thr Asn Leu Thr
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Pro	Glu	Ile	Glu	Ser	Val	Ala	Arg	Ala	Val	Leu	Pro	Lys	Phe	Tyr	Met
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Arg	Val	Val	Val	Thr	Arg	Met	Pro	Arg	Val	Met	Val	Glu	Arg	Phe	Ala
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<212> PRT

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 305 310 315 320
 Lys Pro Ser Gln Arg Lys Gly Cys Leu Phe Val Cys Asn His Arg Thr
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 370 375 380
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 385 390 395 400
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 Glu Val Ser Asp Val Ile Val Pro Val Ala Val Thr Val His Val Thr
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 450 455 460
 Pro Val Ser Gly Ala Thr Cys Gln Asp Pro Asp Gly Lys Leu Lys Phe
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 Glu Val Ala Asn Asn Val Gln Ser Asp Ile Gly Lys Ala Leu Asp Phe
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<211> 1506

<212> DNA

<213> Arabidopsis sp.

<400> 16

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 35 40 45
 Leu Glu Ala Gly Ser Leu Leu Arg Ala Leu Ile Leu Leu Val Ser Val
 50 55 60
 Pro Phe Val Tyr Leu Thr Tyr Leu Thr Ile Ser Glu Thr Leu Ala Ile
 65 70 75 80
 Asn Val Phe Val Phe Ile Thr Phe Ala Gly Leu Lys Ile Arg Asp Val
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 Glu Leu Val Val Arg Ser Val Leu Pro Arg Phe Tyr Ala Glu Asp Val
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 Gly Arg Ala Thr Gly Phe Thr Arg Lys Pro Gly Ile Leu Val Gly Gln
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 Tyr Lys Arg Asp Val Val Leu Arg Glu Phe Gly Gly Leu Ala Ser Asp
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 Leu Pro Asp Leu Gly Leu Gly Asp Ser Lys Thr Asp His Asp Phe Met
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 325 330 335
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 340 345 350
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 Glu Gly Asp Leu Val Ile Cys Pro Glu Gly Thr Thr Cys Arg Glu Pro
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 405 410 415
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<212> DNA

<213> Arabidopsis sp.

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          35          40          45
Gly Phe Glu Phe Asp His Leu Asn Pro Tyr Gly Phe Leu Ser Glu Ser
          50          55          60
Glu Pro Pro Val Leu Gly Pro Thr Thr Val Asp Pro Phe Arg Asn Asn
          65          70          75          80
Thr Pro Gly Val Ser Gly Leu Tyr Glu Ala Ile Lys Leu Val Ile Cys
          85          90          95
Leu Pro Ile Ala Leu Ile Arg Leu Val Leu Phe Ala Ala Ser Leu Ala
          100          105          110
Val Gly Tyr Leu Ala Thr Lys Leu Ala Leu Ala Gly Trp Lys Asp Lys
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Glu Asn Pro Met Pro Leu Trp Arg Cys Arg Ile Met Trp Ile Thr Arg
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 Thr Ile Ile Arg Ala Met Gln Val Ile Tyr Val Asn Arg Phe Ser Gln
 210 215 220
 Thr Ser Arg Lys Asn Ala Val His Glu Ile Lys Arg Lys Ala Ser Cys
 225 230 235 240
 Asp Arg Phe Pro Arg Leu Leu Leu Phe Pro Glu Gly Thr Thr Thr Asn
 245 250 255
 Gly Lys Val Leu Ile Ser Phe Gln Leu Gly Ala Phe Ile Pro Gly Tyr
 260 265 270
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 275 280 285
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 325 330 335
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 Ala Asp Leu Met Leu Leu Asn Lys Ala Thr Glu Leu Lys Leu Glu Asn
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 Pro Ser Asn Tyr Met Val Glu Met Ala Arg Val Glu Ser Leu Phe His
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 Val Ser Ser Leu Glu Ala Thr Arg Phe Leu Asp Thr Phe Val Ser Met
 385 390 395 400
 Ile Pro Asp Ser Ser Gly Arg Val Arg Leu His Asp Phe Leu Arg Gly
 405 410 415
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 Ser Gly His Val Leu Thr Gln Pro Leu Phe Lys Gln Thr Cys Glu Leu
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 Ala Phe Ser His Cys Asp Ala Asp Gly Asp Gly Tyr Ile Thr Ile Gln
 465 470 475 480
 Glu Leu Gly Glu Ala Leu Lys Asn Thr Ile Pro Asn Leu Asn Lys Asp
 485 490 495
 Glu Ile Arg Gly Met Tyr His Leu Leu Asp Asp Asp Gln Asp Gln Arg
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535

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 50 55 60
 Trp Leu Ala Leu Trp Pro Phe Leu Phe Glu Lys Ile Asn Lys Thr Lys
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 115 120 125
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 Gln Ile Val Ser Ser Phe Lys Asp Pro Arg Asp Ala Leu Trp Leu Ala
 165 170 175

Leu Phe Pro Glu Gly Thr Asp Tyr Thr Glu Ala Lys Cys Gln Arg Ser
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 Lys Lys Phe Ala Ala Glu Asn Gly Leu Pro Ile Leu Asn Asn Val Leu
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 210 215 220
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 225 230 235 240
 Pro Ser Phe Leu Asp Asn Val Tyr Gly Ile Glu Pro Ser Glu Val His
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<212> DNA

<213> Arabidopsis sp.

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 aagttctctac actgggcaca actcttttct tcatggaaaag gtatcacgat atcggcgctt
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1170

<210> 23

<211> 389

<212> PRT

<213> Arabidopsis sp.

<400> 23

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Ile	Ser	Gly	Leu	Ala	Val	Asn	Leu	Phe	Gln	Ala	Val	Cys	Tyr	Val	Leu
			20					25					30		
Ile	Arg	Pro	Leu	Ser	Lys	Asn	Thr	Tyr	Arg	Lys	Ile	Asn	Arg	Val	Val
		35					40					45			
Ala	Glu	Thr	Leu	Trp	Leu	Glu	Leu	Val	Trp	Ile	Val	Asp	Trp	Trp	Ala
	50					55					60				
Gly	Val	Lys	Ile	Gln	Val	Phe	Ala	Asp	Asn	Glu	Thr	Phe	Asn	Arg	Met
	65				70					75					80
Gly	Lys	Glu	His	Ala	Leu	Val	Val	Cys	Asn	His	Arg	Ser	Asp	Ile	Asp
				85					90					95	
Trp	Leu	Val	Gly	Trp	Ile	Leu	Ala	Gln	Arg	Ser	Gly	Cys	Leu	Gly	Ser
			100					105					110		
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		115					120					125			
Trp	Ser	Met	Trp	Phe	Ser	Glu	Tyr	Leu	Phe	Leu	Glu	Arg	Asn	Trp	Ala
		130				135					140				
Lys	Asp	Glu	Ser	Thr	Leu	Lys	Ser	Gly	Leu	Gln	Arg	Leu	Ser	Asp	Phe
	145				150					155					160
Pro	Arg	Pro	Phe	Trp	Leu	Ala	Leu	Phe	Val	Glu	Gly	Thr	Arg	Phe	Thr
				165					170					175	
Glu	Ala	Lys	Leu	Lys	Ala	Ala	Gln	Glu	Tyr	Ala	Ala	Ser	Ser	Glu	Leu
			180				185						190		
Pro	Ile	Pro	Arg	Asn	Val	Leu	Ile	Pro	Arg	Thr	Lys	Gly	Phe	Val	Ser
		195				200						205			
Ala	Val	Ser	Asn	Met	Arg	Ser	Phe	Val	Pro	Ala	Ile	Tyr	Asp	Met	Thr
	210					215					220				
Val	Thr	Ile	Pro	Lys	Thr	Ser	Pro	Pro	Pro	Thr	Met	Leu	Arg	Leu	Phe
	225				230					235					240
Lys	Gly	Gln	Pro	Ser	Val	Val	His	Val	His	Ile	Lys	Cys	His	Ser	Met
				245				250						255	
Lys	Asp	Leu	Pro	Glu	Ser	Asp	Asp	Ala	Ile	Ala	Gln	Trp	Cys	Arg	Asp
			260					265					270		
Gln	Phe	Val	Ala	Lys	Asp	Ala	Leu	Leu	Asp	Lys	His	Ile	Ala	Ala	Asp
		275					280					285			
Thr	Phe	Pro	Gly	Gln	Gln	Glu	Gln	Asn	Ile	Gly	Arg	Pro	Ile	Lys	Ser
	290					295					300				
Leu	Ala	Val	Val	Leu	Ser	Trp	Ala	Cys	Val	Leu	Thr	Leu	Gly	Ala	Ile
	305				310					315					320
Lys	Phe	Leu	His	Trp	Ala	Gln	Leu	Phe	Ser	Ser	Trp	Lys	Gly	Ile	Thr

325 330 335
 Ile Ser Ala Leu Gly Leu Gly Ile Ile Thr Leu Cys Met Gln Ile Leu
 340 345 350
 Ile Arg Ser Ser Gln Ser Glu Arg Ser Thr Pro Ala Lys Val Val Pro
 355 360 365
 Ala Lys Pro Lys Asp Asn His His Pro Glu Ser Ser Ser Gln Thr Glu
 370 375 380
 Thr Glu Lys Glu Lys
 385

<210> 24
 <211> 269
 <212> DNA
 <213> Glycine max

<400> 24
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 ataagggtct acttcaacct ccctctccca gaacncattg tccgctacac ctacgagatg 120
 ctggtcatca acctcgctcat ccgctggccac cgccctctctc cgccttcccc cggcaccctc 180
 ggcaacctct acgtctgcaa ccaccgcacc gctctcgacc ccacgtcat cgccattgcc 240
 ctggtggcga aggtctcctg cgtcaccta 269

<210> 25
 <211> 242
 <212> DNA
 <213> Glycine max

<400> 25
 tgatcttcca cgaaggcgt ttcgtgcaga ggccagacc actgaacgt ctcatacct 60
 tcacgtggct ccccttcggc ttcacctctt ccatacataag ggtctacttc aaccttctc 120
 tcccagaacg cattgtccgc tacacctacg agatgctcgg catcaacctc gtcattccgc 180
 gccaccgcc tctctcgcct tcccccgga ccccgga cctctacgtc tgcaaccacc 240
 gc 242

<210> 26
 <211> 272
 <212> DNA
 <213> Glycine max

<400> 26
 gtttgttcaa aggccaactc ctctagcagc cctcttgacc ttcctatggt tgccaattgg 60
 catcatactc tccatnctta aggggtctacc ttaacatccc tttgcctgaa agaattgctt 120
 ggtataacta taagctatta ggaatcagag ttattgtgaa gggtagccct ccaccacctc 180
 caaagaaggg tcaaagtggg gtcctatttg tttgtaacca ccgcacagtt ttagaccctg 240
 tggttactgc agttgcactt ggaagaaaaa tt 272

<210> 27
 <211> 218
 <212> DNA
 <213> Glycine max

<400> 27
 atagcacagg aggggttacat ggtgcctccg agcaaatcag caaaggcagt cccacaggag 60
 cgtctgaaga gcagaatgat cttccacgac gggcgtttcg tgcagaggcc agacccaatg 120
 aatgccctca tcaccttcac atggctccct ttgggtttcg tctctccat cataagggtc 180
 tacttcaacc tccctctccc agaacgcacg gtcgcta 218

<210> 28
 <211> 270
 <212> DNA
 <213> Glycine max

<400> 28
 gtgcctggtg ctgtgaactg caagcagaac atgttctttg gaaccaccgt tcgtggcgctc 60
 aagttctggg acccttaact tacttcttac atgaacccta ggctgtgta cgagggttacc 120

ttaccttgat acctttgccg aggagatgtc ggttaaggct ggggggaagt cgtccattga 180
ggtggccaac cacgtggcag aaggtgctgg gggatgtgtt agggtttgag tgcaccgggt 240
tgactaggaa ggataagtat atgttggttg 270

<210> 29
<211> 252
<212> DNA
<213> Glycine max

<400> 29
catgagggtg ggtttgctca aaggccaact cctctagctg ccctcttgac cttcctatgg 60
ctgccaattg gcatcatact ctccatctta agggctacc ttaacatccc ttgccttgaa 120
agaattgttg gtacaactac aagctcttag gaatcagagt tattgtgaag ggtacccctc 180
caccgcccc aaagaagggt caaagtgggt tctatttgtt tgtaaccacc gcacagtatt 240
agaccctgtt gt 252

<210> 30
<211> 272
<212> DNA
<213> Glycine max

<400> 30
ctgggactgc cttaaactgat gcatggatct tatcaagaaa ggagcctctg tttttttctt 60
tccagagggg acacgcagta aagatggaag actaggcaca ttcaagaagg gtgctttcag 120
tggtgctgca aagacaaatg caccagtagt accaattacc cttattggaa ctgggtcaaat 180
catgcctgca ggaaaggagg gaatagtga cataggttct gtgaaagtgg ttatacataa 240
acctattgtt ggaaaggatc ctgacatgtt at 272

<210> 31
<211> 239
<212> DNA
<213> Glycine max

<400> 31
cgggaaatcaa ggtcatcaga cttcaagggt gtttcagctg ttgtcactga cagaattcga 60
gaagctcatc agaattgagtc tgctccatta atgatgttat ttccagaagg tacaaccaca 120
aatggagagt tcctccttcc attcaagact ggtgggtttt tggcaaaggc accgggtactt 180
cctgtgatat tacgatatca ttaccagaga tttagccctg cctgggattc catatctgg 239

<210> 32
<211> 242
<212> DNA
<213> Glycine max

<400> 32
gaacggcaac ggcaacagcg ttccgcatga ccgtcctctg ctgaagccgg agcctccggt 60
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tgtacggcac catgggacgc ggcgagttgc ctcccaagga gaagctcttg ctcggtttcg 180
cgttgggtcac tcttctcccc attcgagtcg ttctcgcctg caccatattg ctcttttatt 240
ac 242

<210> 33
<211> 248
<212> DNA
<213> Glycine max

<400> 33
ttcttttct ctcactctct aaaaccctaa ctctatacat ggaagggaaa nctcaaattct 60
natgactaat taattaatcc atcgatcaag catggagtcc gaactcaaag acctcaattc 120
gaagccgccc aacggcaacg gcaacagcgt tcgcgatgac cgtcctctgc tgaagccgga 180
gcctccggtc tccgccgaca gcatcgccga tatggagaag aagttcgccg cttacgtccg 240
ccgcgacg 248

<210> 34
<211> 217
<212> DNA
<213> Glycine max

<400> 34
aaaaccctaa ttctatacat ggaagggaaa tctcaaattct aatgactaat taattaatcc 60

atcgatcaag	catggagtcc	gaactcaaag	acctcaattc	gaagccgccc	aacggcaacg	120
gcaacagcgt	tcgcatgac	cgctctctgc	tgaagccgga	gcctccggtc	tccgccgaca	180
gcatcgccga	tatggagaag	aagttcgccc	cttacgt			217

<210> 35
 <211> 257
 <212> DNA
 <213> Glycine max

<400> 35						
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aaatctaata	actaattaat	caatcaatcg	tattaataat	ccatcgatca	agtatggagt	120
ccgaactcaa	agacctcaat	tcgaagccac	ccaactgcaa	cggcaacgcc	aacagcgttt	180
gcgacgaccg	tcctctgctg	aagccggagc	ctccggcctc	ctccgacagc	atcgccgaga	240
tggagaagaa	gttcgcc					257

<210> 36
 <211> 284
 <212> DNA
 <213> Glycine max

<400> 36						
cccgaccaaa	acaggttttt	gtggccaatc	atacttccat	gattgatttc	attatcttag	60
aacagatgac	tgcatttgct	gttattatgc	agaagcatcc	tggatgggtt	ggattattgc	120
agagcaccat	tntggagagt	gtaggggtgta	tctgggtcaa	ccgtacagag	gcaaaggatc	180
gagaagttgt	ggcaaggaaa	ttgagggatc	atgtcctggg	agctaacaac	aaccctcttc	240
ttatatttcc	tgaaggaact	tgtgtaaata	atcactactc	gtca		284

<210> 37
 <211> 246
 <212> DNA
 <213> Glycine max

<400> 37						
ggagatccgc	ataagcaaata	caatcactct	gttccttctc	tatctctgtc	tctgcatttc	60
cctccctaaa	accctaattc	tacatttggg	aaggaantct	caaataat	gataattaat	120
caatcaatcg	tattaataat	ccatcgatca	agtatggagt	ccgaactcaa	agacctcaat	180
tcgaagccac	ccaactgcaa	cggcaacgcc	aacagcgttt	gcgacgaccg	tcctctgctg	240
aagccg						246

<210> 38
 <211> 278
 <212> DNA
 <213> Glycine max

<400> 38						
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cgctcagttc	tgaattggac	cttcacattg	aagattacct	accttctgga	tccagtgttc	120
aacaagaacg	gcatggcaag	ctccgactgt	gtgatttgct	agacatttct	cctagtctat	180
ctgaggcagc	acgtgccatt	gtagatgata	cattcacaag	gtgcttcaag	caaatacctc	240
agaaccttgg	aactggaatg	tttatttggg	tcctttgt			278

<210> 39
 <211> 312
 <212> DNA
 <213> Glycine max

<400> 39						
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cagaggtctt	tggtaganat	gatgtgcagt	ttctgtgggt	catcttggac	tgnggntggt	120
aagnatcatg	gacccaggcc	tagcaggaga	ccaaagcagg	tttttgtagc	caaccatact	180
tcattgattga	tntcattatn	tnagaacaga	tgactgcttt	tgcngttatn	atgcagaagc	240
atcctggatg	ggttggtaag	cntacagnat	gtcaacngtg	tatnaaatat	gntacacnnn	300
acttgcgtct	tc					312

<210> 40
 <211> 255
 <212> DNA
 <213> Glycine max

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<400> 40
ggattattgn ngcanatgca gtcattctgtt ctaagataat ganatcnatc atggaagtat 60
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naccacagtc catgatgcaa canaganact gnacatcatc tccaccaaac ccctctgana 180
ganacgagaa ttgagcaatt tagagtacct tggtttgatg caagtcagta tattcaagtt 240
tctattcatc aaagg                                     255

```

```

<210> 41
<211> 291
<212> DNA
<213> Glycine max

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```

<400> 41
caacctccca tgcaategct caccctctcc gtcacctgaa tctgttttct attccctccg 60
tcgcgtaaca aggatgaatg gcattgggaa actcaaatcg tcgagttctg aattggacct 120
tcacattgaa gattacctgc cttctggatc cagtgttcaa caagaacggc atggcaagct 180
ccgcctgtgt gatttgctag acatttctcc tagtctatct gaggcagcac gtgccattgt 240
agatgataca ttcacaaggt gcttcaagtc aaatcctcca gaaccttgga a 291

```

```

<210> 42
<211> 284
<212> DNA
<213> Glycine max

```

```

<400> 42
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gtaacgaaga tgaatggcat tgggaaactc aaatcgctcg gttctgaatt ggaccttcac 120
attgaagatt acctaccttc tggatccagt gttcaacaag aacggcatgg caagctccga 180
ctgtgtgatt tgctagacat ttctcctagt ctatctgagg cagcacgtgc catgtagatg 240
atacatcaca aggtgctcaa gtcaaatctc cagaaccttg gaat 284

```

```

<210> 43
<211> 268
<212> DNA
<213> Glycine max

```

```

<400> 43
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tcagtgtctgc ggcgatggga ggaaaagtga tgtgtacctt tatgtgggtg tgttcttaat 120
tattcttagt aatgccattg cttcgacccc tttttttgct tttgttttgc cattgctaac 180
tatttatttt taacactttt attaaagata tggcatatat ncacttcagt anacaaagtt 240
gtncacagtaa tttnttttcc aaaaaaaaaa 268

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```

<210> 44
<211> 241
<212> DNA
<213> Glycine max

```

```

<400> 44
gancaaaaatt gccctccatc acttttccttg ttagagttgg tttctgenac ctaccatgca 60
attccctcac ctgaatccgt tttctattgc caggttgtgg aagcgtaacg aagatgaatg 120
gcattgggaa actcaaatcg tcgagttctg aattggacct tcacattgaa gattacctac 180
cttctggatc cagtgttcaa caagaacggc atggcaagct ccgactgtgt gatttgctag 240
a 241

```

```

<210> 45
<211> 247
<212> DNA
<213> Glycine max

```

```

<400> 45
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aggatgcgaa aatgatgaaa aatttgctgg ggcaagggga cctgggtggt tgtcctgaag 120
ggaccacatg tagagaacct tatttattga gggtcagccc tctgttctca gagatgtgcg 180
atgagattgt ccccgttggc agttgattcc cagttatatg ttccacggaa ccaactgtcg 240
tganta 247

```

```

<210> 46
<211> 271
<212> DNA

```

<213> Glycine max

<400> 46

tgcagggggg	cttgtagag	ccatagtttt	ggttcttcta	tacccttttg	tttgtgtcgt	60
aggaaaagag	atgggggttga	agataatggt	catggcatgc	ttcttcggga	tcaaagcatc	120
gagcttcaga	gttggaaggt	ccgttttgcc	cnaattcttc	tnggaggacg	ttngtgcaga	180
aatgttttag	gcactcaaaa	aaggaggga	gacagtggga	gttaccaatt	tacccacgt	240
gatggtggaa	agcttcttga	gagagtattt	g			271

<210> 47

<211> 242

<212> DNA

<213> Glycine max

<400> 47

ttcacagctg	tcacgccgtn	aacggaaaat	ggcaacggcg	agacgcagtt	tccgcctat	60
caccgaatgc	aacggaaacga	cncctgcga	ntctgtngnc	gccgacctcg	agggtacgt	120
cctcatctcc	cgtngctcgt	tcccgtactt	catgctcgtc	gccgtcgaag	ccggcagct	180
cctccgcggc	ctcatgctnc	tcctctccct	tccgttcgtc	atnatcgct	acctcttcat	240
ct						242

<210> 48

<211> 244

<212> DNA

<213> Glycine max

<400> 48

acatattctt	cagttagctc	ccccaaccta	tacacttcac	caccacacca	caaccctacc	60
ctctctctct	gtcatggtea	ttggaggagc	cttccctcgt	ttcgacccaa	tcaccaaagt	120
tagaccaag	accgtctcaa	ccagaccatc	gcctcggacc	tcgatggcac	cctccttgtc	180
tcccggagtg	ccttccccta	ctacttctc	gtcgccctcg	aagccggcag	cgtcttccga	240
gcct						244

<210> 49

<211> 230

<212> DNA

<213> Glycine max

<400> 49

caacattcca	cctagctccc	caatcacatc	ttcaccacac	cataaacctt	cttaatttct	60
ctcttcattt	tctcctctat	tgctcataatc	atgggggacct	tcctctcgct	cgacccaatc	120
accaccaag	accggtccaa	ccagaccgtg	gcctccgacc	ttgacggcac	cctcctcgtc	180
tcccggagcg	ccttccccta	ctacctctc	gttgcccctg	aagccggcag		230

<210> 50

<211> 265

<212> DNA

<213> Glycine max

<400> 50

ctggtgaata	atcctaagtt	atggagtctg	tggtgtgtga	gctagaaggc	acgcttgtga	60
aggacaagga	tgcgttctca	tacttcatgt	tggttgcggt	tgaagcttca	ggtttggttc	120
gtttcgcctt	gttgctaaca	ctattgcccg	tgattcggtt	ccttgacatg	gttggcatga	180
acgatgcac	tctcaagcta	ntnatcttcg	tggtgtgtgg	tggtgttcca	aagtcagaga	240
ttgaatcagt	ggctagggca	gtttt				265

<210> 51

<211> 252

<212> DNA

<213> Glycine max

<400> 51

ctggtgaata	atcctaagtt	atggagtctg	tggtgtgtga	gctagaaggc	acgcttgtga	60
aggacaagga	tgcgttctca	tacttcatgt	tggttgcggt	tgaagcttca	ggtttggttc	120
gtttcgcctt	gttgctaaca	ctattgcccg	tgattcggtt	ccttgacatg	gttggcatga	180
acgatgcac	tctcaagcta	atgatcttcg	tggctgtggc	tgggttccaa	agtcagagat	240
tgaatcagt	gc					252

<210> 52

<211> 218

<212> DNA

<213> Glycine max

<400> 52

aactgcaact	acaacaacat	tcattcattc	acagctgtca	cgccgtgaac	ggaaaatggc	60
aacggcgaga	cgagttttac	ccgcctatac	accgaatgca	acggaacgac	accgtgcgag	120
tctgtggccg	ccgacctcga	cggtacgctc	ctcatntccc	gtagctcgtt	cccgtacttc	180
atgctcgtcg	ccgtcgaagc	cggcagcctc	ctccgcgg			218

<210> 53

<211> 262

<212> DNA

<213> Glycine max

<400> 53

ggttaaggac	attgagatgg	tcgnntcctc	ggtgctgccc	aagttctaca	ccgaggacgt	60
gcnccccag	agctggagag	tcttcaatcc	ttcgggaagc	gttacattgt	cactgctagt	120
ctagggatg	ggtggagcan	tttggttaaga	cgtttcttgg	ggctgataag	gtgcttggga	180
ctgagcttga	ggccacgaaa	tcggggaggt	tcatgggttt	gttaaggagc	ctgggtgtgt	240
tggtggggag	cacaagaaag	tg				262

<210> 54

<211> 212

<212> DNA

<213> Glycine max

<400> 54

gcaactacaa	caacattcat	tcattcacag	ctgtcacgcc	gtgaacggaa	aatggcaacg	60
gcgagacgca	gtttcccgcc	tatcacgaa	tgcaacggaa	cgacgccgtg	cgagtctgtg	120
gccgccgacc	tcgacggtag	gtctctcctc	tcccgtagnc	cgttcccgtg	cttcatgtct	180
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<210> 55

<211> 273

<212> DNA

<213> Glycine max

<400> 55

catggttttc	ttgagcttct	ttggcctcag	aaaggacaca	ttcagaacag	gatcagctgt	60
tctggcaaaag	ttcttcttag	aagatgttgg	attgggaaggc	tttgaggccg	taatatgttg	120
tgagagaaaa	gtggcatcta	gtaagttgcc	aagggtcatg	gttgaaaatt	tcctcaagga	180
ctatttaggg	gttgatgctg	ttatagcaag	agaattgaag	tccttttagtg	gcttcttttt	240
gggagttttt	gagagtaaga	agccaattaa	aat			273

<210> 56

<211> 257

<212> DNA

<213> Glycine max

<400> 56

ctctcaaaaa	aggagggaag	acagtgggag	tcaccaatct	accccatgtg	atgggtggaaa	60
gcttctttgag	agagtatttg	gacattgatt	tcgttgtggg	caggagagctg	aaagttttct	120
gtggatacta	cgtaggattg	atggatgaca	caaaaactat	gcatgccttg	gagctgggta	180
aagaaggaaa	aggatgctcc	gacatgatcg	gaatcacaag	gtttcgcaac	atacgcgacc	240
atgatgattt	tttctcc					257

<210> 57

<211> 240

<212> DNA

<213> Glycine max

<400> 57

gaactaagtg	tgaaccacta	ccaagaaaca	agctttttaag	tccaattatt	tttcatgagg	60
gtaggtttgc	tcaaaggcca	actcctctag	ctgnnctctt	gaccttccta	tggtgcca	120
ttggcatcat	actctccatc	ttaagggtct	accttaacat	ccctttgcct	gaaagaattg	180
cttgggtacaa	ctacaagctc	ttaggaatca	gagttattgt	gaagggtacc	cctccaccgc	240

<210> 58

<211> 254

<212> DNA

<213> Glycine max

<400> 58

cttgggaataa	gggtcattag	gaagggtatc	cctccacccc	cagcnaagaa	gggccaaagt	60
ggagtcctat	ttgtatgcaa	ccacaggaca	gttttagacc	ctgtgggttac	agctgttgca	120
ttaggaagga	aaattagctg	tgtcacatat	agcataagca	aattcactga	aataatttca	180
ccaatcaaag	ctgtggcact	ctctagggag	agggacaaag	atgctgcgaa	catcaagang	240
ttgcttgagg	aagg					254

<210> 59

<211> 267

<212> DNA

<213> Glycine max

<400> 59

gccaganaga	cttgcttggt	acaactacaa	gcttcttggg	ataagggtca	ttaggaaggg	60
tatccctcca	ccccagcaa	agaagggcc	aagtggagtc	ctatttgtat	gcaaccacag	120
gacagtttta	gaccctgtgg	ttacagctgt	tgcattagga	aggaaaatta	gctgtgtcac	180
atatagcata	agcaaattca	ctgaaataat	tcaccaatca	aagctgtggc	actctctagg	240
gagagggacc	nagatgctgc	cnacatc				267

<210> 60

<211> 261

<212> DNA

<213> Glycine max

<400> 60

gtaaccacag	ggtctaaaac	tgtgcggtgg	ttactgcagt	tgcaacttgn	nagaaaaatt	60
tgcttatgct	atatgtgaca	cagctaattc	actgnaataa	tttcaccaat	taaagctgtg	120
gcactctcaa	ggganngaga	gaaagatgct	gccaatatcc	ngagactact	tgaggaaggg	180
gacttggtga	tttgccctga	aggcacaact	tgtagagagc	cttcctcttg	aggttcagtg	240
cactatttgc	tgaactcact	g				261

<210> 61

<211> 258

<212> DNA

<213> Glycine max

<400> 61

caaggagctc	acatgcagtg	gagggaaatc	agctattgaa	gttgcaaact	acattcaaag	60
ggttcttgca	gggacttttg	gatttgagtg	cacaaatttg	actaggaaga	gcaaatatgc	120
catgcttgca	ggcacagatg	ggacagttcc	atctaaggag	aaggcttgan	aagggagaga	180
aattaagttc	tcccttttga	ttattctgta	ttggtgccca	atgtgtttcc	aaaacactta	240
gaattatgat	agaaataa					258

<210> 62

<211> 258

<212> DNA

<213> Glycine max

<400> 62

attggcataa	tcctctccat	cctaagggtc	tatctcaaca	tcctcttgcc	agaaagactt	60
gcttgntaca	actacaagct	tcttggaata	agggtcatta	ggaagggtat	ccctccaccc	120
ccagcaaaga	agggccaaag	tgagagctat	ttgtatgcaa	ccacaggaca	gttttagacc	180
ctgtgggttac	agctgttgca	ttaggaagga	aaattagctg	tgtcacatat	agcataagca	240
aattcactga	aataattt					258

<210> 63

<211> 239

<212> DNA

<213> Glycine max

<400> 63

cacttcacca	ccacaccaca	accctaccct	ctctctctgt	catggtcatt	ggaggagcct	60
tcctctgttt	cgacccaatc	accaaagtga	gcacccaaga	ccgctccaac	cagaccatcg	120
cctcggacct	cgatggcacc	ctccttgtct	cccggagtgc	cttcccctac	tacttctctg	180
tcgcccctga	agccggcagc	gtcttccgag	ccctccttct	cttaaccttc	gtccccttc	239

<210> 64

<211> 531

<212> DNA

<213> Glycine max

<400> 64

ccgagaaccg	gtctaaccac	accgtggcct	cggacttgga	cggcacccctc	ctgggtgtccc	60
ccagcgcatt	tccttactac	atgctggctc	ccatcgaagc	cggcagcttc	ctccgtggcc	120
ttgtcctcct	tgccctcgtc	cctttcgtgt	attcacgtac	atattcctct	ccgagaccgc	180
ggccatcaag	tccctgatct	tcacgcctt	cgcgggcctg	aaggtcagg	acgttgagat	240
ggtcgcgtgc	tcggtgctgc	ccaagtctta	cgcgcacata	ttcttcagtt	agctccccca	300
acctatacac	ttcaccacca	caccacaacc	ctaccctctc	tctctgtcat	ggtcattgga	360
ggagccttcc	ctcgtttcga	cccaatcacc	aaatgtagca	cccaagaccg	ctccaaccag	420
accatcgctt	cggacctcga	tggcacccctc	cctgtctccc	ggagtgcctt	ccctacttac	480
ttcctcgctg	ccctcgaagc	cggcagcgctc	ttccgagccc	tccttctctt	a	531

<210> 65

<211> 256

<212> DNA

<213> Glycine max

<400> 65

acatattctt	cagttagctc	ccccaaccta	tacacttcac	caccacacca	caaccctacc	60
ctctctctct	gtcatgggtc	ttggaggagc	cttccctcgt	ttcgacccaa	tcaccaaagt	120
tagcacccaa	gaccgctcca	accagaccat	cgcctcggac	ctcgatggca	ccctccttgt	180
ctcccgaggt	gccttccctt	actacttctt	cgctcgccctc	gaagccggca	gcgtcttccg	240
agccctcctt	ctctta					256

<210> 66

<211> 260

<212> DNA

<213> Glycine max

<400> 66

ccatccaaca	tattcttcag	ttagctcccc	caacctatac	acttcaccac	cacaccacaa	60
ccctaccctc	tctctctgtc	atgggtcattg	gaggagcctt	ccctcgtttc	gacccaatca	120
ccaaatgtag	cacccaagac	cgctccaacc	agactatcgc	ctcggacctc	gatggcacc	180
tccttgcttc	cggagtgc	ttccccctact	acttccctcgt	cgccttcgaa	gccggcagcg	240
tcttcagagc	cctccttctc					260

<210> 67

<211> 248

<212> DNA

<213> Glycine max

<400> 67

caccaacca	acctcactct	ccctttctctc	cctgaccctc	tccctgccat	ggtcattggga	60
gcctttggcc	acttcgaacc	ggctctccaaa	tgcagcaccg	agaaccggtc	taaccaaac	120
gtggcctcgg	acttggacgg	cacctctctg	gtgtccccc	gcgcatttcc	ttactacatg	180
ctgggcgcca	tcgaagccgg	cagcttctctc	cgtggccttg	tcctccttgc	ctccgtccct	240
ttcgtgta						248

<210> 68

<211> 283

<212> DNA

<213> Glycine max

<400> 68

ttcttcccca	ccatcacacc	aancaaacct	cactctncct	ggccatgggtc	atgnnnngcct	60
ttccgccact	tcgaaccggg	ttccaaatgc	agcaccgaaa	accggtttta	ccaaaccgtg	120
gcctcggact	tggacggcac	cctcctgggtg	tccccctagcg	cctttcctta	ctacatgctc	180
gtcgccatcg	aagccggcag	cttctctcgt	ggccttgctc	tccttggatc	cgcccccttc	240
gtgtacttca	cgtacatatt	cttctccgag	accgcggcca	tca		283

<210> 69

<211> 258

<212> DNA

<213> Glycine max

<400> 69

ctcttcttcc	ccaccatcnn	accaaccaaa	cctcactctc	cctgaccatg	gtcatggggag	60
cctttcgcca	cttcgaaccg	gtttccaaat	gcagcaccga	aaaccggttt	aaccaaacccg	120

tggcctcgga cttggacggc accctcctgg tgtcccctag cgcctttcct tactacatgc 180
tcgtcgccat cgaagccggc agcttcctcc gtggccttgt cctccttgga tccgtccctt 240
tcgtgtactt cacgtaca 258

<210> 70
<211> 256
<212> DNA
<213> Glycine max

<400> 70
tgcaactaca acaacattca ttcattcaca gctgtcacgc cgtgaacgga aaatggcaac 60
ggcgagacgc agtttccgc ctatcaccga atgcaacgga acgacaccgt gcgagtctgt 120
ggcgcgcgac ctcgacggta cgctcctcat ctcccgtagc tcgttcccggt acttcatgct 180
cgtcgcccgc gaagccggca gcntcctccg cggcctcatc ctctcctng ccantccgtt 240
cgctcatcanc gcctac 256

<210> 71
<211> 259
<212> DNA
<213> Glycine max

<400> 71
cttccccacc atcacaccan ggcnaacctc antctccctt tctccacnga cctctccctt 60
gccatngtca tgggancctt tggccacttc gaaccggtct ccaaatgcag caccgagaac 120
cggnctaacc aaaccgtggc ctcggacttg gacggcaccc tcttggtgtc ccncagcgca 180
tttcttact acatgctggc ngccatcgaa gccggcagct tcttccgtgg ccttgctctc 240
cttgccctcg tccctttcg 259

<210> 72
<211> 249
<212> DNA
<213> Glycine max

<400> 72
ccaacatatt cttcagttag ctcccccaac ctatacactt caccaccaca ccacaaccct 60
accctctctc tctgtcatgg tcattggagg agccttccct cgtttcgacc caatcaccaa 120
atgtagcacc caagaccgct ccaaccagac catcgctcgc gacctcgatg gcaccctnct 180
tgtctcccgg agtgccctcc cctactactt cctcgctcgc ctcgaagccg gcagcgtctt 240
ncgagccct 249

<210> 73
<211> 257
<212> DNA
<213> Glycine max

<400> 73
caaccctctt cttccccacc atcacacca ncaaacctca ctctcccttt ctccccctgac 60
cctctccctg ccatggctcat gggagccttt ggccacttcg aaccggtctc caaatgcagc 120
accgagaacc ggtctaacca aaccgtggcc tcggacttgg acggcacccct cctgggtgtcc 180
cccagcgcac ntcccttacta catgctggtc gccatcgaag ccggcagctt cctccgtggc 240
cttgctcctc ttgcctg 257

<210> 74
<211> 255
<212> DNA
<213> Glycine max

<400> 74
gccgaagacg tgcacccgga gagttggaga gtgttcaact ctttcgggaa gcgttacatt 60
gtcacggcta gtcctagggg gatggtggag ccgtttgtta aggcgtttct cggggctgac 120
aaggtgcttg ggactgaact tgaggccacc aaatcgggga cgttcaactgg gtttggttaag 180
aagcctggtg tgcttggttg ggagcataag aaagtggctc tgggtgaagga gtttcagggt 240
aattacctga cttgg 255

<210> 75
<211> 244
<212> DNA
<213> Glycine max

<400> 75

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caacaacatt cattcattca cagctgtcac gccgtgaacg gaaaatggca acggcgagac 60
gcagttttccc gcctatcacc gaatgcaacg gaacgcacac gtgcgagtct gtggccgccg 120
acctcgacgg tacgtctctc atcncccgta gctcgtttccc gtacttcattg ctcgctgccg 180
tcgaagccgg cagcctctct cgcggcctca tgcnttcctg ggtttanttt gagnaacctt 240
gagg 244

```

<210> 76
 <211> 240
 <212> DNA
 <213> Glycine max

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<400> 76
gctggctacc ctcttcttcc ccaccatcac accaatcaaa cctcactcta ccctggccat 60
ggtcattggga gcctttncgc cacttcgaac cggttttccaa atgcagcacc gaanaccgg 120
ttnacccanac cgtggcctcg gncttggacg gcaccctcct ggtgtcccct agcgcctttc 180
cttactacat gctcgtcgcc atcgaagccg gcagcttctt ccgtggcttg tctctcttgg 240

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<210> 77
 <211> 263
 <212> DNA
 <213> Glycine max

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<400> 77
gtttctcggg gctgacaagg tgcttgggac tgaacttgag gccaccaa at cggggacgtt 60
cactgggttt gttaagaagc ctgggtgtgct tgttggggag cataagaaag tggctctggt 120
gaaggagttt cagggttaatt tacctgactt gggcttaggt gatagtaaaa gtgattatga 180
cttcattgtca atttgcaagg aagggtacat ggtgccaaga actaagtgtg aaccactacc 240
aagaacaag cttttaagtc caa 263

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<210> 78
 <211> 258
 <212> DNA
 <213> Glycine max

```

<400> 78
ggccacgaaa tcggggagggt tcaactgggtt tgtaaggag cctgggtgtgc ttgttgggga 60
gcacaagaaa gtggctgttg tgaaggaggt tcagggtaat ttacctgact tgggactagg 120
agatagtaaa agtgattatg acttcattgtc aatttgcaag gaagggtaca tgggtgccaag 180
gactaagtgt gaaccactac caagaaacaa actttttaagt ccaattattt ntcattgagg 240
taggtttgtt caaaggcc 258

```

<210> 79
 <211> 260
 <212> DNA
 <213> Glycine max

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<400> 79
ctctttcttcc ccaccatcac accaancaaa cctcactctc cctttctccc ctgacctctt 60
ccctgccatg gtcattgggag ccttttgcca cttcgaaccg gtctccaaat gcagcaccga 120
gaaccggtct aaccaaaccg tggcctcgga cttggacggc accctcctgg tgccccccag 180
cgcatttctt tactacatgc tggtcgccat cgaagccggc agcttctctc gtgggccttg 240
tctctcttgc ctccgtccct 260

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<210> 80
 <211> 257
 <212> DNA
 <213> Glycine max

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<400> 80
gggaacaaca acaaatggca ngaaccttat ctcttccaa cttgggtgcat ttatccctgg 60
atacccaate cagcctgtaa ttgtacgcta tctctatgtg cactttgacc aatcctgggg 120
tcattgtntct ttgggaaagc ttatgttcag aatgttcact caatttcaca acttttttga 180
ggtagaatat cttcctgtca tttatccccct ggatgataag gaaactgctg tancttntcg 240
ggagaggact agccggg 257

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<210> 81
 <211> 272
 <212> DNA
 <213> Glycine max

<400> 81
 catacctttt gttggcacca ttattagagc aatgcaggtc atatatgtta acagattctt 60
 accatcatca aggaagcagg ctgttaggga aataaaggaa ctgaataaca gagaagggcc 120
 tcttgtgata aatttcctcg agtactatta ttcccggagg gaacaacaac taatggcagg 180
 aaccttatct ccttccaact tgggtgcatt atccctggat acccaatcca gcctgtaatt 240
 atacgctatc ctcatgtaca ctttgacca tc 272

<210> 82
 <211> 245
 <212> DNA
 <213> Glycine max

<400> 82
 gggcatttca catactagag ttcattccag tgaaaagaaa gtgggaggct gatgaatcaa 60
 tcatgcgcca tatgttttct acattcaagg atccacaaga tcctctctgg cttgcgcttt 120
 tcccagaagg cactgatttc actgagcaaa agtgccttcg gagtcaaaaa tatgtgctg 180
 aacataagtt accggttctg aaaaatgttt tacttccaag gacaaagggg cttctgtgccc 240
 gcttg 245

<210> 83
 <211> 268
 <212> DNA
 <213> Glycine max

<400> 83
 cagtgtcctt cctttctgga caatgttttt ggtgttgacc cttcagaagt gcacctgcat 60
 gtgcggcgta ttccgggtgga ggagattcca gcttctgaaa ccaaagctgc ttcttggtta 120
 atcgacacat tccagatcaa ggaccaattg ctttcggatt tcaagattca aggccatttc 180
 cctaaccaac taaatgaaaa tgaaatttct agatttaaga gcctactctc ttttatggtg 240
 atagtttctt ttactgccat gtttattt 268

<210> 84
 <211> 265
 <212> DNA
 <213> Glycine max

<400> 84
 gaaagagact gggcaaaaaga tgaaacatca ctgaagtcag gttttaggca tctagagcac 60
 atgccattcc ctttctggtt ggcccttttt gttgaaggaa ctcgtttcac gcagacaaag 120
 cttttacaag ctcaagagtt tgctgcttca aaagggtgc ctatacctag aaatgttttg 180
 attcctcgta ctaagggttt tgtcacagca gnacaaagcc ttcggccatt tcgttccagc 240
 catttatgat tgcacatatg cagtt 265

<210> 85
 <211> 265
 <212> DNA
 <213> Glycine max

<400> 85
 gaaagagact gggcaaaaaga tgaaacatca ctgaagtcag gttttaggca tctagagcac 60
 atgccattcc ctttctggtt ggcccttttt gttgaaggaa ctcgtttcac gcagacaaag 120
 cttttacaag ctcaagagtt tgctgcttca aaagggtgc ctatacctag aaatgttttg 180
 attcctcgta ctaagggttt tgtcacagca gnacaaagcc ttcggccatt tcgttccagc 240
 catttatgat tgcacatatg cagtt 265

<210> 86
 <211> 301
 <212> DNA
 <213> Zea mays

<400> 86
 ctcgctgta agggcaccac gccgcccgcg cccaagaagg gccaccggg cgctctcttc 60
 gtctgcaacc accgcaccgt gctcgacccc gtcgaggtgg ccgtggcgct gcgcccgaag 120
 gtcagctgcg tcacctacag catctccaag ttctccgagc tcatctcgcc catcaaggcc 180
 gtcgctgtgt cgcgggaggc gacaaggacg ccgagaacat ccgcccctg ctggaggagg 240
 gcgacctggt catctgcccc gagggnaaca actgccgcga gcccttctctg ctgcgttcag 300
 g 301

<210> 87
 <211> 309

<212> DNA

<213> Zea mays

<400> 87

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cgctcatgcg gtgtacatca acctgccgct gcccgagcgc atcgtctact acacctacaa 60
gctcatgggc atcaggctcg tcgtcaaggg caccgcccgc ccgcccgcga agaagggccca 120
ccggggcgct ctcttcgtct gcaaccaccg caccgtgctc gaccccgctc aggtggccgt 180
ggcgctgcgc cgcaagggtca gctgcgtcac ctacagcatc tccaagttct ccgagctcat 240
ctcgcccatc aaggccgctc cgctgtcggg gaggcgacaa ggacgccgag aacatccgcc 300
gcctgctgg 309

```

<210> 88

<211> 304

<212> DNA

<213> Zea mays

<400> 88

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tggctgtgca ggaggcctac ctggtgacgt caaggaagta cagcccggtg cccaggaacc 60
agctgtgag cccgctgatt cgtgcacgac ggccgcctcg tgcagcgcgc gacgccgctc 120
gtcgcgctcg tcaccttcct ctggatgccg ttccgcttcg cgtggcgct catgctgctg 180
tacatcaacc tgcgctgcc cgagcgcac gtctactaca cctacaagct catgggcac 240
aggctcgctg tcaagggcac ccgcccgcgc ccgcccgaaga agggccaccc gggcgctcctc 300
ttcg 304

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<210> 89

<211> 312

<212> DNA

<213> Zea mays

<400> 89

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gggtcatcca cttgtgttgc tattngaccg gtaccgtagg agagcacagc actancatcg 60
caaagatttn gggctacggt gacaatctcc atgttctaca atcttnaggt cgaaggaatg 120
gagaatctgc ctccaaatag ctgtcctggt gtctatgttg ctaaccatca gagcttcttg 180
gatatttata cccttctaac tctagggagg tgcttcaaat ttataagcaa gaccagcatc 240
tttatgttcc ctattatagg gtgggcaatg tatctcttgg gtgtgattcc tctgcggcgt 300
atggacagca gg 312

```

<210> 90

<211> 264

<212> DNA

<213> Zea mays

<400> 90

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ggtgctgtat ctgaaagaat ccacgtgct catcaacaga aaaatgcacc aatgatgcta 60
ctcttcccct gagggcacaa ctacaaatgg ggattatctc cttccattca aaacaggtgc 120
ttttcttgca aaggcaccag ttcaaccagt cattttgaga tacccttaca aaagatttaa 180
tgcagcatgg gattccatgt caggggcacg tcatgtattt ctgctgctct gtcaatttgt 240
aaattaccta gaggtggtcc gctt 264

```

<210> 91

<211> 212

<212> DNA

<213> Zea mays

<400> 91

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aaatgtcttg gatgcatttt tgttcagcgg gagtcgaaaa caccagattt caaagggtgt 60
tcagggtgctg tatttgaaag aatccatcgt gctcatcaac agaaaaatgc accaatgatg 120
ctactcttcc ctgagggcac aactacaaat ggggattatc tccttccatt caaaacaggt 180
gcttttcttg caaaggcacc agttcaacca gt 212

```

<210> 92

<211> 267

<212> DNA

<213> Zea mays

<400> 92

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gtctaaagaa atngaaaggc gtggggnaat tgtgtctaata catgtntctt atgtggatat 60
tctttatcan atgtcagcct cttttcctag tttgttgct aagagatcag tggntagatt 120
gcctctagtt ggtctcataa gcaaatgtct tggatgcatt tttgttcagc gggagtnnaa 180
aatncanatt tcaaagggtgt ttaagggtgt gnatctgaaa gaatccatcg tgctcatcaa 240

```

cagaaaaatg caccaatgat gctactc

267

<210> 93

<211> 152

<212> DNA

<213> Zea mays

<400> 93

ctacaaatgg	ggattacctt	cttccattta	agactggagc	ctttnttgca	ggtgcaccag	60
tgcagccagt	cattttgaaa	tacccttaca	ggagatttag	tccagcatgg	gattcaatgg	120
atggagcacg	tcatgtgtta	ttgctgctct	gt			152

<210> 94

<211> 274

<212> DNA

<213> Zea mays

<400> 94

aaaatataaa	ttaatatggt	cttaatccca	ccatataaat	aacgtttctct	ttctgcaggg	60
caatttagtt	ctttctaata	ttgggctggc	agagaagcgc	gtgtaccatg	cagcactgac	120
tggtagtagt	ctacctggcg	ctagacatga	gaaagatgat	tgaaagacgt	tgcgctcgctt	180
tttctgtaac	agacagccga	ggaacactta	aaaatgtaac	tgtgtgcggtg	ttttataacc	240
tgtaatgtgg	cagtttattt	gtttgaggag	gctg			274

<210> 95

<211> 295

<212> DNA

<213> Zea mays

<400> 95

aatagctatc	aagtacaata	aaatatttgt	tgatgccttt	tggaacagta	agaagcaatc	60
ttttacaatg	cacttggtcc	ggctgatgac	atcatgggct	gttgtgtgtg	atgtttggta	120
cttacctcct	caatatctga	gggagggaga	gacggcaatt	gcatttgctg	agagagtaag	180
ggacatgata	gctgctagag	ctggactaaa	gaaggttcct	tgggatggct	atctgaaaca	240
caaccgtcct	agtcccaaac	acactgaaga	gaacaacgca	tattgccgat	ctgtc	295

<210> 96

<211> 273

<212> DNA

<213> Zea mays

<400> 96

gngccatctc	accggcggn	ggcctgcggc	cggcaaccgg	aggcgatggc	gagctngtct	60
gtgggtggcg	acatggagca	ntaccgcccc	aacctggagg	actacctccc	gcccgaactcg	120
ctcccgagg	aggcgcccag	gaatctccat	ctgcgcgatc	tgcttgacat	ctcgccggtg	180
ctaaccgagg	cagcgggtgc	catagtcgat	gattcattca	cccgttgctt	taagtcgaat	240
tctccagaac	catggaatgg	aacatatatt	tgt			273

<210> 97

<211> 127

<212> DNA

<213> Zea mays

<400> 97

ctcaatatct	ganggagga	gagactgcaa	ttgcgtttgc	tgagagagta	agggacatga	60
tagcagctag	agctggtctt	aagaaggctc	cgtgggatgg	ctatctgaag	cacaaccgcc	120
ctagtcc						127

<210> 98

<211> 286

<212> DNA

<213> Zea mays

<400> 98

gaaccgtacg	cgccctatta	cgccccatcca	cgtgctcgcc	tctccccatc	gcataatttt	60
nectggcggc	gtcgccatct	ccanccggcng	cnggcctgcn	gccggcaacc	ggaggcgatg	120
gcgagctcgt	ctgtggcggc	ggacatggag	ctggaccgcc	ccaacctgga	ggactacntc	180
ccgccccgant	cgctccccga	ggaggcgacc	aggaatctcc	atctgngcga	tctgcttgan	240
atctcgccgg	tgctaaccga	ggcagcgggt	gccatagtcg	atgatt		286

<210> 99
 <211> 308
 <212> DNA
 <213> Zea mays

<400> 99
 cgccatctca tcggcggcgg gcgtgcggcc ggccggcngag gcgaggngcg attggcgagc 60
 tcgtctgtgg cgccggacat ggagctggac cgcccanacc tggaggacta nctcccgccc 120
 gactcgnncc cgcagaggcg ccccggaatc tccanctgcg cgatctgctg gacatcncgc 180
 cggtgctcac cgaggcagcg ggtgccattg tcgatgactc cttcacacgg ngctttaagt 240
 caaattctcc agagccatgg aattggaaca tatatctgtt ccccttatgt gctttggtgt 300
 ataataag 308

<210> 100
 <211> 282
 <212> DNA
 <213> Zea mays

<400> 100
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 gagcaactat gcaattttaat gccatgctgt gactaacttc tagtttctgg cattaaatta 120
 ctgtttggct actaggaaga ccgaggtaga gaagcaaata taagaatacc ctccaacgca 180
 canccaaatg acagagtaaa tgaaggtagg gttcaccttc ttgaacatga ccgtatactg 240
 gttgttaaca caagttcctc tgggaaaatc agagagggtt tt 282

<210> 101
 <211> 282
 <212> DNA
 <213> Zea mays

<400> 101
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 acnggcattgt cgtggcggct caaagggtn gcccngngc ttgcnnngcc gtgctccggc 120
 gggcgctgnc agctgttcgt gtgcaacnac cggacgctga tcgaccngt gtacgtgtcc 180
 gtagcgtgga ccgggaaatg cgcgncgtgt nctacagnct gangcggntn tcggagctca 240
 tctcccccat ngncggaang tgcacctgan accgggaacg gg 282

<210> 102
 <211> 290
 <212> DNA
 <213> Zea mays

<400> 102
 ggacgcggca ccatgcgcgc cgagctggcc agtggcgacg tggccgtgtg ccccgagggc 60
 accacgtgcc gggagccctt cctgctccgc ttctccaagc tcttcgcgga gctcagcgac 120
 aggatcgtgc ccgtggcgat gaactaccgc gtggggctct tccacccgac gacggcgcg 180
 ggggtggaaag ccatggaccc catcttcttc ttcatgaacn gcggcccgtg tacgaggtga 240
 cgttcctgaa ccantccccg caaagcgacg tgcgcggcgg ggaagagccc 290

<210> 103
 <211> 279
 <212> DNA
 <213> Zea mays

<400> 103
 acgaggtgac gttcctgaac cagctccccg cagaggcgac gtgcgcggcg gggaagagcc 60
 ccgttgatgt agccaactac gttcagcgga tactcgctgc cacgctcggg ttcgagtga 120
 ccaccctcac aaggaaggac aaatacacgg tgctcgccgg caacgacggc gtcctgaacg 180
 ccaagccggc ggccggcccg aagccggctt ggcagagccg cgtgaaggaa gtcctcgggt 240
 tctgctccac taacaattac accttgccca gatctggac 279

<210> 104
 <211> 315
 <212> DNA
 <213> Zea mays

<400> 104
 gcccagagcgc atcgtctact acacctacaa gctcatgggc atcaggctcg tcgtcaaggg 60
 caccgcccg ccgccgccca agaagggcca cccgggcgtc ctcttcgtct gcaaccaccg 120
 caccgtgctc gaccccgctc aggtggccgt ggcgctgcgc cgcaangtca gctgcgtcac 180

tacagcatct ccaagttctc cgagctcatc tgcgccatca aggccgtagc agnaaagcag 240
gtcgcaaattg gagcagnagc gagtcgatgg aagngaattg gcgactgggc atctgcncga 300
agnnacactg cggag 315

<210> 105

<211> 314

<212> DNA

<213> Zea mays

<400> 105

cgagacaccg agcacgtact accagcaaga tgggtggcgtc tcccagattc aagcccatcg 60
aggagtgtg ctcggagggg cggtcggagc agacgggtggc cgccgacctg gacggcacgc 120
tgctcatctc caggagcgcg tccccctact acctctcgt ggctctcgag gccggcagcg 180
tctccgcgc cgcgctgctg ctctgttcg tgccgttcgt ctacgtcacc tacgccttct 240
tctccgagtc gctggccatc agcacgctgg tgtacatctc cgtggcgggg ctgaagggtgc 300
gcanatcgag atgg 314

<210> 106

<211> 291

<212> DNA

<213> Zea mays

<400> 106

ctctgggtct ggggccgaga caccgagcac gtactaccag caagatgggtg gcgtctccca 60
gattcaagcc catcgaggag tgctgtctcg aggggcggtc ggagcagacg gtggccgccc 120
acctggacgg cagctgtctc atntccagga gcgcgttccc ctactacctc ctctgtggctc 180
tcgaggccgg cagcgtcttc cgcgcgcgc tgctgtctct gtccgtgccc ttcgtctacg 240
tcacctagc cttcttctcc gagtcgctgg ccacacgac gctggtgtac a 291

<210> 107

<211> 300

<212> DNA

<213> Zea mays

<400> 107

gcacgcagca gtacgacgtc tctctcttgg gtctggggcc gagacaccga gcacgtacta 60
ccagcaagat ggtggcgtct cccagattca agcccatcga ggagtgtgc tcggaggggc 120
ggtcggagca gacgggtggc gccgacctgg acggcacgct gctcatctcc aggagcgcgt 180
tcccctacta cctctctgtg gtctctgagg ccggcagcgt cctccgcgcc gcgtgtctgc 240
tctgttcgt gccgttcgtc tacgtcacct acgccttctt ctccgagtcg ctggccatca 300

<210> 108

<211> 284

<212> DNA

<213> Zea mays

<400> 108

gnggccgaga caccgagcac gtactaccag cagatgggtg gcgtctccca gattcangcc 60
antcgaggag tgctgtctcg aggggcggtc ggagcagacg gtggccgccc acctggacgg 120
cagctgtctc atctccagga gcgcgttccc ctacnacctc ctctgtggctc tcgaggccgg 180
cagcgtcttc cgcgcgcgc tgctgtctct gtccgtgccc ttcgtctacg tcaactacgc 240
ttcttctccg agtcgctggc catcaanacg ctggtgtaca tctc 284

<210> 109

<211> 280

<212> DNA

<213> Zea mays

<400> 109

ctctcttggg tctggggccg agacaccgag cacgtactac cagcaagatg gtggcgtctc 60
ccagattcaa gccatcgag gagtgctgct cggaggggcg gtcggagcag acgggtggccg 120
ccgacctgga cggcacgctg ctcatctcca ggagcgcgtt ccnctactac ctctctgtgg 180
ctctcgaggc cggcagcgtc ctccgcgcgg cgtgtgtgt cctgtccgtn ccgttcgtct 240
acgtcaccta cgenttnttc tccgagtcgc tggccatcag 280

<210> 110

<211> 287

<212> DNA

<213> Zea mays

<400> 110
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 gtctcccaga ttcaagccca tcgaggagtg ctgctcggag gggcggtcgg agcagacggg 120
 ggccgcccag ctggacggca gctgctcatc tccaggagcg cgttccccta ctacctctc 180
 gtggctctcg aggcggcag cgtcctccgc gccgcgtgc tgctcctgtc cgtgccgttc 240
 gtctacgtca ctacggcttc ttctccgagt cgctggccat cagcacg 287

<210> 111
 <211> 286
 <212> DNA
 <213> Zea mays

<400> 111
 cgcacagtta cgacgtctct cctctgggtc tggggccgag acaccgagca cgtactacca 60
 gcaagatggg ggcgtctccc agattcaagc ccatcgagga gtgctgctcg gaggggcggg 120
 cggagcagac ggtggccgcc gacctggacg gcacgctgct catctccagg agcgcgttcc 180
 cctactactc ctctgtctct cgaggccggc aggtcctccg cgcgcgctg tgctcctgtc 240
 gtgcgttcgt ctagtcaacta cgtttttctc gancgtggca ataana 286

<210> 112
 <211> 323
 <212> DNA
 <213> Zea mays

<400> 112
 gttattccct gaaggtagca caacaaatgg gagattcctg atttcgttcc aacatgggtgc 60
 attcatacct ggctaccctg ttcaacctgt tgttggtccg tatccacatg tgcactttga 120
 tcaatcatgg gggnatatat cgttattaaa gctcatgttt aagatgttca cccaatttca 180
 taatttcacg gaggtagagt accttcctgt tgtctaccct cctgagatca agcaagagaa 240
 tgcccttcat tttgcggagg ataccagcta tgctatggca cgtgccctca atgtcttgcc 300
 aacttcctat tcatatggtg att 323

<210> 113
 <211> 312
 <212> DNA
 <213> Zea mays

<400> 113
 cgataaggcc cttttcgaag agcttctacc gtcggatcaa cagattcttg gccgagctgc 60
 tgtggcttca gcttgtctgg gtgggtggact ggtgggcagg tgttaaggta caactgcatg 120
 cagatgagga aacttacaga tcaatgggta aagagcatgc actcatcata tcaaatcatc 180
 ggagtgatat tgattggctc attggatgga tattggccca gcgttcaggg tgccttgga 240
 gtacacttgc tgtcatgaag aagtcattca agttccttc agttattggc tggtaaatgt 300
 ggtttgcaga gt 312

<210> 114
 <211> 279
 <212> DNA
 <213> Zea mays

<400> 114
 agtgggggtc ccaaagggtg aaagacttcc ctagaccatt ttggctagct ctttttgttg 60
 agggtagctc ctttactcca gcaaagcttc tcgcagctca ggagtatgcg gcttcccagg 120
 gcttaccagc tcctagaaat gtacttattc cacgtaccaa gggatttgta tctgccgtaa 180
 gtattatgcg agattttgtt ccagccattt acgatacaac tgtaatatgt cctaaagatt 240
 cccctcaacc aacaatgctg cggattttga aagggaat 279

<210> 115
 <211> 304
 <212> DNA
 <213> Zea mays

<400> 115
 cgtcaacgcc atccaggccg tcctatttgt gacgataagg cccttttcga agagcttcta 60
 ccgtcggatc aacagattct tggccgagct gctgtggctt cagcttgtct ggggtggtgga 120
 ctggtgggca ggtgttaagg tacaactgca tgcagatgag gaaacttaca gatcaatggg 180
 taaagagcat gcaatcatca tatcaaatca tcggagtgat attgattggc tcatggatgg 240
 atattggccc agcgttcagg gtgccttggg agtacattgc tgtcatgaag aagtcattca 300
 agtt 304

<210> 116
 <211> 259
 <212> DNA
 <213> Zea mays

<400> 116
 cttcctcctg tccggcctca tcgtcaacgc catccaggcc gtcctatttg tgacgataag 60
 gcccntttcg aagagcttct aacgtcggat caacagattc ntggccgagc tgctgtggct 120
 tcagcttggtc tgggtgggtg acnggtgggc aggtgttaag gtacaactgc atgcngatga 180
 ggaaacttac agatcnatgg gtanagagca tgcactcatc atatcaaate atcggagtga 240
 tattgattgg cncattgga 259

<210> 117
 <211> 235
 <212> DNA
 <213> Zea mays

<400> 117
 attccacgta ccaagggtatt tgtatctgct gtaagtatta tgcgagattt tgttccagcc 60
 atttatgata caactgtaat agttcctaaa gattcccctc aaccaacaat gctgcggatt 120
 ttgaaagggc aatcatcagt gatacatgtc cgcatgaaac gtcattgcaat gaggatgatg 180
 ccaaaatcag atgaggatgt ttcaaaatgg tgtaaagaca tttttgtggc aaagg 235

<210> 118
 <211> 282
 <212> DNA
 <213> Zea mays

<400> 118
 tgagatgcca aaatcagatg atgacgtttc aaaatgggtg aaagacattt ttgtgacaaa 60
 ggatgcctta ctggacaaac atttggcaac aggcactttc gatgaggaga ttagacctat 120
 cggccgcccc gtgaaatcat tgctgggtgac cctgttttgg tcgtgcctgc tgttgtttgg 180
 tgccatcgag ttcttcaagt ggacgcagct cctatcgaca tggagaggag tggcattcac 240
 tgccgcagga tggcgctcgt gacaggggtc atgcacgtct tc 282

<210> 119
 <211> 166
 <212> DNA
 <213> Zea mays

<400> 119
 ctgggtgggca ggcgttaagg tacaactaca tgcggatgag gacacttacc gatcaatggg 60
 taaagagcat gcactcgtca tatcaaatca tcgaagtgat attgattggc ttattgggatg 120
 gatattggcc cagcgtcag ggtgccttgg aagtaacgctc gctgtc 166

<210> 120
 <211> 234
 <212> DNA
 <213> Zea mays

<400> 120
 agtcanccaa gntccttcca gtcattggct ggtcaatgtg gtttgcagag tacctctttt 60
 nggagaggag ctgggccaag gatgaaaaga cactaaagtg ggtctccaa aggttgaaag 120
 acttccctag accatttngg ctagctcttn tttgtngagg gnantcgctt tactccagca 180
 angnttntng aggnnncagn agnnncgggn ttcccanggg ttaacagncc cana 234

<210> 121
 <211> 210
 <212> DNA
 <213> Zea mays

<400> 121
 gtgagatgcn aaaatcagat gatgacgttt caaaatgggt taaagacatt tttgtggaca 60
 aaggatgcct tactggacaa acatttggca acaggcactt tcgatgagga gattagacct 120
 atcgcccgcc cagtgaatc atngctgggt accctgtntt ggtcgtgcct gctgttgttt 180
 ggtgccatcg agntcttcaa gtggacgcag 210

<210> 122
 <211> 274
 <212> DNA

<213> Zea mays

<400> 122

acncccgaat	cgcgcgcgcg	cgcnccgtcc	tcgtcgccgg	cggaggcgcc	cgcnaccgcc	60
cacagcagcc	tatcgccgga	gaaggaacgc	cgcggggagc	ttttccacng	ccatctcccg	120
tctgaccct	ccgagatcgn	aagcggcgcc	catggcgatc	ccgctcgtgc	tcgtcgtgct	180
cccgtcggc	ctcctcttcc	tcctgtccgg	cctcatcgtc	aacaccatcc	aggccatcct	240
atthgtgaca	ataaggccct	tttccaagag	cttg			274

<210> 123

<211> 305

<212> DNA

<213> Zea mays

<400> 123

ttgcactgag	gaaaggccat	tagggatata	tcaagtacat	acataagagc	agcttgatga	60
agttgcctat	ttttagctgg	gcatttcaca	tttttgagtt	tatcccggta	gaacgggaaat	120
gggagattga	tgaagcaatt	attcagaaca	agctatcaaa	atttaagaac	ccgagagatc	180
ctatctgggt	ggcgggtttt	cctgaaggca	cggattatac	tgagaagaaa	tgcatcatga	240
gtcaagagta	tgcttcagaa	catggcttgc	ctatgctaga	acatgtcctc	cttccaaaga	300
caagg						305

<210> 124

<211> 279

<212> DNA

<213> Zea mays

<400> 124

ccagattttc	tggacaatgt	gtatggcggt	gatccttctg	aagtccacat	ccacgtcaga	60
atggttcagc	tccatcacat	ccccacaaca	gaagacaaga	taacagaatg	gatggncgag	120
aggttttaggc	agaaggacca	gctcctggca	gatttcttca	tgaaggggca	tttctgatg	180
aaaggaactg	aaaggagatc	tgctcgacgc	gagtgcctgg	caaactttct	taaccagtag	240
tatgcttgac	ggccnatctg	gtttgtacct	aaactcttt			279

<210> 125

<211> 219

<212> DNA

<213> Zea mays

<400> 125

agattttntg	gacaatgtgt	atggngttga	tccttntgaa	gtncacatcc	acgtnagaat	60
ggttcagctc	catcacatcc	ccacaacagn	agacaagata	acagaangga	tggttagagag	120
gttttaggcag	aaggaccagc	tcctggcaga	tttcttcatg	aaggggcact	ttcctgatga	180
aggaactgaa	ggagatctgt	cgacgccgaa	gtgcctggc			219

<210> 126

<211> 293

<212> DNA

<213> Zea mays

<400> 126

taccatagat	gctgtgtacg	acatcacgat	cgcntacaaa	caccggcngc	ngacattttct	60
ngacaacgtc	tacngcgtgg	ntccttcgga	agtccacatc	cacatcanca	gcattccaggt	120
ctccgacata	ncggcgctccg	aaaaacgggg	tggtctggcng	gntnngtgga	gcggttcaag	180
gcntnganna	acgagctngc	tggtcggggc	tttctaccgc	ggctggggcc	aatttcnccc	240
cgaacgaaag	ggaaaaaggg	gaaccgaagg	ggggaacctg	ttngaacggg	ncc	293

<210> 127

<211> 6

<212> PRT

<213> conserved sequence

<400> 127

Val	Xaa	Asn	His	Xaa	Ser
1					5

<210> 128

<211> 6

<212> PRT

<213> conserved sequence

<400> 128

Val Thr Tyr Ser Xaa Ser
1 5

<210> 129

<211> 7

<212> PRT

<213> conserved sequence

<400> 129

Val Xaa Leu Thr Arg Xaa Arg
1 5

<210> 130

<211> 5

<212> PRT

<213> conserved sequence

<400> 130

Cys Pro Glu Gly Thr
1 5

<210> 131

<211> 5

<212> PRT

<213> conserved sequence

<400> 131

Ile Val Pro Val Ala
1 5

<210> 132

<211> 7

<212> PRT

<213> conserved sequence

<400> 132

Leu Xaa Xaa Gly Asp Leu Val
1 5

<210> 133

<211> 6

<212> PRT

<213> conserved sequence

<400> 133

Phe Xaa Xaa Gly Ala Phe
1 5

<210> 134

<211> 6

<212> PRT

<213> Synthetic Oligonucleotide

<400> 134

Val Ala Asn Xaa Xaa Gln
1 5

<210> 135

<211> 30

<212> DNA

<213> Synthetic Oligonucleotide
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<210> 136
<211> 31
<212> DNA
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<400> 136
tccctgtctt gcttgatgaa cttaaagctt g 31
<210> 137
<211> 30
<212> DNA
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<400> 137
acagcaggag tgtctgatga tggcagattc 30
<210> 138
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide
<400> 138
actggagttc cagccaaaaa tgcacctgtc 30
<210> 139
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<400> 139
gatacaccct tgaaatcagg cgatttttgc 30
<210> 140
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<400> 140
ttgcaaattc aattcctgtt tcaccgggccc 30
<210> 141
<211> 30
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<213> Synthetic Oligonucleotide
<400> 141
gttttctgct attccagaag gcgtcaacaa 30
<210> 142
<211> 32
<212> DNA
<213> Synthetic Oligonucleotide
<400> 142
cattgaagat ccgtccgtga agttncctta cc 32
<210> 143
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<400> 143
tcgagctgtg atcgatgatt ggctgtgaag 30
<210> 144

<211> 30
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<400> 144
gtctcttcaa aaacacacac acacgtctct 30

<210> 145
<211> 30
<212> DNA
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<400> 145
gtctcttcaa aaacacacac acacgtctct 30

<210> 146
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 146
gtagagagcc ttacttgctt cggtttagtc 30

<210> 147
<211> 30
<212> DNA
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<400> 147
acgtcatcgt acctggtgct attgactcac 30

<210> 148
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 148
acttttccat tgtcaggac tcctcgacac 30

<210> 149
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 149
acggtgtagg aagggaagg attcaaaagg 30

<210> 150
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<213> Synthetic Oligonucleotide

<400> 150
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<210> 151
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<212> DNA
<213> Synthetic Oligonucleotide

<400> 151
ccggtttacg agattacgtt cttgaaccag 30

<210> 152
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 152
caatggagac aaggctcgaa agtgctaacc 30

<210> 153
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 153
attctctgaa catagttcgc cacggtcatg 30

<210> 154
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 154
gaaatccaac gccttcccaa tatcactctg 30

<210> 155
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 155
cttcaacttt ccatcaggat cttggcacgt 30

<210> 156
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 156
accacttggt agagacctta cctgcttagg 30

<210> 157
<211> 30
<212> DNA
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<400> 157
tcctacctac accatccaat ttctcgaccc 30

<210> 158
<211> 30
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<213> Synthetic Oligonucleotide

<400> 158
ctgcgtcaag tgagcaactc agttcttgca 30

<210> 159
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 159
tggaagcag cacgttggtc agtatcgga 30

<210> 160
<211> 30
<212> DNA
<213> Synthetic Oligonucleotide

<400> 160
tagcctctgt gtaatctgtg ccctcgggga 30

<210> 161
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<212> DNA
<213> *Simmondsia chinensis*

<400> 161
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tagtataatt atatctgggt aatcttgaat ttgttggtga ggccatgggg atcccagctg 180
cggctgtgat tgtaccgctt ggcttgctct tcttcttctc tggctctctc atcaacttca 240
ttcaggcaat ttgttttgtg ctctgtgcgc cactgtcaaa gnntacatac agaaggatta 300
acagggtgct ggtggaattg ttgtggcttg agctgatatg gctcgtagat tgggtgggcaa 360
gtgttaagat caagtgtgtc acagatcctg atacctttcg gctaattgggt aaagagcatg 420
cacttgtgat atcaaaccac agaagtgata ttgattggct tgttggatgg gtgttggccc 480
agagatcagg ctgcctggga agcacactgg ctgtcatgaa gaaatcatca aagtttctcc 540
cggtcacagg ttggtctatg tggttttctg agtacctttt tcttgagaga agctggggcca 600
aggatgaaag cacattgaag ttaggtcttc aacgcctcaa ggactaccct ctgcctttct 660
ggttggctct tttcgtagaa ggaacacgat ttacccaagc taaactttta gcagctcaag 720
aatatgctac ttcaatggga ttgccagttc ctagaatac tttgatccct cgtactaagg 780
gatttgttcc agccgtgagc catatgcgtt cgtttgtccc ggccatatac gatgtaacgg 840
tggccatccc taaatcttct tcgcagccta caatgctcag acttttcaaa ggccagccat 900
ccacggttca tgtacacatc aagcgccgct cgatgaaaga tctccctgaa gcagcagatg 960
atgttgcaac atggtgtcga gacacattcg tcgcaaagga tgcactcctg gacaagcata
1020
atgtagatga cacttttcgga gatgagtatc tgcaggacac tggccggcct ttgaaatctc
1080
tctttgtagc agtctcttgg gcattgattc tcatcctggg aggtttgaaa ttcttacgat
1140
ggtcgtccct tctatcatca tgggaaggggg tcgccttctc agccgcatgc cttgtgctcg
1200
tcaccattct tatgcagatc ttaatccaat tttctcaatc cgagcgctcg actcctgcta
1260
aggtagcccc aggaaagccc aagaacatgg tatcagaacc cacggaaacg caacgacata
1320
agcagcacta aaagtatata tggaccccaa ctaagaagat tcagacgcaa gccacagttg
1380
attcaactgt tcagaatgtc aaatatagtt tgagaaacaa aagatcaaga ttagctgatg
1440
aagagcctaa tgaacctaca tacttggatc tgtcgtcgcc accgtctgct gctagctcgt
1500
tatcagaatt cgtgattccg ggaccgatcc cggatcttag ccttctatgc atggattatg
1560
atagtatctt aaatttcttt aatgatgtac cggaattata atgttagtta attaggggga
1620
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1680
aattcgattc gagtgtctctg aa
1702

<210> 162

<211> 387

<212> PRT

<213> *Simmondsia chinensis*

<400> 162

Met Gly Ile Pro Ala Ala Ala Val Ile Val Pro Leu Gly Leu Leu Phe
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Phe Phe Ser Gly Leu Phe Ile Asn Phe Ile Gln Ala Ile Cys Phe Val
20 25 30
Leu Val Arg Pro Leu Ser Lys Thr Tyr Arg Arg Ile Asn Arg Val Leu
35 40 45
Val Glu Leu Leu Trp Leu Glu Leu Ile Trp Leu Val Asp Trp Trp Ala
50 55 60
Ser Val Lys Ile Lys Leu Phe Thr Asp Pro Asp Thr Phe Arg Leu Met
65 70 75 80
Gly Lys Glu His Ala Leu Val Ile Ser Asn His Arg Ser Asp Ile Asp
85 90 95
Trp Leu Val Gly Trp Val Leu Ala Gln Arg Ser Gly Cys Leu Gly Ser
100 105 110

Thr Leu Ala Val Met Lys Lys Ser Ser Lys Phe Leu Pro Val Ile Gly
 115 120 125
 Trp Ser Met Trp Phe Ser Glu Tyr Leu Phe Leu Glu Arg Ser Trp Ala
 130 135 140
 Lys Asp Glu Ser Thr Leu Lys Leu Gly Leu Gln Arg Leu Lys Asp Tyr
 145 150 155 160
 Pro Leu Pro Phe Trp Leu Ala Leu Phe Val Glu Gly Thr Arg Phe Thr
 165 170 175
 Gln Ala Lys Leu Leu Ala Ala Gln Glu Tyr Ala Thr Ser Met Gly Leu
 180 185 190
 Pro Val Pro Arg Asn Thr Leu Ile Pro Arg Thr Lys Gly Phe Val Ser
 195 200 205
 Ala Val Ser His Met Arg Ser Phe Val Pro Ala Ile Tyr Asp Val Thr
 210 215 220
 Val Ala Ile Pro Lys Ser Ser Ser Gln Pro Thr Met Leu Arg Leu Phe
 225 230 235 240
 Lys Gly Gln Pro Ser Thr Val His Val His Ile Lys Arg Arg Ser Met
 245 250 255
 Lys Asp Leu Pro Glu Ala Ala Asp Asp Val Ala Gln Trp Cys Arg Asp
 260 265 270
 Thr Phe Val Ala Lys Asp Ala Leu Leu Asp Lys His Asn Val Asp Asp
 275 280 285
 Thr Phe Gly Asp Glu Tyr Leu Gln Asp Thr Gly Arg Pro Leu Lys Ser
 290 295 300
 Leu Phe Val Ala Val Ser Trp Ala Leu Ile Leu Ile Leu Gly Gly Leu
 305 310 315 320
 Lys Phe Leu Arg Trp Ser Ser Leu Leu Ser Ser Trp Lys Gly Val Ala
 325 330 335
 Phe Ser Ala Ala Cys Leu Val Leu Val Thr Ile Leu Met Gln Ile Leu
 340 345 350
 Ile Gln Phe Ser Gln Ser Glu Arg Ser Thr Pro Ala Lys Val Ala Pro
 355 360 365
 Gly Lys Pro Lys Asn Met Val Ser Glu Pro Thr Glu Thr Gln Arg His
 370 375 380
 Lys Gln His
 385

<210> 163

<211> 43

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 163

aagcttgcat gcgtcgacac aatggttcat gcgaccaagt cag

43

<210> 164

<211> 35

<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 164
ggatccgctcg actcacttct tgggtgttgtt gatag 35

<210> 165
<211> 44
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 165
ggatccgctcg ccgcacaatg acgagcttta ctacttcct tcat 44

<210> 166
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 166
ggatcccctg caggtagag atccattgat tctgcaat 38

<210> 167
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 167
ggatccgctcg ccgcataatg gaatcagagc tcaaagat 38

<210> 168
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 168
ggatcccctg caggtcattc ttctttctga tggaaatc 38

<210> 169
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 169
ggatccgctcg ccgcacaatg actcgttcac aagatgttc a 41

<210> 170
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 170
ggatcccctg caggtcactt ctcttccaat ctagccag 38

<210> 171
<211> 46
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 171
ggatccgcgg ccgcacaatg tccggtaata agatctcgac tcttca 46

<210> 172
<211> 46
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 172
ggatcccctg caggttattt tttcttgaca actccgttat taccgg 46

<210> 173
<211> 39
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 173
atatccgcgg ccgcacaatg gttatggagc aagctggaa 39

<210> 174
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 174
ggatcccctg caggtcaatg gagacaaggc tcgaaagt 38

<210> 175
<211> 42
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 175

ggatccgcgg cgcacaatg tccgccaaga tttcaatatt cc

42

<210> 176

<211> 38

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 176

ggatcccctg caggtttaatt tttcttaact actccatt

38

<210> 177

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 177

ggatccgcgg cgcacaatg ggagctcagg agaaacggcg cc

42

<210> 178

<211> 38

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 178

ggatcccctg caggtcacgt cttctccttc ttcaccgg

38

<210> 179

<211> 44

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 179

ggatccgcgg cgcacaatg gcggatcctg atctgtcttc tcct

44

<210> 180

<211> 44

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 180

ggatcccctg caggttatgt tggggccaag tcagggtgcaa agat

44

<210> 181

<211> 44

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 181
ggatccgcgg ccgcaaaatg gaaaaaaaga gtgtaccaa ttct 44

<210> 182
<211> 46
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 182
ggatcccctg caggttatgt gtttactaat ttgaggggaat tttttg 46

<210> 183
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 183
tcgacctgca ggaagcttaa ggatggatgat tgctgc 36

<210> 184
<211> 31
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 184
ggatccgcgg ccgcttactt ctccttctcc g 31

<210> 185
<211> 39
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 185
ggatccgcgg ccgcacaatg tcttttaggg atgtcctag 39

<210> 186
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 186
ggatcccctg cagggtcaatc atccttacct tttggtttac c 41

<210> 187
<211> 60
<212> DNA
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 187

atgtctttta gggatgtcct agaaagagga gatgaatttt ctgtgcggta ttccacaccg 60

<210> 188

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 188

tcaatcatcc ttaccctttg gtttaccctc tggaggcaga agattgtact gagagtgcac 60

<210> 189

<211> 44

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 189

ggatccgcgg cgcacaatg aagcattccc aaaaataccg tagg

44

<210> 190

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 190

ggatcccctg cagggtcaatg attttttttc atcacaaata c

41

<210> 191

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 191

atgaagcatt cccaaaaata ccgtaggtat ggaatttatg ctgtgcggta ttccacaccg 60

<210> 192

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 192

tcaatgattt tttttcatca caaatacaag aataagaaaa agattgtact gagagtgcac 60

<210> 193

<211> 43

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 193

ggatccgcgg ccgcacaatg ggttttggtg atttcttcga aac

43

<210> 194

<211> 45

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 194

ggatcccgctg caggttatctt ggtctcaatt ttaatatttt ttgac

45

<210> 195

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 195

atgggttttg ttgatttctt cgaaacatat atggtcgggt ctgtgcggta tttcacaccg 60

<210> 196

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 196

ttatttggtc tcaattttta ttttttttg caaggactcg agattgtact gagagtgcac 60

<210> 197

<211> 44

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 197

ggatccgcgg ccgcacaatg gaaaagtaca ccaattggag agac

44

<210> 198

<211> 42

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 198

ggatcccgctg caggctactt cctcttttta cgttgatcgc tg

42

<210> 199

<211> 60

<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 199
atggaaaagt acaccaattg gagagacaat ggtacgggaa ctgtgcggtta tttcacaccg 60

<210> 200
<211> 60
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 200
ctacttcctc tttttacgtt gatcgctgat atattccttc agattgtact gagagtgcac 60

<210> 201
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 201
ggatccgcgg ccgcacaatg cctgcaccaa aactcacgga g 41

<210> 202
<211> 38
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 202
ggatccctcg caggctacgc atctccttct ttcccttc 38

<210> 203
<211> 60
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 203
atgcctgcac caaaactcac ggagaaatct gcctcttcca ctgtgcggtta tttcacaccg 60

<210> 204
<211> 60
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 204
ctacgcattc cttcttttcc cttcttcttc ttcttctct agattgtact gagagtgcac 60

<210> 205
<211> 46
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 205
ggatccgcgg cgcacaatg tctgctcccg ctgccgatca taacgc

46

<210> 206
<211> 44
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 206
ggatccctcg caggtcattc tttcttttcg tgttctcttt tctg

44

<210> 207
<211> 60
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 207
atgtctgctc ccgctgccga tcataacgct gccaaacctc ctgtgctgta tttcacaccg 60

<210> 208
<211> 60
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 208
tcattctttc ttttcgtgtt ctcttttctg tcttaccagc agattgtact gagagtgcac 60

<210> 209
<211> 49
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 209
ggatccgcgg cgcacaatg ctgcatcaaa aaatagctca taaagttcg

49

<210> 210
<211> 49
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 210

ggatcccctg cagggtcaaaa aataaaacaa taaagtttat aaactaacc

49

<210> 211

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 211

atgctgcatc aaaaaatagc tcataaagtt cgaaaagtcg ctgtgcggtg tttcacaccg 60

<210> 212

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 212

tcaaaaaata aaacaataaa gtttataaac taaccaaatt agattgtact gagagtgcac 60

<210> 213

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 213

ggatccgcgg cgcgacaatg agtgtgatag gtaggttctt g

41

<210> 214

<211> 41

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 214

ggatcccctg cagggttaatg catctttttt acagatgaac c

41

<210> 215

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 215

atgagtgtga taggtaggtt cttgtattac ttgaggtccg ctgtgcggtg tttcacaccg 60

<210> 216

<211> 60

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:Synthetic
Oligonucleotide

<400> 216
 ttaatgcac ttttttacag atgaaccttc gttatgggta agattgtact gagagtgcac 60

<210> 217
 <211> 381
 <212> PRT
 <213> *Saccharomyces* sp.

<220>

<400> 217
 Met Ser Phe Arg Asp Val Leu Glu Arg Gly Asp Glu Phe Leu Glu Ala
 1 5 10 15
 Tyr Pro Arg Arg Ser Pro Leu Trp Arg Phe Leu Ser Tyr Ser Thr Ser
 20 25 30
 Leu Leu Thr Phe Gly Val Ser Lys Leu Leu Leu Phe Thr Cys Tyr Asn
 35 40 45
 Val Lys Leu Asn Gly Phe Glu Lys Leu Glu Thr Ala Leu Glu Arg Ser
 50 55 60
 Lys Arg Glu Asn Arg Gly Leu Met Thr Val Met Asn His Met Ser Met
 65 70 75 80
 Val Asp Asp Pro Leu Val Trp Ala Thr Leu Pro Tyr Lys Leu Phe Thr
 85 90
 Ser Leu Asp Asn Ile Arg Trp Ser Leu Gly Ala His Asn Ile Cys Phe
 100 105 110
 Gln Asn Lys Phe Leu Ala Asn Phe Phe Ser Leu Gly Gln Val Leu Ser
 115 120 125
 Thr Glu Arg Phe Gly Val Gly Pro Phe Gln Gly Ser Ile Asp Ala Ser
 130 135 140
 Ile Arg Leu Leu Ser Pro Asp Asp Thr Leu Asp Leu Glu Trp Thr Pro
 145 150 155 160
 His Ser Glu Val Ser Ser Ser Leu Lys Lys Ala Tyr Ser Pro Pro Ile
 165 170 175
 Ile Arg Ser Lys Pro Ser Trp Val His Val Tyr Pro Glu Gly Phe Val
 180 185 190
 Leu Gln Leu Tyr Pro Pro Phe Glu Asn Ser Met Arg Tyr Phe Lys Trp
 195 200 205
 Gly Ile Thr Arg Met Ile Leu Glu Ala Thr Lys Pro Pro Ile Val Val
 210 215 220
 Pro Ile Phe Ala Thr Gly Phe Glu Lys Ile Ala Ser Glu Ala Val Thr
 225 230 235 240
 Asp Ser Met Phe Arg Gln Ile Leu Pro Arg Asn Phe Gly Ser Glu Ile
 245 250 255
 Asn Val Thr Ile Gly Asp Pro Leu Asn Asp Asp Leu Ile Asp Arg Tyr
 260 265 270
 Arg Lys Glu Trp Thr His Leu Val Glu Lys Tyr Tyr Asp Pro Lys Asn
 275 280 285
 Pro Asn Asp Leu Ser Asp Glu Leu Lys Tyr Gly Lys Glu Ala Gln Asp
 290 295 300
 Leu Arg Ser Arg Leu Ala Ala Glu Leu Arg Ala His Val Ala Glu Ile

Ala Val Glu Lys Leu Ala Pro Ser Leu Asp Ala Ile Tyr Asp Val Thr
 260 265 270
 Ile Gly Tyr Ser Pro Ala Leu Arg Thr Glu Tyr Val Gly Thr Lys Phe
 275 280 285
 Thr Leu Lys Lys Ile Phe Leu Met Gly Val Tyr Pro Glu Lys Val Asp
 290 295 300
 Phe Tyr Ile Arg Glu Phe Arg Val Asn Glu Ile Pro Leu Gln Asp Asp
 305 310 315 320
 Glu Val Phe Phe Asn Trp Leu Leu Gly Val Trp Lys Glu Lys Asp Gln
 325 330 335
 Leu Leu Glu Asp Tyr Tyr Asn Thr Gly Gln Phe Lys Ser Asn Ala Lys
 340 345 350
 Asn Asp Asn Gln Ser Ile Val Val Thr Thr Gln Thr Thr Gly Phe Gln
 355 360 365
 His Glu Thr Leu Thr Pro Arg Ile Leu Ser Tyr Tyr Gly Phe Phe Ala
 370 375 380
 Phe Leu Ile Leu Val Phe Val Met Lys Lys Asn His
 385 390 395

<210> 219

<211> 479

<212> PRT

<213> Saccharomyces sp.

<220>

<400> 219

Met Gly Phe Val Asp Phe Phe Glu Thr Tyr Met Val Gly Ser Arg Val
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 Gln Phe Lys Gln Leu Asp Ile Ser Asp Trp Leu Ser Leu Thr Pro Arg
 20 25 30
 Leu Leu Ile Leu Phe Gly Tyr Phe Tyr Leu His Ser Phe Phe Thr Ala
 35 40 45
 Ile Asn Gln Phe Leu Gln Phe Ile Asn Thr Asn Ser Phe Cys Leu Arg
 50 55 60
 Leu His Leu Leu Tyr Asp Arg Phe Trp Ser His Val Pro Ile Ile Gly
 65 70 75 80
 Glu Tyr Lys Ile Arg Leu Leu Ser Arg Ala Leu Thr Tyr Ser Lys Leu
 85 90 95
 Lys Ile Ile Pro Thr Leu Asp Lys Val Leu Glu Ala Ile Glu Ile Trp
 100 105 110
 Phe Gln Leu His Leu Val Glu Met Thr Phe Glu Lys Lys Lys Asn Val
 115 120 125
 Gln Ile Phe Ile Thr Glu Gly Ser Asp Asp Leu Asn Phe Phe Lys Asp
 130 135 140
 Ser Lys Phe Gln Thr Thr Leu Met Ile Cys Asn His Arg Ser Val Asn
 145 150 155 160
 Asp Tyr Thr Leu Ile Asn Tyr Leu Phe Leu Lys Ser Cys Pro Thr Lys
 165 170 175

Phe Tyr Thr Lys Trp Glu Phe Leu Gln Lys Leu Arg Lys Gly Glu Asp
 180 185 190
 Leu Ala Glu Trp Pro Gln Leu Lys Phe Leu Gly Trp Gly Lys Met Phe
 195 200 205
 Asn Phe Pro Arg Leu Asp Leu Leu Lys Asn Ile Phe Phe Lys Asp Glu
 210 215 220
 Thr Leu Ala Leu Ser Ser Asn Glu Leu Arg Asp Ile Leu Glu Arg Gln
 225 230 235 240
 Asn Asn Gln Ala Ile Thr Ile Phe Pro Glu Val Asn Ile Met Ser Leu
 245 250 255
 Glu Leu Ser Ile Ile Gln Arg Lys Leu His Gln Asp Phe Pro Phe Val
 260 265 270
 Ile Asn Phe Tyr Asn Leu Leu Tyr Pro Arg Phe Lys Asn Phe Thr Thr
 275 280 285
 Leu Met Ala Ala Phe Ser Ser Ile Lys Asn Ile Lys Arg Lys Lys Asn
 290 295 300
 Arg Asn Asn Ile Ile Lys Glu Ala Arg Tyr Leu Phe His Arg Glu Leu
 305 310 315 320
 Asp Lys Leu Val His Lys Ser Met Lys Met Glu Ser Ser Lys Val Ser
 325 330 335
 Asp Lys Thr Thr Pro Pro Met Ile Val Asp Asn Ser Tyr Leu Leu Thr
 340 345 350
 Lys Lys Glu Glu Ile Ser Ser Gly Lys Pro Lys Val Val Arg Ile Asn
 355 360 365
 Pro Tyr Ile Tyr Asp Val Thr Ile Ile Tyr Tyr Arg Val Lys Tyr Thr
 370 375 380
 Asp Ser Gly His Asp His Thr Asn Gly Asp Leu Arg Leu His Lys Gly
 385 390 395 400
 Tyr Gln Leu Glu Gln Ile Ser Pro Thr Ile Phe Glu Met Ile Gln Pro
 405 410 415
 Glu Met Glu Ser Glu Asn Asn Ile Lys Asp Lys Asp Pro Ile Val Val
 420 425 430
 Met Val Asn Val Lys Lys His Gln Ile Gln Pro Leu Leu Ala Tyr Asn
 435 440 445
 Asp Glu Ser Leu Glu Lys Trp Leu Glu Asn Arg Trp Ile Glu Lys Asp
 450 455 460
 Arg Leu Ile Glu Ser Leu Gln Lys Asn Ile Lys Ile Glu Thr Lys
 465 470 475

<210> 220

<211> 300

<212> PRT

<213> Saccharomyces sp.

<400> 220

Met Glu Lys Tyr Thr Asn Trp Arg Asp Asn Gly Thr Gly Ile Ala Pro
 1 5 10 15

Phe Leu Pro Asn Thr Ile Arg Lys Pro Ser Lys Val Met Thr Ala Cys
 20 25 30

Leu Leu Gly Ile Leu Gly Val Lys Thr Ile Ile Met Leu Pro Leu Ile
 35 40 45
 Met Leu Tyr Leu Leu Thr Gly Gln Asn Asn Leu Leu Gly Leu Ile Leu
 50 55 60
 Lys Phe Thr Phe Ser Trp Lys Glu Glu Ile Thr Val Gln Gly Ile Lys
 65 70 75 80
 Lys Arg Asp Val Arg Lys Ser Lys His Tyr Pro Gln Lys Gly Lys Leu
 85 90 95
 Tyr Ile Cys Asn Cys Thr Ser Pro Leu Asp Ala Phe Ser Val Val Leu
 100 105 110
 Leu Ala Gln Gly Pro Val Thr Leu Leu Val Pro Ser Asn Asp Ile Val
 115 120 125
 Tyr Lys Val Ser Ile Arg Glu Phe Ile Asn Phe Ile Leu Ala Gly Gly
 130 135 140
 Leu Asp Ile Lys Leu Tyr Gly His Glu Val Ala Glu Leu Ser Gln Leu
 145 150 155 160
 Gly Asn Thr Val Asn Phe Met Phe Ala Glu Gly Thr Ser Cys Asn Gly
 165 170 175
 Lys Ser Val Leu Pro Phe Ser Ile Thr Gly Lys Lys Leu Lys Glu Phe
 180 185 190
 Ile Asp Pro Ser Ile Thr Thr Met Asn Pro Ala Met Ala Lys Thr Lys
 195 200 205
 Lys Phe Glu Leu Gln Thr Ile Gln Ile Lys Thr Asn Lys Thr Ala Ile
 210 215 220
 Thr Thr Leu Pro Ile Ser Asn Met Glu Tyr Leu Ser Arg Phe Leu Asn
 225 230 235 240
 Lys Gly Ile Asn Val Lys Cys Lys Ile Asn Glu Pro Gln Val Leu Ser
 245 250 255
 Asp Asn Leu Glu Glu Leu Arg Val Ala Leu Asn Gly Gly Asp Lys Tyr
 260 265 270
 Lys Leu Val Ser Arg Lys Leu Asp Val Glu Ser Lys Arg Asn Phe Val
 275 280 285
 Lys Glu Tyr Ile Ser Asp Gln Arg Lys Lys Arg Lys
 290 295 300

<210> 221

<211> 759

<212> PRT

<213> Saccharomyces sp.

<400> 221

Met Pro Ala Pro Lys Leu Thr Glu Lys Phe Ala Ser Ser Lys Ser Thr
 1 5 10 15
 Gln Lys Thr Thr Asn Tyr Ser Ser Ile Glu Ala Lys Ser Val Lys Thr
 20 25 30
 Ser Ala Asp Gln Ala Tyr Ile Tyr Gln Glu Pro Ser Ala Thr Lys Lys
 35 40 45
 Ile Leu Tyr Ser Ile Ala Thr Trp Leu Leu Tyr Asn Ile Phe His Cys
 50 55 60

Phe Phe Arg Glu Ile Arg Gly Arg Gly Ser Phe Lys Val Pro Gln Gln
 65 70 75 80
 Gly Pro Val Ile Phe Val Ala Ala Pro His Ala Asn Gln Phe Val Asp
 85 90 95
 Pro Val Ile Leu Met Gly Glu Val Lys Lys Ser Val Asn Arg Arg Val
 100 105 110
 Ser Phe Leu Ile Ala Glu Ser Ser Leu Lys Gln Pro Pro Ile Gly Phe
 115 120 125
 Leu Ala Ser Phe Phe Met Ala Ile Gly Val Val Arg Pro Gln Asp Asn
 130 135 140
 Leu Lys Pro Ala Glu Gly Thr Ile Arg Val Asp Pro Thr Asp Tyr Lys
 145 150 155 160
 Arg Val Ile Gly His Asp Thr His Phe Leu Thr Asp Cys Met Pro Lys
 165 170 175
 Gly Leu Ile Gly Leu Pro Lys Ser Met Gly Phe Gly Glu Ile Gln Ser
 180 185 190
 Ile Glu Ser Asp Thr Ser Leu Thr Leu Arg Lys Glu Phe Lys Met Ala
 195 200 205
 Lys Pro Glu Ile Lys Thr Ala Leu Leu Thr Gly Thr Thr Tyr Lys Tyr
 210 215 220
 Ala Ala Lys Val Asp Gln Ser Cys Val Tyr His Arg Val Phe Glu His
 225 230 235 240
 Leu Ala His Asn Asn Cys Ile Gly Ile Phe Pro Glu Gly Gly Ser His
 245 250 255
 Asp Arg Thr Asn Leu Leu Pro Leu Lys Ala Gly Val Ala Ile Met Ala
 260 265 270
 Leu Gly Cys Met Asp Lys His Pro Asp Val Asn Val Lys Ile Val Pro
 275 280 285
 Cys Gly Met Asn Tyr Phe His Pro His Lys Phe Arg Ser Arg Ala Val
 290 295 300
 Val Glu Phe Gly Asp Pro Ile Glu Ile Pro Lys Glu Leu Val Ala Lys
 305 310 315 320
 Tyr His Asn Pro Glu Thr Asn Arg Asp Ala Val Lys Glu Leu Leu Asp
 325 330 335
 Thr Ile Ser Lys Gly Leu Gln Ser Val Thr Val Thr Cys Ser Asp Tyr
 340 345 350
 Glu Thr Leu Met Val Val Gln Thr Ile Arg Arg Leu Tyr Met Thr Gln
 355 360 365
 Phe Ser Thr Lys Leu Pro Leu Pro Leu Ile Val Glu Met Asn Arg Arg
 370 375 380
 Met Val Lys Gly Tyr Glu Phe Tyr Arg Asn Asp Pro Lys Ile Ala Asp
 385 390 395 400
 Leu Thr Lys Asp Ile Met Ala Tyr Asn Ala Ala Leu Arg His Tyr Asn
 405 410 415
 Leu Pro Asp His Leu Val Glu Glu Ala Lys Val Asn Phe Ala Lys Asn
 420 425 430

Leu Gly Leu Val Phe Phe Arg Ser Ile Gly Leu Cys Ile Leu Phe Ser
 435 440 445
 Leu Ala Met Pro Gly Ile Ile Met Phe Ser Pro Val Phe Ile Leu Ala
 450 455 460
 Lys Arg Ile Ser Gln Glu Lys Ala Arg Thr Ala Leu Ser Lys Ser Thr
 465 470 475 480
 Val Lys Ile Lys Ala Asn Asp Val Ile Ala Thr Trp Lys Ile Leu Ile
 485 490 495
 Gly Met Gly Phe Ala Pro Leu Leu Tyr Ile Phe Trp Ser Val Leu Ile
 500 505 510
 Thr Tyr Tyr Leu Arg His Lys Pro Trp Asn Lys Ile Tyr Val Phe Ser
 515 520 525
 Gly Ser Tyr Ile Ser Cys Val Ile Val Thr Tyr Ser Ala Leu Ile Val
 530 535 540
 Gly Asp Ile Gly Met Asp Gly Phe Lys Ser Leu Arg Pro Leu Val Leu
 545 550 555 560
 Ser Leu Thr Ser Pro Lys Gly Leu Gln Lys Leu Gln Lys Asp Arg Arg
 565 570 575
 Asn Leu Ala Glu Arg Ile Ile Glu Val Val Asn Asn Phe Gly Ser Glu
 580 585 590
 Leu Phe Pro Asp Phe Asp Ser Ala Ala Leu Arg Glu Glu Phe Asp Val
 595 600 605
 Ile Asp Glu Glu Glu Glu Asp Arg Lys Thr Ser Glu Leu Asn Arg Arg
 610 615 620
 Lys Met Leu Arg Lys Gln Lys Ile Lys Arg Gln Glu Lys Asp Ser Ser
 625 630 635 640
 Ser Pro Ile Ile Ser Gln Arg Asp Asn His Asp Ala Tyr Glu His His
 645 650 655
 Asn Gln Asp Ser Asp Gly Val Ser Leu Val Asn Ser Asp Asn Ser Leu
 660 665 670
 Ser Asn Ile Pro Leu Phe Ser Ser Thr Phe His Arg Lys Ser Glu Ser
 675 680 685
 Ser Leu Ala Ser Thr Ser Val Ala Pro Ser Ser Ser Glu Phe Glu
 690 695 700
 Val Glu Asn Glu Ile Leu Glu Glu Lys Asn Gly Leu Ala Ser Lys Ile
 705 710 715 720
 Ala Gln Ala Val Leu Asn Lys Arg Ile Gly Glu Asn Thr Ala Arg Glu
 725 730 735
 Glu Glu Glu Glu Glu Glu Glu Glu Glu Glu Glu Glu Glu Glu Glu
 740 745 750
 Glu Gly Lys Glu Gly Asp Ala
 755

<210> 222

<211> 743

<212> PRT

<213> Saccharomyces sp.

<400> 222

Met Ser Ala Pro Ala Ala Asp His Asn Ala Ala Lys Pro Ile Pro His
 1 5 10 15
 Val Pro Gln Ala Ser Arg Arg Tyr Lys Asn Ser Tyr Asn Gly Phe Val
 20 25 30
 Tyr Asn Ile His Thr Trp Leu Tyr Asp Val Ser Val Phe Leu Phe Asn
 35 40 45
 Ile Leu Phe Thr Ile Phe Phe Arg Glu Ile Lys Val Arg Gly Ala Tyr
 50 55 60
 Asn Val Pro Glu Val Gly Val Pro Thr Ile Leu Val Cys Ala Pro His
 65 70 75 80
 Ala Asn Gln Phe Ile Asp Pro Ala Leu Val Met Ser Gln Thr Arg Leu
 85 90 95
 Leu Lys Thr Ser Ala Gly Lys Ser Arg Ser Arg Met Pro Cys Phe Val
 100 105 110
 Thr Ala Glu Ser Ser Phe Lys Lys Arg Phe Ile Ser Phe Phe Gly His
 115 120 125
 Ala Met Gly Gly Ile Pro Val Pro Arg Ile Gln Asp Asn Leu Lys Pro
 130 135 140
 Val Asp Glu Asn Leu Glu Ile Tyr Ala Pro Asp Leu Lys Asn His Pro
 145 150 155 160
 Glu Ile Ile Lys Gly Arg Ser Lys Asn Pro Gln Thr Thr Pro Val Asn
 165 170 175
 Phe Thr Lys Arg Phe Ser Ala Lys Ser Leu Leu Gly Leu Pro Asp Tyr
 180 185 190
 Leu Ser Asn Ala Gln Ile Lys Glu Ile Pro Asp Asp Glu Thr Ile Ile
 195 200 205
 Leu Ser Ser Pro Phe Arg Thr Ser Lys Ser Lys Val Val Glu Leu Leu
 210 215 220
 Thr Asn Gly Thr Asn Phe Lys Tyr Ala Glu Lys Ile Asp Asn Thr Glu
 225 230 235 240
 Thr Phe Gln Ser Val Phe Asp His Leu His Thr Lys Gly Cys Val Gly
 245 250 255
 Ile Phe Pro Glu Gly Gly Ser His Asp Arg Pro Ser Leu Leu Pro Ile
 260 265 270
 Lys Ala Gly Val Ala Ile Met Ala Leu Gly Ala Val Ala Ala Asp Pro
 275 280 285
 Thr Met Lys Val Ala Val Val Pro Cys Gly Leu His Tyr Phe His Arg
 290 295 300
 Asn Lys Phe Arg Ser Arg Ala Val Leu Glu Tyr Gly Glu Pro Ile Val
 305 310 315 320
 Val Asp Gly Lys Tyr Gly Glu Met Tyr Lys Asp Ser Pro Arg Glu Thr
 325 330 335
 Val Ser Lys Leu Leu Lys Lys Ile Thr Asn Ser Leu Phe Ser Val Thr
 340 345 350
 Glu Asn Ala Pro Asp Tyr Asp Thr Leu Met Val Ile Gln Ala Ala Arg
 355 360 365
 Arg Leu Tyr Gln Pro Val Lys Val Arg Leu Pro Leu Pro Ala Ile Val

<210> 223

<211> 397

<212> PRT

<213> Saccharomyces sp.

<400> 223

Met Leu His Gln Lys Ile Ala His Lys Val Arg Lys Val Val Val Pro
1 5 10 15Gly Ile Ser Leu Leu Ile Phe Phe Gln Gly Cys Leu Ile Leu Leu Phe
20 25 30Leu Gln Leu Thr Tyr Lys Thr Leu Tyr Cys Arg Asn Asp Ile Arg Lys
35 40 45Gln Ile Gly Leu Asn Lys Thr Lys Arg Leu Phe Ile Val Leu Val Ser
50 55 60Ser Ile Leu His Val Val Ala Pro Ser Ala Val Arg Ile Thr Thr Glu
65 70 75 80Asn Ser Ser Val Pro Lys Gly Thr Phe Phe Leu Asp Leu Lys Lys Lys
85 90 95Arg Ile Leu Ser His Leu Lys Ser Asn Ser Val Ala Ile Cys Asn His
100 105 110Gln Ile Tyr Thr Asp Trp Ile Phe Leu Trp Trp Leu Ala Tyr Thr Ser
115 120 125Asn Leu Gly Ala Asn Val Phe Ile Ile Leu Lys Lys Ser Leu Ala Ser
130 135 140Ile Pro Ile Leu Gly Phe Gly Met Arg Asn Tyr Asn Phe Ile Phe Met
145 150 155 160Ser Arg Lys Trp Ala Gln Asp Lys Ile Thr Leu Ser Asn Ser Leu Ala
165 170 175Gly Leu Asp Ser Asn Ala Arg Gly Ala Gly Ser Leu Ala Gly Lys Ser
180 185 190Pro Glu Arg Ile Thr Glu Glu Gly Glu Ser Ile Trp Asn Pro Glu Val
195 200 205Ile Asp Pro Lys Gln Ile His Trp Pro Tyr Asn Leu Ile Leu Phe Pro
210 215 220Glu Gly Thr Asn Leu Ser Ala Asp Thr Arg Gln Lys Ser Ala Lys Tyr
225 230 235 240Ala Ala Lys Ile Gly Lys Lys Pro Phe Lys Asn Val Leu Leu Pro His
245 250 255Ser Thr Gly Leu Arg Tyr Ser Leu Gln Lys Leu Lys Pro Ser Ile Glu
260 265 270Ser Leu Tyr Asp Ile Thr Ile Gly Tyr Ser Gly Val Lys Gln Glu Glu
275 280 285Tyr Gly Glu Leu Ile Tyr Gly Leu Lys Ser Ile Phe Leu Glu Gly Lys
290 295 300Tyr Pro Lys Leu Val Asp Ile His Ile Arg Ala Phe Asp Val Lys Asp
305 310 315 320Ile Pro Leu Glu Asp Glu Asn Glu Phe Ser Glu Trp Leu Tyr Lys Ile
325 330 335

Trp Ser Glu Lys Asp Ala Leu Met Glu Arg Tyr Tyr Ser Thr Gly Ser
 340 345 350
 Phe Val Ser Asp Pro Glu Thr Asn His Ser Val Thr Asp Ser Phe Lys
 355 360 365
 Ile Asn Arg Ile Glu Leu Thr Glu Val Leu Ile Leu Pro Thr Leu Thr
 370 375 380
 Ile Ile Trp Leu Val Tyr Lys Leu Tyr Cys Phe Ile Phe
 385 390 395

<210> 224
 <211> 303
 <212> PRT
 <213> Saccharomyces sp.

<400> 224
 Met Ser Val Ile Gly Arg Phe Leu Tyr Tyr Leu Arg Ser Val Leu Val
 1 5 10 15
 Val Leu Ala Leu Ala Gly Cys Gly Phe Tyr Gly Val Ile Ala Ser Ile
 20 25 30
 Leu Cys Thr Leu Ile Gly Lys Gln His Leu Ala Gln Trp Ile Thr Ala
 35 40 45
 Arg Cys Phe Tyr His Val Met Lys Leu Met Leu Gly Leu Asp Val Lys
 50 55 60
 Val Val Gly Glu Glu Asn Leu Ala Lys Lys Pro Tyr Ile Met Ile Ala
 65 70 75 80
 Asn His Gln Ser Thr Leu Asp Ile Phe Met Leu Gly Arg Ile Phe Pro
 85 90 95
 Pro Gly Cys Thr Val Thr Ala Lys Lys Ser Leu Lys Tyr Val Pro Phe
 100 105 110
 Leu Gly Trp Phe Met Ala Leu Ser Gly Thr Tyr Phe Leu Asp Arg Ser
 115 120 125
 Lys Arg Gln Glu Ala Ile Asp Thr Leu Asn Lys Gly Leu Glu Asn Val
 130 135 140
 Lys Lys Asn Lys Arg Ala Leu Trp Val Phe Pro Glu Gly Thr Arg Ser
 145 150 155 160
 Tyr Thr Ser Glu Leu Thr Met Leu Pro Phe Lys Lys Gly Ala Phe His
 165 170 175
 Leu Ala Gln Gln Gly Lys Ile Pro Ile Val Pro Val Val Val Ser Asn
 180 185 190
 Thr Ser Thr Leu Val Ser Pro Lys Tyr Gly Val Phe Asn Arg Gly Cys
 195 200 205
 Met Ile Val Arg Ile Leu Lys Pro Ile Ser Thr Glu Asn Leu Thr Lys
 210 215 220
 Asp Lys Ile Gly Glu Phe Ala Glu Lys Val Arg Asp Gln Met Val Asp
 225 230 235 240
 Thr Leu Lys Glu Ile Gly Tyr Ser Pro Ala Ile Asn Asp Thr Thr Leu
 245 250 255
 Pro Pro Gln Ala Ile Glu Tyr Ala Ala Leu Gln His Asp Lys Lys Val
 260 265 270

Asn Lys Lys Ile Lys Asn Glu Pro Val Pro Ser Val Ser Ile Ser Asn
 275 280 285
 Asp Val Asn Thr His Asn Glu Gly Ser Ser Val Lys Lys Met His
 290 295 300

<210> 225
 <211> 1146
 <212> DNA
 <213> *Saccharomyces* sp.

<400> 225
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 agcccccttt ggagatttct ttcatacagt acatcattac tgaccttcgg tgtatcaaaa 120
 ctgcttcttt tcacatgcta taatgtcaaa ttgaatgggt ttgaaaaatt agaaactgcc 180
 ttggaacggt ccaaaaggga aaatagaggc cttatgacgg tcatgaacca tatgagtatg 240
 gtcgatgatc cgttagtttg ggcaacacta ccatataagt tatttacgtc tttggacaac 300
 ataagatggt ctttgggtgc acataatatt tgctttcaaa ataaatttct ggccaacttt 360
 ttctcacttg gccaaagtcct ttcaacagaa agatttgggg tgggcccatt tcaaggttct 420
 atagatgctt caataagatt gttaagccct gacgacact tagacttgga atggaccct 480
 cactctgagg tctcttcttc gctaaaaaaa gcctactccc cgcccataat aagggtcgaag 540
 ccattctggg tccatgttta tccagaagga tttgtactac aattatatcc gccttttgaa 600
 aattcgatga ggtatttttaa atggggtatt accagaatga tcctagaagc aacaaagccg 660
 cccattgtag tactaatatt tgctacaggg tttgaaaaaa tagcatccga agcagtcaca 720
 gattcaatgt ttagacaaat tctaccaaga aactttggct ctgaaataaa tgttaccata 780
 ggggatcctt taaatgatga tttaatcgac aggtatagaa aagaatggac acatttgggt 840
 gaaaaatact atgatcccaa aaatcctaac gacctctctg acgaattgaa atatggtaaa 900
 gaggcgcaag atttaagaag cagattagcc gctgaactga gagcccatgt tgctgaaatt 960
 agaaatgaag ttcgcaaatt accacgcgaa gaccctaggt tcaaatcccc ctcatggtgg
 1020
 aagcgggttca acaccacgga aggtaaatcg gaccagatg ttaaagtcatt tggcgaaaat
 1080
 tgggcaataa ggaggatgca aaagtctctg cctccagagg gtaaaccaaa gggtaaggat
 1140
 gattga
 1146

<210> 226
 <211> 1191
 <212> DNA
 <213> *Saccharomyces* sp.

<400> 226
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 ataaaagggt tgcaaaggct gcttatcgct tgcttgttca tttcaggctc gctgagtatt 120
 gtcgtttttc agatctgtct acagggtgctt ctcccttgga gcaagattag atttcaaat 180
 ggtataaatc aaagtaagaa ggctttttatc gttttattat gcatgatctt gaacatgggtg 240
 gctccctctt ctttgaatgt cacttttgaa acatcgcggc cattgaagaa ctcttctaac 300
 gccaaagccat gcttttagatt taaagacagg gctataataa ttgcaaatca tcaaatgtat 360
 gcagactgga tttatctctg gtggctttcc tttgtttcaa atttgggtgg taacgtttat 420
 atcatcctga agaaagctct gcagtaacata ccattactgg gatttggcat gcgaaatttt 480
 aagttttatat ttttaagtag gaactggcaa aaggatgaga aagctttaac aaatagtttg 540
 gtttctatgg acttaaacgc gaggtgcaag gggcccccta caaattataa gagttgttat 600
 tccaagacaa atgaatccat tgccgcttat aatttaacata tgttccctga gggtaacaaat 660
 ctaagcctca agacaagaga aaaaagcgag gcattctgtc aaagagcaca tttggaccat 720
 gtccaattaa gacatttgtt attaccgcac tctaaaggct tgaagtttgc agtagaaaaa 780
 cttagctccta gtttagatgc tatctacgat gtcactattg gatattctcc cgccttgaga 840
 acggaatagc tcggcaccaa attcaccttg aagaaaaat tcttaattgg tgctatccg 900
 gagaaagtag atttttatat tagggaattt agagttaatg agatcccttt gcaagatgac 960
 gaagtttttt tcaattgggt actgggcgtg tggaaagaaa aagatcaact gctagaagac
 1020
 tactacaaca caggccaatt taaaagtaat gctaaaaatg acaaccaatc catcgttggt
 1080
 acgacacaaa cgactggatt tcagcacgaa acattgacac cccgtatcct ttcattattac
 1140
 gggttcttcg cttttcttat tcttgtattt gtgatgaaaa aaaatcattg a
 1191

<210> 227
<211> 1440
<212> DNA
<213> *Saccharomyces* sp.

<400> 227
atggggttttg ttgatttctt cgaaacatat atgggtcggtt ctagggtcca gttcaaacag 60
ttagatatatt ctgattgggt gagtctgacc ccaagggtgc ttattctttt tggctatttt 120
taccttcatt ctttttttac tgcaatcaat caattcctac agttcattaa cacgaattcc 180
ttctgtctta gactgcattt actatatgac agattttgggt cgcatgtgcc cataataggt 240
gagtacaaaa ttcggtgct ctcgagggca ctgacatata gtaaaactgaa aataatacca 300
actttagaca aggtgctgga ggcgattgaa atttggtttc agctacattt agttgaaatg 360
accttcgaaa aaaaaaaaaa cgtccaaatt ttcataaccg agggaagtga tgacctaaac 420
ttttttaaag atagcaaatt ccaaaccaca ttaatgatat gtaatcatcg atcagtgaat 480
gactacacat tgattaatta cctttttctc aaaagtgtgc ccaccaagtt ttatactaaa 540
tggaattttc tacaaaagct gaggaagggg gaagatctag ctgaatggcc tcagttaaaa 600
tttcttggtt ggggaaaaat gtttaacttt cctcgattgg atctactaaa gaacatatct 660
ttcaaagatg aaacactcgc actctcatcg aatgagttaa gagatatatt agaaagacaa 720
aacaatcaat ctattactat ttttcccgaa gtcaatatca tgagtttgga actatcaatt 780
attcaaagaa aattacacca agattttccc ttgtttataa acttctataa tttattatac 840
ccaagattta aaaactttac cactttgatg gctgcttttt catcaattaa aaacatcaaa 900
agaaagaaaa accgtaacaa tataatcaaa gaggccgat acctgtttca cagagaactt 960
gacaaattag ttcacaagag catgaaaatg gagtcttcca aggtatccga taagacgacg
1020
ccgcccata tctgtagataa ttcatactta cttacaaaaa aggaagaaat cagcagcggc
1080
aagcccaagg tggtagaat caatccatac atatatgatg tcaccataat ttattaccga
1140
gtcaaatata ctgatagtgg gcatgatcat accaacggag atttgagact tcataaagggt
1200
tatcaattag agcaaatatc tccgacaatc tttagatga ttcaaccaga aatggagtct
1260
gaaaacaaca taaaggataa ggacccatt gttgtgatgg taaatgtaaa aaagcatcaa
1320
attcaaccat tactcgcata caatgatgag agtttagaaa agtggcttga aaataggtgg
1380
atagaaaaag atagattaat cgagtccttg caaaaaata ttaaaattga gaccaaataa
1440

<210> 228
<211> 903
<212> DNA
<213> *Saccharomyces* sp.

<400> 228
atggaaaagt acaccaattg gagagacaat ggtacgggaa tagctccatt tctaccaaac 60
acaatcagga aacctagtaa ggtgatgaca gcgtgtttgt tgggtatcct aggggtgaaa 120
accattataa tgctaccatt gattatgctg taccttctaa ctggccagaa caacttactg 180
ggtttgatat tgaagtttac attcagttgg aaagaggaaa ttaccgtgca aggaatcaag 240
aaacgtgcag taaggaaatc caagcattat ccacagaagg gcaagcttta tatttgcaat 300
tgtacctcac ctttagatgc tttttcagtg gtgttattag ctcaagggcc tgttacgttg 360
ttgggtccat ccaatgacat tgtatacaaa gtttccataa gagaattcat caacttcac 420
ctcgccgggtg ggtagatat aaaactctat ggccacgagg tagcagagct atctcaattg 480
ggcaataccg tgaattttat gtttgcagag ggtacctcat gtaatggtaa aagcgtctta 540
ccgttttagta taaccgggaa aaaacttaaa gaattcatag acccttcaat aaccacaatg 600
aaccgccgaa tggccaaaac taaaaaattt gaattgcaga ccatccaaat caaaactaat 660
aaaactgcca tcaccacatt gccatctccc aatatggagt atttatctag atttctgaac 720
aagggcatta atgttaaag caagatcaac gagccacaag tactctcgga taatttagag 780
gaattacgag ttgcattaaa cgggtggcag aaatataaac tagtctcacg gaagttagat 840
gttgaatcta agaggaattt tgtgaaggaa tatatcagcg atcaacgtaa aaagaggaag 900
tag 903

<210> 229
<211> 2280
<212> DNA
<213> *Saccharomyces* sp.

<400> 229
atgcctgcac caaaactcac ggagaaattt gcctcttcca agagcacaca gaaaactacg 60
aattacagtt ccatcgaggc caaaagcgtc aagacgtcgg ctgatcaggc atacatctac 120

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caagagccta ggcgtaccaa gaagatactt tactccatcg ccacatggct gttgtacaac 180
atcttccact gcttcttttag agaaatcaga ggccggggca gtttcaagggt accgcaacag 240
ggaccgggtga tctttgttgc ggctccgcat gctaaccagt tcgtcgaccc tgtaatcctt 300
atgggcgagg tgaagaaatc tgtcaacaga cgtgtgtcct tcttgattgc ggagagctca 360
ttaagcaac ccccatagg gtttttggct agtttcttca tggccatagg cgtggtaagg 420
ccgcaggata atttgaaacc ggcagaagggt actatccgcg tagatccaac agactacaag 480
agagttatcg gccacgacac gcatttcttg actgattgta tgccaaagggt tctcatcggg 540
ttacccaaat caatgggatt tggagaaatc cagtccatag aaagtgcac gagtttgacc 600
ctaagaaaag agttcaaaat ggccaaacca gagattaaaa ctgctttact caccggcact 660
acttataaat atgccgctaa agtcgaccaa tcttgcgctt accatagagt ttttgagcat 720
ttggcccata acaactgcat tgggatcttt cctgaagggt ggtcccacga cagaacaaac 780
ttgttgcgcc tgaagcagg ttgtgctctg gttgcatgga taagatcctt 840
gacgtcaatg ttaagattgt tccctgcggg atgaattatt tccatccaca taagttcagg 900
tcgagagcgg ttgttgaatt cggtgacccc attgaaatac cgaaggaact agtcgccaaag 960
taccacaacc cggaaacgaa cagagatgca gtgaaagaat tattagatac catatcgaag
1020
ggtttacaat cegttaccgt tacatgttct gattatgaaa ctttgatgggt ggttcaaacg
1080
ataagaagac tatatatgac acaatttagc accaagttac cgttgccctt gattgtggaa
1140
atgaacagaa gaatgggtcaa aggttacgaa ttctatagaa acgatcctaa aatagcggac
1200
ttgaccaaaag atataatggc atataatgcc gccttgagac actataatct tcctgatcac
1260
cttgtggagg aggcaaagggt aaatttcgca aaaaacctcg gacttgtttt ttttagatcc
1320
atcgggctct gcacccctctt ttcggttagcc atgccaggta tcattatggt ctcacctgtc
1380
ttcatattag ccaagagaat ttctcaagaa aaggcccgtc cgcctttgtc caagtctaca
1440
gttaaaataa aggctaacga tgtcattgcc acgtggaaaa tcttgattgg gatgggattt
1500
gcgcccttgc tttacatctt ttgggtccgtt ttaatcactt attacctcag acataaacca
1560
tggaataaaa tatatgtttt ttccgggtct tacatctcgt gtgttatagt cacgtattcc
1620
gccttaatcg tgggtgatat tgggtatggat ggtttcaaat ctttgagacc actgggtttta
1680
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1740
agaataatcg aagttgtaaa taactttgga agcgaattat tccccgattt cgatagtggc
1800
gccctacgtg aagaattcga cgtcatcgat gaagaggaag aagatcgaag aacctcagaa
1860
ttgaatcgca ggaaaatgct aagaaaacag aaaataaaaa gacaagaaaa agattcgtca
1920
tcacctatca tcagccaacg tgacaaccac gatgcctatg aacaccataa ccaagattcc
1980
gatggcgtct cattgggtcaa tagtgacaat tccctctcta acattccatt attctcttct
2040
acttttcac gtaagtcaga gtcttcctta gcttcgacat ccgttgaccc ttcttcttcc
2100
tccgaatttg aggtagaaaa cgaaatcttg gaggaaaaaa atggattagc aagtaaaatc
2160
gcacaggccg tcttaaacia gagaattggg gaaaatactg ccagggaaga ggaagaggaa
2220
gaagaagagg aagaagaaga agaggaagaa gaagaagaag ggaaagaagg agatgcgtag
2280

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<210> 230

<211> 2232

<212> DNA

<213> *Saccharomyces* sp.

<400> 230

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atgtctgctc ccgctgccga tcataacgct gccaaaccta ttcctcatgt acctcaagcg 60
tcccgcggtg acaaaaattc atacaatgga ttcgatataca atatacatatc atggctgtat 120
gatgtgtctg tatttctgtt taatatcttg ttactatttt tcttcagaga aattaaggta 180
cgtgggtgcat ataacgttcc cgaagttggg gtgccaaacca tccttggtgtg tgccccctcat 240
gcaaatcagt tcacgcaccc ggctttggta atgtcgcaaa cccgtttgct gaagacatca 300

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gcgggaaagt cccgatccag aatgccttgt tttgttactg ctgagtcgag ttttaagaaa 360
agatttatct ctttcttttg tcacgcaatg ggcggatttc ccgtgcctag aattcaggac 420
aacttgaagc cagtggatga gaatcttgag atttacgctc cggacttgaa gaaccacccg 480
gaaatcatca agggccgctc caagaaccca cagactacac cagtgaactt tacgaaaagg 540
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2100
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<210> 231

<211> 1194

<212> DNA

<213> *Saccharomyces* sp.

<400> 231

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<210> 232

<211> 912

<212> DNA

<213> Saccharomyces sp.

<400> 232

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<213> Artificial Sequence

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Oligonucleotide

<400> 233

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<210> 234

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<212> DNA

<213> Artificial Sequence

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Oligonucleotide

<400> 234

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Oligonucleotide

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Oligonucleotide

<400> 241

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28

